



First records of the genus *Aristosyrphus* Curran, 1941 (Diptera, Syrphidae) from Ecuador

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Abstract

We record *Aristosyrphus carpenteri* (Hull, 1945) for the first time from South America. This species was previously known from Panama and Costa Rica. New specimens, collected in Pichincha Province of Ecuador, represent the first reported occurrence of this flower fly genus from the country. Images, diagnosis, and DNA barcodes are provided to help with the identification of this species.

Keywords

Ant flies, *Aristosyrphus carpenteri*, flower flies, hoverflies, Microdontinae, new record

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Introduction

Aristosyrphus Curran, 1941 (Diptera, Syrphidae) is a small genus of flower flies found in Central and South America (Cheng and Thompson 2008; Reemer and Ståhls 2013a). The biology of *Aristosyrphus* is unknown, but it is presumed that larvae are myrmecophilous like most microdontines (Reemer 2013).

Aristosyrphus belongs to the subfamily Microdontinae, also known as ant flies (Thompson 2020), and it is divided into two distinct subgenera: *Aristosyrphus sensu stricto* with four described species and *Aristosyrphus (Eurypterosyrphus)* with three species, although there are several taxa waiting for a formal description (Reemer and Ståhls 2013a). Five out of the seven described species occur in Brazil, namely *A. (A.) minutus* Thompson in Marinoni & Thompson, 2004, *A. (A.) primus* Curran,

1941, *A. (A.) boraceiensis* (Papavero, 1962), *A. (E.) macropterus* (Curran, 1941), and *A. (E.) melanopterus* (Barretto & Lane, 1947). *Aristosyrphus (E.) currani* (van der Goot, 1964), a new name for *Microdon clavicornis* Curran, 1940, is only known from the female holotype collected in Guyana. One species, *A. (A.) carpenteri* (Hull, 1945), was originally described from Panama (Hull 1945), but it has also been reported from Costa Rica (GBIF 2022).

The Neotropical Region is worldwide one of the least sampled areas for animals (Hughes et al. 2021), and insects are usually an understudied animal group (Troudet et al. 2017). Consequently, it is not surprising that in a megadiverse country like Ecuador (Mittermeier et al. 2005), a species-rich family like Syrphidae

is underexplored and poorly collected (Amorim 2009). Marín-Armijos et al. (2017) reviewed the literature records of Ecuadorian syrphids and listed 201 species belonging to 51 genera and subgenera. They concluded that the syrphid fauna in Ecuador is far from being completely known. The project “Diversidad de moscas florícolas (Insecta: Diptera) del Ecuador” (MAAE-DBI-CM-2021-0167) shared between the Zoological Research Museum Koenig (Bonn, Germany) and the Instituto Nacional de Biodiversidad (Quito, Ecuador) aims to fill the knowledge gap for flies in this country. With the help of DNA barcodes (Hebert et al. 2003) and morphological identifications, the project intends to create a species list of dipterans that will help to monitor this group by providing identification tools. As a result of the ongoing efforts, in the present work, we report for the first time the genus *Aristosyrphus* from Ecuador.

Methods

Fieldwork was carried out between December 2019 and January 2020 in the Parroquia Pedro Vicente Maldonado, Pichincha Province, Ecuador. Sampling was performed using a double Malaise trap. This trap consists of two single Townes Malaise traps (Townes 1972) sewn back to back. The double malaise traps were placed in the transition zone between a lightly managed organic farming area and an old secondary forest. Behind the secondary forest is a primary forest with a stream, and pastures are predominant in the surrounding area (more than 100 m away from the trap in any direction). The sampling was done on the northwestern slopes of the Andes between the evergreen forest of the equatorial Choco lowlands and Piedmont evergreen forest of the western Andean Range (Noh et al. 2020; Ron 2020).

Collecting jars were filled with 96% ethyl alcohol and specimens were transferred to a vial with new 96% ethyl alcohol once they arrived at the lab and kept in a freezer at -20°C . Specimens are deposited in Instituto Nacional de Biodiversidad (INABIO) and Zoological Research Museum Koenig (ZFMK-DIP).

Specimens of the subfamily Microdontinae were identified to genus using the key provided by Reemer and Ståhls (2013a) and to species using a manuscript identification key (Thompson unpublished, in Primer taller de identificación de Syrphidae del Neotrópico, 21 a 27 de febrero de 2006, Facultad de Ciencias, Universidad del Valle, Cali, Colombia). The adult terminology used follows Cumming and Wood (2017), except the terms for male genitalia that follow Reemer and Ståhls (2013a).

Specimens were dried using the Leica EM CPD300 Automated Critical Point Dryer. Pinned specimens were photographed using a Canon EOS 7D[®] mounted on a P-51 Cam-Lift (Dun Inc., Virginia, USA) and stacked using the software Zerene Stacker[®] v. 1.04 (Richland, Washington, USA) with the help of Adobe Lightroom[®] v. 5.6 to export the images. SimpleMappr (Shorthouse

2010) was used to create Figure 1. Google Earth[®] was used to obtain the geographic coordinates of the holotype of *A. carpenteri*, and we also included coordinates from publicly available records (GBIF 2021; BOLD, <https://www.boldsystems.org>) (see Appendix Table A1).

For the selected specimens, the 5'-end of the mitochondrial cytochrome oxidase c subunit I (COI) gene was sequenced. One leg from the selected specimens was used for DNA extraction. DNA was extracted following standard protocols of the commercially available DNeasy Blood & Tissue Kit (QIAGEN[®]). The COI barcode region was amplified using the forward primer LCOI-1490 (5'-GCTCAACAAATCATAAAGATATTGG-3'; Folmer et al. 1994) and the reverse primer COI-Dipt-2183R, also known as COI-780R (5'-CCAAAAAATCARAATARR TGYTG-3'; Gibson et al. 2011). PCR amplification, purification, sequencing protocols, and editing were carried out as described in Rozo-Lopez and Mengual (2015). The Ecuadorian Ministerio del Ambiente y Agua gave access to the genetic resources with the Marco Contract number MAAE-DBI-CM-2021-0167.

The PCR product was visualized on 1.5% agarose gel. PCR products were cleaned using the commercially available QIAquick PCR Purification Kit (QIAGEN[®]). Bidirectional sequencing reactions were carried out by Macrogen Europe BV (Amsterdam, the Netherlands). Chromatograms were edited in Geneious v. 7.1.3. All new sequences were submitted to GenBank via BOLD. GenBank accession numbers are listed for each sequenced specimen in Results.

The software Geneious v. 7.1.3 was used to run a distance-based neighbor-joining (NJ) analysis using the Jukes-Cantor model, in which several publicly accessible DNA barcode sequences of the genus *Aristosyrphus* from BOLD were included together with the newly obtained. The DNA barcode of *Mixogaster mexicana* Macquart, 1846 (BOLD Sample ID: BIOUG58205-F02) was constrained as the root for the NJ tree. All COI sequences can be accessed in BOLD under the Dataset DS-ARISTOSY (<https://doi.org/10.5883/ds-aristosity>). Bootstrap support (BS) values were estimated from 1,000 replicates as spawned in Geneious v. 7.1.3. FigTree v. 1.3.1 (Rambaut 2018) and Adobe[®] Illustrator CS 5.1 were used to draw the NJ tree (Fig. 3).

Results

Aristosyrphus carpenteri (Hull, 1945)

Ceratophya carpenteri Hull 1945: 76. Type locality: Panama, Coclé Province, El Valle (holotype ♀ at the Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts, USA).

Figures 1–3

New records. ECUADOR – **Pichincha** • Parroquia Pedro Vicente Maldonado, near San Pancraccio, roadway to Pachijal, path to the river near forest; 00.1155°N, 078.9584°W; 737 m alt.; 26.XII.2019–02.I.2020; I. Kilian leg.; double malaise trap; 3 ♂ ZFMK-DIP-00067491

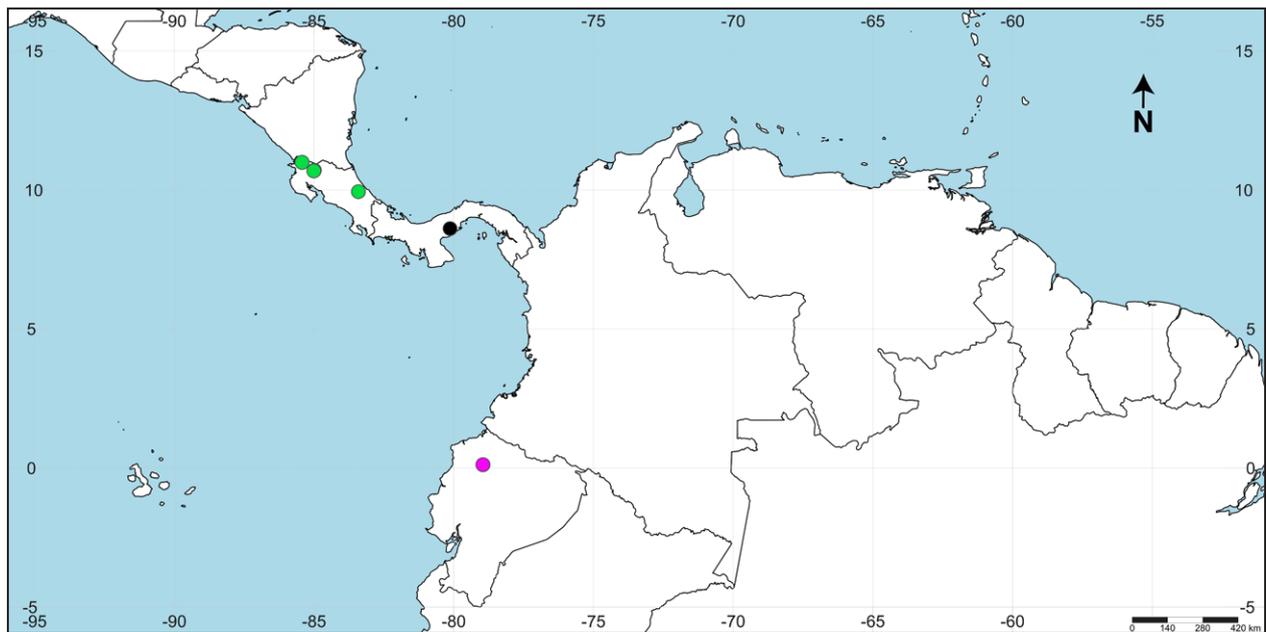


Figure 1. Records of *Aristosyrphus carpenteri*. The black circle is the type locality; green circles indicated localities for specimens in GBIF and BOLD; the pink circle is our new record from Ecuador.

[ZFMK; GenBank ON943477], ZFMK-DIP-00091519 [ZFMK], ZFMK-DIP-00091520 [ZFMK]; 1 ♀ ZFMK-DIP-00067499 [ZFMK; GenBank ON943475] – **Pichincha** • Parroquia Pedro Vicente Maldonado, near San Pancraccio, roadway to Pachijal, area behind platform; 00.1186°N, 078.9580°W; 770 m alt.; 25–28.I.2020; I. Kilian leg.; double malaise trap; 7♂ ZFMK-DIP-00067462 [ZFMK; GenBank ON943474], ZFMK-DIP-00067466 [ZFMK; GenBank: ON943473], ZFMK-DIP-00067476 [ZFMK; GenBank ON943476], ZFMK-DIP-00091515 [ZFMK], ZFMK-DIP-00091516 [ZFMK], ZFMK-DIP-00091517 [INABIO], ZFMK-DIP-00091518 [INABIO].

Identification. Images of the female holotype were studied (available at <https://mczbase.mcz.harvard.edu/guid/MCZ:Ent:26031>) and male genitalia were compared with those of Costa Rican specimens (<https://doi.org/10.15468/dl.c429d9>). No morphological differences were noticed between Central American specimens and the Ecuadorian individuals (Fig. 1).

Species with a convex face, without a facial tubercle, head wider than thorax, and with long antennae, longer than the distance between antennal fossa and anterior oral margin, with postpedicel longer than scape (Fig. 2). As a member of *Aristosyrphus*, it has the postpronotum pilose, the anepimeron entirely pilose, the mesonotum with an incomplete transverse suture, wing vein R_{4+5} without a posterior appendix, and abdomen parallel-sided. Moreover, the ejaculatory hood of the male terminalia is apicodorsally developed into a prong-like structure, separate from the actual phallus (Fig. 2C).

Aristosyrphus carpenteri is most similar to *A. primus* (Reemer and Ståhls 2013a: figs. 27–29); both have wings vividly yellow on the anterobasal half, including veins, contrasting with the infuscated, dark apical half (Fig. 2A, B). *Aristosyrphus carpenteri* differs by having

legs entirely orange (legs entirely black in *A. primus*), a bare katapisternum (katapisternum dorsally pilose in *A. primus*), mesonotum golden pilose (mesonotum mostly black pilose in *A. primus*), and the first three abdominal segments entirely orange (second tergite with a medial dark marking and third tergite with anterior half or more black in *A. primus*). In addition, male genitalia for both species are distinct: surstylus with a dorsal truncated projection, curved dorsal margin, and rounded ventral tip (surstylus with dorsal and ventral triangular tips and almost straight dorsal margin in *A. primus*), and ejaculatory hood strongly curved dorsoapically (ejaculatory hood almost straight apically in *A. primus*).

We successfully sequenced five specimens of *A. carpenteri* collected in Ecuador (GenBank accession numbers ON943473, ON943474, ON943475, ON943476, and ON943477). These five COI sequences were very similar to one another (p -distance = 0.0–0.46%) and also quite similar to other barcode sequences from specimens collected in Costa Rica (0.98–2.13%) (Appendix Table A2). This intraspecific variability in the COI sequence is also seen among the members of an undescribed species from Costa Rica, *Aristosyrphus* sp. 1 (p -distance = 0.0–3.26%) (Appendix Table A2). In our NJ tree (Fig. 3), all specimens identified as *A. carpenteri* clustered together. The Barcode Index Number (BIN) (Ratnasingham and Hebert 2013) for all the *A. carpenteri* specimens is BOLD:ABY2236 (<https://doi.org/10.5883/BOLD:aby2236>).

Discussion

Nothing is known about the biology of *Aristosyrphus* (Reemer 2013), and no molecular data are available to infer its phylogenetic relationships, besides a handful of

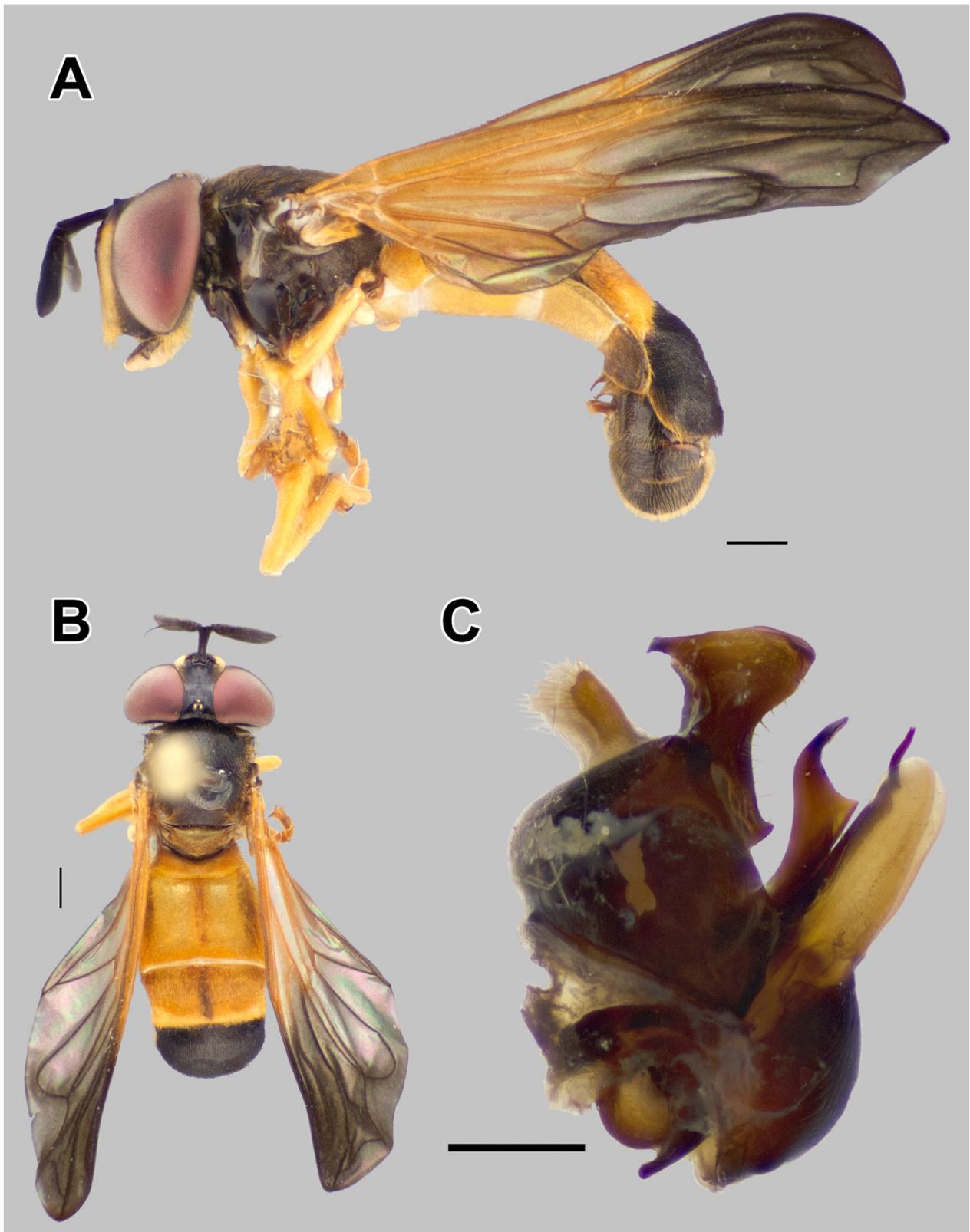


Figure 2. *Aristosyrphus carpenteri*, male, ZFMK-DIP-00091520. **A.** Habitus, lateral view. **B.** Habitus, dorsal view. **C.** Male genitalia, lateral view. Scale bars: A, B = 1 mm; C = 0.5 mm.

COI barcodes (Reemer and Ståhls 2013b; public data in BOLD). Our study reports the first records of the genus *Aristosyrphus* and of *A. carpenteri* from Ecuador. Our data extends the known geographical range of the species by more than 950 km southward (Fig. 1) from the type locality at El Valle [de Antón], Coclé Province,

Panama (08.6098°N, 080.1317°W). Furthermore, the new records are the first specimens of *A. carpenteri* reported from South America, as the species has been only known from Central America until now.

In Costa Rica, specimens of *A. carpenteri* were collected in the provinces of Alajuela, Cartago and

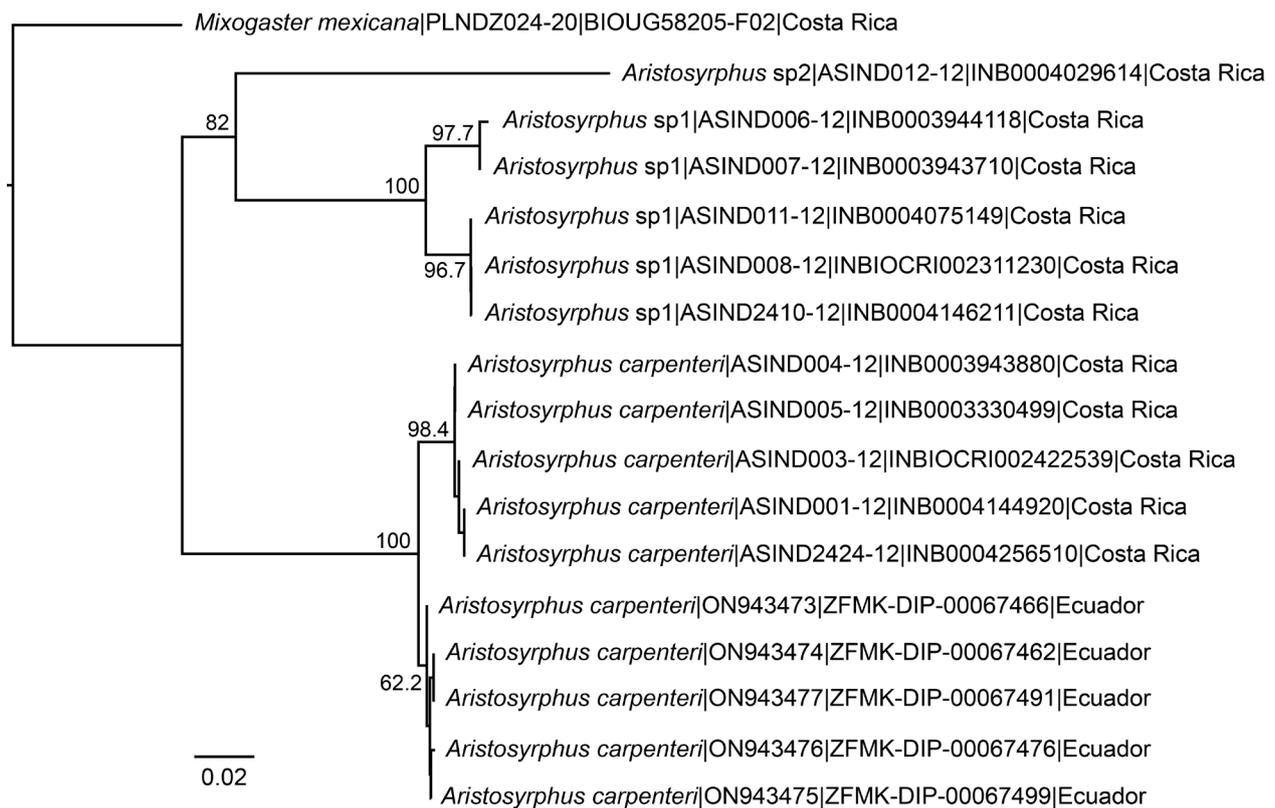


Figure 3. Neighbour-joining tree using Jukes-Cantor model based on public COI sequences of *Aristosyrphus*, with *Mixogaster mexicana* constrained as the outgroup. The name for each specimen has: the name of the species | Process ID or GenBank accession number | sample ID | country of origin. Bootstrap support values are given at the nodes.

Guanacaste, and in Panama the female holotype of *A. carpenteri* was collected in Coclé Province. All these localities belong to the Guatuso-Talamanca biogeographic province (Morrone 2001, 2014) and are located in two distinct ecoregions (Olson et al. 2001): Talamanca montane forests in Costa Rica and Isthmian-Pacific moist forest in Panama. Our new records come from the Cauca biogeographic province (Morrone 2014) and are within the Northwestern Andean montane forest ecoregion. Both biogeographic provinces are threatened by human activities, mostly changes in land use (conversion to agricultural land and pasture) and natural and anthropogenic fires (Dinerstein et al. 1995; Brown and Kappelle 2001; Morrone 2001); however, the Talamanca montane forests still cover 75% of their original area (WWF 2001).

DNA barcoding supports that the specimens from Central America and Ecuador belong to the same species. We believe that the biogeographic range of *Aristosyrphus* species may be larger than what the current records suggest, as reported here by us. Published records of the species from Costa Rica and Panama are from May to July, and our Ecuadorian specimens were collected in December and January. These Ecuadorian specimens may indicate a longer phenology of *A. carpenteri* (possibly year round), that the flight periods differ between Central America and Ecuador, or that the species is not univoltine.

Of the more than 6,200 flower fly species (Skevington

et al. 2019), almost a third are described from the Neotropical Region (Thompson et al. 2010), and this number may represent only half of the actual number of species (Reemer 2016). More faunistic studies are needed to understand the current distribution of flower flies in Central and South America, especially for the Microdontinae since they are highly diverse in the Neotropics (Reemer and Ståhls 2013a; Reemer 2014).

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Authors' Contributions

Conceptualization: XM. Data curation: XM, ICK, APP. Formal analysis: XM. Funding acquisition: XM. Investigation: ICK. Methodology: XM, ICK. Project administration: XM, APP. Resources: ICK, APP. Validation: XM. Visualization: XM. Writing – original draft: XM. Writing – review and editing: XM, ICK, APP.

References

- Amorim DS (2009) Neotropical Diptera diversity: richness, patterns, and perspectives. In: Pape T, Bickel D, Meier R (Eds.) Diptera diversity: status, challenges and tools. Brill, Leiden, the Netherlands, 71–97. <https://doi.org/10.1163/ej.9789004148970.1-459.17>
- Barretto MP, Lane J (1947) Novos Microdontinae brasileiros (Diptera, Syrphidae). *Revista de Entomologia* 18: 139–148.
- Brown AD, Kappelle M (2001) Introducción a los bosques nublados del neotrópico: una síntesis regional. In: Kappelle M, Brown AD (Eds.) Bosques nublados del Neotrópico. Instituto Nacional de Biodiversidad, Santo Domingo de Heredia, Costa Rica, 25–40.
- Cheng X-Y, Thompson FC (2008) A generic conspectus of the Microdontinae (Diptera: Syrphidae) with the description of two new genera from Africa and China. *Zootaxa* 1879: 21–48. <https://doi.org/10.11646/zootaxa.1879.1.3>
- Cumming JM, Wood DM (2017) Adult morphology and terminology. In: Kirk-Spriggs AH, Sinclair BJ (Eds.) Manual of Afrotropical Diptera. Volume 1. Introductory chapters and keys to Diptera families. Suricata 4. South African National Biodiversity Institute, Pretoria, South Africa, 89–133.
- Curran CH (1941) New American Syrphidae. *Bulletin of the American Museum of Natural History* 78: 243–304.
- Dinerstein E, Olson DM, Graham DJ, Webster AL, Primm SA, Bookbinder MP, Ledec G (1995) Una evaluación del estado de conservación de las ecorregiones terrestres de América Latina y el Caribe. World Bank, Washington DC, USA, 135 pp.
- Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from metazoan invertebrates. *Molecular marine Biology and Biotechnology* 3: 294–299.
- GBIF (2022) GBIF occurrence download, *Aristosyrphus carpenteri* (Hull, 1945). <https://doi.org/10.15468/dl.c429d9>. Accessed on: 2015-7-1.
- Gibson JF, Kelso S, Jackson MD, Kits JH, Miranda GFG, Skevington JH (2011) Diptera-specific polymerase chain reaction amplification primers of use in molecular phylogenetic research. *Annals of the Entomological Society of America* 104: 976–997. <https://doi.org/10.1603/an10153>
- Hebert PDN, Cywinska A, Ball SL, deWaard JR (2003) Biological identifications through DNA barcodes. *Proceedings of the Royal Society of London B* 270: 313–321. <https://doi.org/10.1098/rspb.2002.2218>
- Hughes AC, Orr MC, Ma K, Costello MJ, Waller J, Provoost P, Yang Q, Zhu C, Qiao H (2021) Sampling biases shape our view of the natural world. *Ecography* 44: 1259–1269. <https://doi.org/10.1111/ecog.05926>
- Hull FM (1945) Some undescribed syrphid flies. *Proceedings of the New England Zoölogical Club* 23: 71–78.
- Macquart PJM (1846) Diptères exotiques nouveaux ou peu connus. Supplement. [1]. *Memoires de la Société Royal des Sciences, de l'Agriculture et des Arts à Lille* 1844: 133–364, 20 pls.
- Marín-Armijos D, Quezada-Ríos N, Soto-Armijos C, Mengual X (2017) Checklist of the flower flies (Diptera, Syrphidae) of Ecuador. *ZooKeys* 691: 163–199. <https://doi.org/10.3897/zookeys.691.13328>
- Marinoni L, Thompson FC (2004) Flower flies of southeastern Brazil (Diptera: Syrphidae). Part I. Introduction and new species. *Studia Dipterologica* 10: 565–578.
- Mittermeier RA, Mittermeier CG, Gil PR, da Fonseca GAB, Brooks T, Pilgrim J, Konstant WR (2005) Megadiversity: Earth's biologically wealthiest nations. CEMEX, Mexico, 501 pp.
- Morrone JJ (2001) Biogeografía de América Latina y el Caribe. Manuales y Tesis SEA, Volumen 3. Sociedad Entomológica Aragonesa, Zaragoza, Spain, 148 pp.
- Morrone JJ (2014) Biogeographical regionalisation of the Neotropical region. *Zootaxa* 3782: 1–110. <https://doi.org/10.11646/zootaxa.3782.1.1>
- Noh JK, Echeverria C, Kleemann J, Koo H, Fürst C, Cuenca P (2020) Warning about conservation status of forest ecosystems in tropical Andes: national assessment based on IUCN criteria. *PLoS ONE* 15: e0237877. <https://doi.org/10.1371/journal.pone.0237877>
- Olson DM, Dinerstein E, Wikramanayake ED, Burgess ND, Powell GVN, Underwood EC, D'Amico JA, Itoua I, Strand HE, Morrison JC, Loucks CJ, Allnutt TF, Ricketts TH, Kura Y, Lamoreux JF, Wettengel WW, Hedao P, Kassem KR (2001) Terrestrial ecoregions of the world: a new map of life on Earth. *Bioscience* 51: 933–938. [https://doi.org/10.1641/0006-3568\(2001\)051\[0933:teotwaj\]2.0.co;2](https://doi.org/10.1641/0006-3568(2001)051[0933:teotwaj]2.0.co;2)
- Papavero N (1962) Quatro novas espécies de Microdontinae do Brasil (Diptera, Syrphidae). *Papéis Avulsos de Zoologia* 15: 317–326.
- Rambaut A (2018) FigTree v. 1.4.4: Tree Figure Drawing Tool. <https://github.com/rambaut/figtree/releases>. Accessed on: 2021-12-1.
- Ratnasingham S, Hebert PDN (2013) A DNA-based registry for all animal species: the Barcode Index Number (BIN) system. *PLoS ONE* 8: e66213. <https://doi.org/10.1371/journal.pone.0066213>
- Reemer M (2013) Review and phylogenetic evaluation of associations between Microdontinae (Diptera: Syrphidae) and ants (Hymenoptera: Formicidae). *Psyche*: 538316. <https://doi.org/10.1155/2013/538316>
- Reemer M (2014) A review of Microdontinae (Diptera: Syrphidae) of Surinam, with a key to the Neotropical genera. *Tijdschrift voor Entomologie* 157: 27–57. <https://doi.org/10.1163/22119434-00002035>
- Reemer M (2016) Syrphidae (Diptera) of Surinam: Eristalinae and synthesis. *Tijdschrift voor Entomologie* 159: 97–142. <https://doi.org/10.1163/22119434-00002035>
- Reemer M, Ståhls G (2013a) Generic revision and species classification of the Microdontinae (Diptera, Syrphidae). *ZooKeys* 288: 1–213. <https://doi.org/10.3897/zookeys.288.4095>
- Reemer M, Ståhls G (2013b) Phylogenetic relationships of Microdontinae (Diptera: Syrphidae) based on molecular and morphological characters. *Systematic Entomology* 38: 661–688. <https://doi.org/10.1111/syen.12020>
- Ron SR (2020) Regiones naturales del Ecuador. BIOWEB. Pontificia Universidad Católica del Ecuador. <https://bioweb.bio/faunaweb/amphibiaweb/RegionesNaturales>. Accessed on: 2022-9-27.
- Rozo-Lopez P, Mengual X (2015) Mosquito species (Diptera, Culicidae) in three ecosystems from the Colombian Andes: identification through DNA barcoding and adult morphology. *ZooKeys* 513: 39–64. <https://doi.org/10.3897/zookeys.513.9561>
- Shorthouse DP (2010) SimpleMapp, an online tool to produce publication-quality point maps <https://www.simplemapp.net>. Accessed on: 2022-7-4.
- Skevington JH, Locke MM, Young AD, Moran K, Crins WJ, Marshall SA (2019) Field guide to the flower flies of northeastern North America. Princeton University Press, Princeton, USA, 512 pp. <https://doi.org/10.1515/9780691192512>
- Thompson FC (2020) Ant and flower flies: two different groups. *Fly Times* 64: 24–25.
- Thompson FC, Rotheray GE, Zumbado MA (2010) Family Syrphidae. In: Brown BV, Borkent A, Cumming JM, Wood DM, Woodley NE, Zumbado MA (Eds.) Manual of Central American Diptera. Volume 2. NRC Research Press, Ottawa, Canada, 763–792.
- Townes H (1972) A light-weight Malaise trap. *Entomological News* 83: 239–247.
- Troudet J, Grandcolas P, Blin A, Vignes-Lebbe R, Legendre F (2017) Taxonomic bias in biodiversity data and societal preferences. *Scientific Report* 7: 9132. <https://doi.org/10.1038/s41598-017-09084-6>
- WWF 2001. Talamancan montane forests (NT0167). World Wildlife Fund. <https://web.archive.org/web/20100308075124/http://www.nationalgeographic.com/wildworld/profiles/terrestrial/nt/nt0167.html>. Accessed on: 2022-7-4.

