NOTES ON GEOGRAPHIC DISTRIBUTION

 $\bigtriangledown$ 

 $\bigtriangledown$ 

Check List 17 (6): 1521–1531 https://doi.org/10.15560/17.6.1521



Check List the journal of biodiversity data

# Out of Indochina: confirmed specimen record and first molecular identification of *Psammophis indochinensis* Smith, 1943 (Squamata, Psammophiidae) from Bali, Indonesia

Nia Kurniawan<sup>1\*</sup>, Luhur Septiadi<sup>2</sup>, Muhammad Fathoni<sup>1</sup>, Gigih Setia Wibawa<sup>1</sup>, Panupong Thammachoti<sup>2</sup>

- 2 Department of Biology, Faculty of Science, Chulalongkorn University, Bangkok, Thailand LS: luhur.septiadi@gmail.com https://orcid.org/0000-0001-5195-5550 PT: panupong.th@chula.ac.th https://orcid.org/0000-0003-3477-8964
- \* Corresponding author

 $\square$ 

 $\bigtriangledown$ 

#### Abstract

*Psammophis indochinensis* Smith, 1943 was reported in the eastern Java and Bali of Indonesia despite its primary geographic range being in the Indochina region. We confirm its presence in Bali based on a newly collected specimen and provide morphological and genetic data. The specimen was found in a lowland, urban areas near open grassland habitat, which confirms the distribution of *P. indochinensis* along the northern coast of Bali. We note some character aberrations in the supralabials compared to Thailand specimens, suggesting an extended character. The basal clade position of *P. indochinensis* raises the possibility of an intercontinental dispersal scenario of this African-origin snake.

#### Keywords

Disjunct distribution, herpetofauna, reptile, snake, underestimated region

Academic editor: Jesse Grismer | Received 21 July 2021 | Accepted 25 October 2021 | Published 10 November 2021

Citation: Kurniawan N, Septiadi L, Fathoni M, Wibawa GS, Thammachoti P (2021) Out of Indochina: confirmed specimen record and first molecular identification of *Psammophis indochinensis* Smith, 1943 (Squamata, Psammophiidae) from Bali, Indonesia. Check List 17 (6): 1521–1531. https://doi.org/10.15560/17.6.1521

# Introduction

*Psammophis indochinensis* Smith, 1943, Indo-Chinese Sand Snake (Bahasa Indonesia: Ular Pasir Asia), is the only psammophiid snake species occurring in the Indochina region. This species is distributed in Myanmar, Thailand, Lao People's Democratic Republic (Lao PDR), Vietnam, Cambodia, and Indonesia, where it occurs in lowland habitats (Hartmann et al. 2011). However, there are few known occurrences of this species in Indonesia, where has been reported from East Java (Mertens 1957) and Bali (Ineich and Dueve 1990), or are doubtful (Kusrini et al. 2021). Based solely on these accounts, several field guides (McKay 2006; de Lang 2017; Somaweera 2017; Rusli 2020), include East Java and Bali within the range of *P. indochinensis*. Although an extensive

survey in 2013–2019 has been conducted in Java, the species was not encountered at that time, and the previously reported specimen in the Museum Zoologicum Bogoriense (MZB) (Kurniawan et al. 2021) was presumably lost. Nonetheless, a recent survey in Bali Barat National Park (TNBB) in 2015 rediscovered this species (Amarasinghe et al. 2021), but unfortunately, voucher specimens were not collected (Amarasinghe pers. comm. 16 February 2021). As most members of genus *Psammophis* are primarily distributed in Africa, this has raised questions on the biogeographic history and taxonomic status of *P. indochinensis* (Kelly et al. 2008; Hartmann et al. 2011).

Herein, we confirm the presence of *P. indochinensis* in Bali based on a newly collected specimen. We provide morphological and genetic data, as well as updated information on the geographic distribution of this species and its congeners. The collected specimen is the first of *P. indochinensis* from Indonesia after three decades and includes genetic data. We discuss the possibility of intercontinental dispersal of this African-origin snake.

## Methods

The individual of Psammophis indochinensis was caught by hand and immediately transported to the laboratory for examination. We recorded information on habitat, elevation, and geographic coordinates using a Garmin GPS Map 64s receiver. No scientific permits were needed as the voucher specimen was found in an urban area and not listed in the national protected fauna and flora of Indonesia under regulation by the Ministry of Environment and Forestry (PERMEN LHK; Ministry of the Environment 2018). Photographs were taken of the specimen alive in the laboratory (ex-situ) using a Nikon D5200 camera. The snake was euthanized with a dose of 7.5% benzocaine, fixed over 24 h in 10% formalin, preserved in 70% ethanol, and accessioned in the Laboratory of Animal Diversity and Ecology Collection, Biology Department, Universitas Brawijaya, Indonesia (NK), under the catalog number NK 2007. A tissue sample was collected in 95% ethanol for molecular study.

For the morphological characters, we compared the newly collected specimen (NK 2007) to four specimens available in Thailand Natural History Museum, Pathum Thani, Thailand (THNHM) including THNHM 1777–1779 and 1781, which collected by locals on 15 September 1965. We mapped the overall geographic distribution of *Psammophis* species (Fig. 6A) extracted from the literature (Supplemental file, Table S1), which corresponds to each representative clade on phylogenetic trees (Fig. 5). The geographical distribution of *P. indochinensis* was presented along with museum number (only for the collected specimens) obtained from Hartmann et al. (2011), Amarasinghe et al. (2021), and coordinates of the newly collected specimen. We used QGIS v. 3.8.1 (QGIS

Development Team, Zürich, Switzerland) to map the geographic distributions of *Psammophis* species.

Scale counts and nomenclature of external morphology follow Peters (1964); the first number of ventral scales (first plate bordered on both sides by the first row of dorsals) follows Dowling's method (1951), and the terminal scute (precloacal plate) is not included in the number of ventrals. The dorsal scale row counts are given at one head length behind the head, at mid-body (onehalf of the total number of ventrals), and one head length before the vent. The counts for paired head characters are listed in Table 2 in left/right order. All body measurements are to the nearest 0.1 mm. Character abbreviations follow Grismer et al. (2014): ASR = dorsal scale rows at neck, MSR = dorsal scale rows at mid-body, PSR = dorsal scale rows before vent, VEN = number of ventral plates, SC = number of subcaudal scales, ANA = anal plate single or divided, L = number of loreal scales, SL = number of supralabials, SL/Eye = number of supralabials touching eye, Larg SL = largest supralabial, IL = number of infralabials,  $IL/1^{st}$  child = number of infralabials touching anterior chin shield, PreOc = number of preoculars, PostOc = number of postoculars, SuprOc = number of supraoculars, ATem = number of anterior temporals, PTem = number of posterior temporals, SVL = snout-vent length (mm), TaL = tail length (mm), TL = total length (mm), HL = head length measured from the caudal border of exoccipitals to the tip of snout (mm), HW = head width measured at the maximum width of head (mm), Rel TL = tail length relative to total length  $(TaL/TL\times 100)$ , and Rel HW = head width relative to head length (HW/HL×100).

Mitochondrial DNA of the cytochrome b (cytB) gene was extracted from the liver and muscle tissue sample of the specimen NK 2007 using QIAamp DNA Mini Kit (Qiagen) following the manufacturer's protocol. We amplified and sequenced the cytB using primers H16064 (forward: 5'-CTTTGGTTTACAAGAACAAT-GCTTTA-3') and L14910 (reverse: 5'-GACCTGTGAT MTGAAAACCAYCGTTGT-3') (Burbrink et al. 2000). Our derived sequence was deposited in GenBank (MZ970474) and compared with congeners (including 27 Psammophis taxa) and 2 outgroup taxa, Psammophylax rhombeatus (Linneaus, 1758) and Dipsina multimaculata (Smith, 1847), obtained from GenBank (Table 1). Sequences were aligned using ClustalW in MEGA v. 7 with default parameters (Kumar et al. 2016) and manually adjusted, resulting in 1,100 base pairs (439 parsimony-informative, 49 singleton sites, 612 constant sites). Uncorrected *p*-distance was calculated in MEGA v. 7 (Kumar et al. 2016). Nucleotide substitution model was estimated using jModelTest2 (Darriba et al. 2012). The GTR+G was considered as the best-fit substitution model for maximum likelihood (ML) and Bayesian inference (BI) of the cytB alignment. Phylogenetic inferences based on the ML was estimated using IQ-Tree (Nguyen et al. 2015) with 1,000 ultrafast bootstrap replicates (Hoang et al. 2018), and BI was estimated by

Table 1. Samples used in the molecular analyses with GenBank accession numbers. An asterisk (*) indicates identification of the species
was changed from that provided by GenBank.

Species	GenBank AN	Country	Locality	Reference
Psammophylax rhombeatus	DQ486342	South Africa	Grahamstown	Kelly et al. 2008
Dipsina multimaculata	DQ486357	South Africa	Namaqualand	Kelly et al. 2008
sammophis crucifer	DQ486360	South Africa	Somerset East	Kelly et al. 2008
sammophis crucifer	DQ486397	Zimbabwe	Nyanga	Kelly et al. 2008
sammophis jallae	DQ486409	Botswana	Kazungula	Kelly et al. 2008
sammophis trigrammus	DQ486458	Namibia	Sesfontein	Kelly et al. 2008
sammophis trigrammus	DQ486469	Namibia	Brandberg	Kelly et al. 2008
sammophis lineolatus	DQ486450	Turkmenistan	Nephtezavodsk	Kelly et al. 2008
sammophis condanarus	AF471075	Myanmar	Ayeyarwady Division	Lawson et al. 2005
sammophis indochinensis	MZ970474	Indonesia	Buleleng Regency, Bali Province	This study
sammophis angolensis	DQ486416	Zambia	Kabwe	Kelly et al. 2008
sammophis angolensis	DQ486439	Botswana	Kasane	Kelly et al. 2008
sammophis aegyptius	MG002976	Egypt	_	Gonçalves et al. 2018
sammophis aegyptius	MG002972	Egypt	_	Gonçalves et al. 2018
sammophis schokari	DQ486365	Morocco	_	Kelly et al. 2008
sammophis schokari	AY612034	Tunisia	Bou Hedma	Gonçalves et al. 2018
sammophis namibensis	DQ486455	South Africa	Port Nolloth	Kelly et al. 2008
sammophis leightonii	DQ486467	South Africa	Piketberg	Kelly et al. 2008
sammophis notostictus	DQ486362	South Africa	Grahamstown	Kelly et al. 2008
sammophis notostictus	DQ486463	South Africa	Port Nolloth	Kelly et al. 2008
ammophis praeornatus	MH997959	Senegal	Fafakourou	Trape et al. 2019
sammophis punctulatus trivirgatus	DQ486432	Tanzania	Arusha Region	Kelly et al. 2008
ammophis elegans univittatus	MK005679	Chad	Bon Amdaoud	Trape et al. 2019
ammophis elegans	MH997943	lvory Coast	Drekro	Trape et al. 2019
ammophis tanganicus	DQ486378	Tanzania	Dodoma Region	Kelly et al. 2008
ammophis tanganicus	DQ486379	Tanzania	Dodoma Region	Kelly et al. 2008
sammophis biseriatus	DQ486389	Tanzania	Arusha	Kelly et al. 2008
ammophis biseriatus	DQ486448	Kenya	Watamu	Kelly et al. 2008
ammophis lineatus*	FJ404313	Cameroun	Wataniu	Vidal et al. 2008
	MH997938	Chad	 Baïbokoum	Trape et al. 2008
sammophis lineatus		Namibia		
ammophis leopardinus	DQ486456		Opuwo Groothare Dage	Kelly et al. 2008
ammophis leopardinus	DQ486462 MH997952	Namibia	Grootberg Pass	Kelly et al. 2008
ammophis philipsii		Togo	Djiguengué Kindia	Trape et al. 2019
sammophis philipsii	MH997955	Guinea	Kindia Usta in the Wall	Trape et al. 2019
sammophis brevirostris	DQ486402	South Africa	Hole in the Wall	Kelly et al. 2008
sammophis brevirostris	DQ486395	Zimbabwe	Marondera	Kelly et al. 2008
sammophis sibilans	MH997970	Ethiopia	Kutaworje	Trape et al. 2019
sammophis sibilans	MH997934	Egypt	Faiyum	Trape et al. 2019
sammophis mossambicus	MH997940	Chad	Baïbokoum	Trape et al. 2019
sammophis mossambicus	DQ486400	South Africa	Sodwana Bay	Kelly et al. 2008
sammophis rukwae	DQ486443	Kenya	Lake Baringo	Kelly et al. 2008
sammophis rukwae	MH997939	Chad	Baïbokoum	Trape et al. 2019
ammophis afroccidentalis	MH997961	Burkina Faso	Pâ	Trape et al. 2019
sammophis afroccidentalis	MH997960	Senegal	Dar Salam	Trape et al. 2019
sammophis orientalis	DQ486380	Tanzania	Handeni	Kelly et al. 2008
sammophis orientalis	DQ486396	Mozambique	Gorongosa	Kelly et al. 2008
sammophis subtaeniatus	DQ486408	Botswana	Kazungula	Kelly et al. 2008
sammophis subtaeniatus	DQ486415	Zimbabwe	Kariba	Kelly et al. 2008
sammophis sudanensis	MH997948	Chad	Baïbokoum	Trape et al. 2019
sammophis sudanensis	MH997950	Chad	Baïbokoum	Trape et al. 2019

MrBayes v. 3.2.7a (Ronquist et al. 2012) with Monte Carlo Markov Chains (MCMC) for 1,000,000 generations. Both were analyzed through the CIPRES Science Gateway (Miller et al. 2012). We visualized the trees using FigTree v. 1.3.1 (http://tree.bio.ed.ac.uk/software/ figtree/), where colors for each clade (Fig. 5) correspond to overall geographic distribution of *Psammophis* species (Fig. 6A).

# Results

#### *Psammophis indochinensis* Smith, 1943 Figures 1, 2, 3

**New record.** INDONESIA – **Bali Province** • Buleleng Regency, Gerokgak District; 08°09'29.40"S, 114° 42'56.80"E; 9 m alt.; 15.VI.2020; Gigih S. Wibawa leg.;



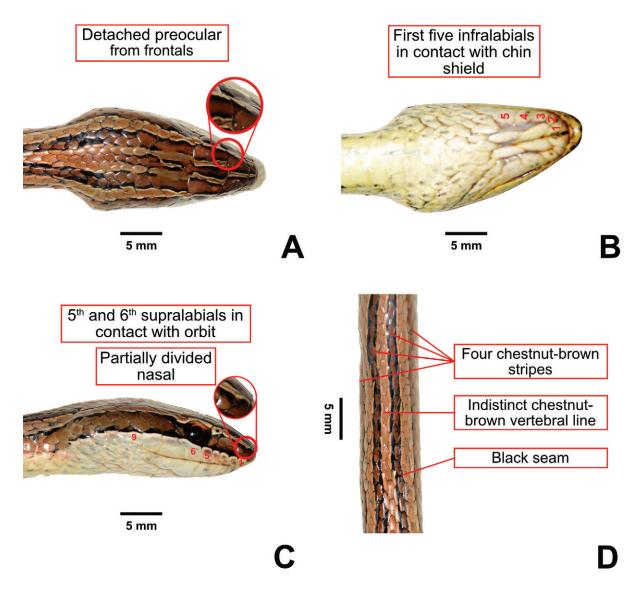
Figure 1. Psammophis indochinensis (NK 2007) from Gerokgak District, Buleleng Regency, Bali Province, Indonesia. A. In life (ex-situ). B. Entire dorsum. C. Entire ventrum. Photographs by M. Fathoni.

lowland and urban area near grassland habitat; 1  $\bigcirc$ , NK 2007.

The specimen was collected in broad daylight (10:00 h), moving fast around G.S. Wibawa's yard. This snake had possibly come from a nearby open grassland habitat (Fig. 4).

Identification. *Psammophis indochinensis* is the only species of its genus that occurs in Indochina. Its nearest congeners which are distributed mainly on the Indian subcontinent (i.e., *P. condanarus* (Merrem, 1820); *P. longifrons* Boulenger, 1890; *P. schokari* (Forskål, 1775); and *P. leithii* Günther, 1869) (Whitaker and Captain 2004; Hartmann et al. 2011; Vyas and Patel 2013). We identified the species using the key provided by Haidar et al. (2020): (1) smooth dorsals; (2) four chestnutbrown stripes, dorsally; and (3) two thin ventrolateral lines. *Psammophis indochinensis* is most similar to *P. condanarus* (characters in parentheses) but differs by the following diagnostic traits (Hughes 1999): (1) ventral scales fewer, numbering 156–173 (165–179); (2) subcaudal scales fewer, numbering 66–85 (75–93); (3) dorsal stripes four and lighter (dorsal stripes five and darker).

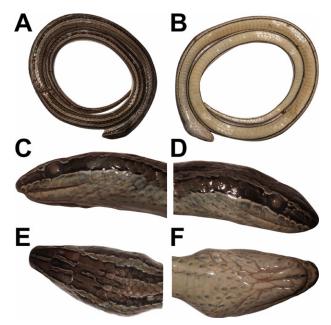
We compare our specimen with the scale ranges reported by Somaweera 2017 (characters in parentheses) as follows: 17:17:13 rows of smooth and shiny dorsals (17 or 19 at midbody); 163 ventrals (150–173); divided anal shield; and 27 paired subcaudals along



**Figure 2.** *Psammophis indochinensis* (NK 2007) from Gerokgak District, Buleleng Regency, Bali Province, Indonesia. **A.** Dorsal head. **B.** Ventral head. **C.** Right lateral head. **D.** Pattern of dorsum at midbody. Photographs by M. Fathoni.

an incomplete tail (75-85 in complete tails). The SVL of our specimen is 540 mm, HL is 19 mm, HW is 13 mm, and TaL is 57 mm but the tail is incomplete. Dorsal head surface shiny; rostral wider than high, hardly visible from above; internasals slightly longer than wide; prefrontals longer than wide; frontal narrowing posteriorly, elongate; supraoculars wider than frontal; parietals slightly shorter than frontal; two pair of chin shields, anterior larger than posterior; nasals partly divided, two times longer than high; two preoculars, upper part reaching top of head, not touching frontal; suboculars absent; two small postoculars; nine supralabials, fifth and sixth contacting orbit, seventh largest; 10 infralabials, first five in contact with chin shields; one temporal in 1st row followed by three temporals in 2<sup>nd</sup> row. In life, body pale olive, with four chestnutbrown stripes from snout to body, bordered by a narrow black margin, most prominent on midbody and anterior; head with an additional median stripe; chin and throat white and mottled with black; venter uniformly whitish yellow with two parallel thin longitudinal black ventrolateral lines from neck to tail (Figs. 1, 2, Table 2). Color after preservation dull and paler (Fig. 3). NK 2007 differs from THNHM 1777–1779 and 1781 (characters in parentheses) by having 5<sup>th</sup> and 6<sup>th</sup> supralabials contacting orbit (4<sup>th</sup> and 5<sup>th</sup>, collectively); higher Rel HW: 0.68 (0.50-0.61, collectively).

**Phylogeny.** Based on BI and ML phylogenetic analyses (Fig. 5), *P. indochinensis* is a sister taxon to the *P. condanarus* from Myanmar (clade 2). These closely related species differ from one another by an uncorrected *p*-distance of 0.0419 (4% in percentage; Supplemental file, Table S2). *Psammophis crucifer* (Daudin, 1803) from southern Africa (clade 1) and three Eurasian species— *P. lineolatus* (Brandt, 1838) from Turkmenistan, *P. condanarus* from Myanmar, and *P. indochinensis* from Indonesia—(clade 2) are the earliest-diverging lineages of the genus.



**Figure 3.** *Psammophis indochinensis* (NK 2007) from Gerokgak District, Buleleng Regency, Bali Province, Indonesia, after preservation. **A.** Entire dorsum. **B.** Entire ventrum. **C.** Left lateral head. **D.** Right lateral head. **E.** Dorsal head. **F.** Ventral head. Photographs by M. Fathoni.

### Discussion

Morphological characters of Psammophis indochinensis have been reported from specimens from the Indochina region. In our Bali specimen (NK 2007), we noted aberrations in some characters (see above and Table 1). Haidar et al (2020) noted that the supralabials touch the eyes in both P. condanarus and P. indochinensis, which differs from P. schokari (4th and 5th vs. 5th and 6th), and NK 2007 had the 5<sup>th</sup> and 6<sup>th</sup> supralabials touching eye (Fig. 2C). Cox et al. (2012), Das (2012), and de Lang (2017) recognized P. indochinensis by having fewer supralabials (SL: 8), which differs from NK 2007 and THNHM 1777 which have nine supralabials (Fig. 2C). Thereby, characters on supralabials touching eye and total number of supralabials should be extended for P. indochinensis. Although the specimen from East Java (MZB 65 1) was lost (Kurniawan et al. 2021), the brief description of this specimen by Mertens (1957) showed that it is similar to Thailand specimens in having the 4<sup>th</sup> and 5<sup>th</sup> supralabials touching the eye. Additional comparisons of specimens, such as MNHN 1989-49 reported from Bali (Ineich and Dueve 1990), may reveal if this character state is unique to the Bali population.

Sex determination on our specimen was only possible by dissection, as sexual dimorphism (e.g., longer tail, longer SVL) was not observed in *P. condanarus* (Cottone et al. 2009) and most other species of *Psammophis* (de Haan 1999). Besides, sex determination using tail length character in *P. indochinensis* is impractical since most of our observed specimens had lost their tail. This is possibly due to caudal autotomy, which is an antipredator mechanism that is also possessed by other *Psammophis* 

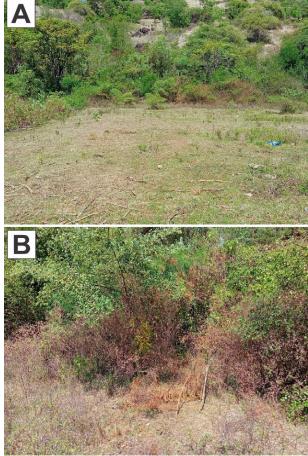


Figure 4. Landscape view where *Psammophis indochinensis* (NK 2007) was found in Gerokgak District, Buleleng Regency, Bali Province, Indonesia. A. Open grassland. B. Nearby dry grasslands habitat. Photographs by G. S. Wibawa.

species (Broadley 1987).

Our specimen was found in lowland and urban areas near open grassland habitats comprise of scattered vegetations (Fig. 4). Psammophis indochinensis has been previously reported in Borassus palm savannah near the sea (Ineich and Dueve 1990), in a wide range of habitats including grassland, open forest, agricultural areas, scrubland, and savannah (Das 2012; Rusli 2020), and in lowland deciduous monsoon forest and moist forest (Amarasinghe et al. 2021). The elevational range has been reported between 100 and 2000 m (McKay 2006; Das 2012), between 983 and 2000 m (Cox et al. 2012); and at sea level (new data herein). Considering those similar habitats and elevations are present on adjacent islands (e.g., Lesser Sunda), with more sampling it is likely that P. indochinensis may also be found to occur on nearby islands. Our new record eliminates the doubt by Kusrini et al. (2021) that P. indochinensis occurs in Indonesia.

We confirm the exact location of the species in Buleleng Regency (this study; NK 2007), as it was previously reported near Mount Merbuk in Bali of Indonesia (Ineich and Dueve 1990; MNHN 1989-49). This species also occurs at Bali Barat National Park (TNBB) (Amarasinghe et al. 2021), 26 km west from Buleleng Regency. In Java, it was recorded in Gresik of East Java (Mertens

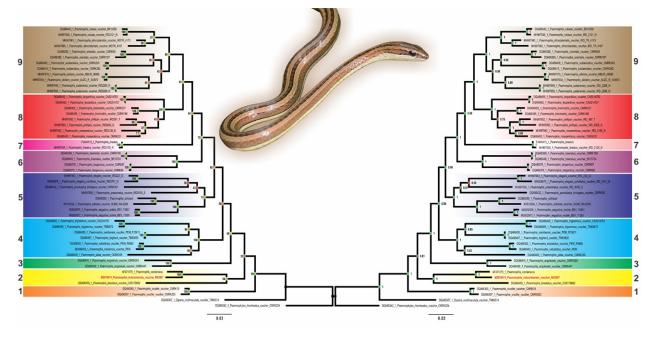
Table 2. Scalation and measurement of P. indochinensis from Indonesia (NK 2007) and Thailand (THNHM 1777–79 and 1781). Abbreviations
are listed in the Methods. An en dash (-) indicates that the tail was incomplete.

Specimen ID	<b>THNHM 1777</b>	<b>THNHM 1778</b>	<b>THNHM 1779</b>	<b>THNHM 1781</b>	NK 2007
Locality		Thai	land		Indonesia
	Pathum Thani	Pathum Thani	Pathum Thani	Pathum Thani	Bali
SEX	ę	ę	ę	ę	ę
ASR	17	17	19	19	17
MSR	17	16	17	17	17
PSR	12	13	13	13	13
VEN	161	164	171	167	163
SC	71	28 ()	83	76	27 (–)
ANA	Single	Single	Single	Single	Single
L	1/1	1/1	1/1	1/1	1/1
SL	9/9	8/8	8/8	8/8	9/9
SL/Eye	4.5/4.5	4.5/4.5	4.5/4.5	4.5/4.5	5.6/5.6
Larg SL	7/7	6/6	7/7	6/6	7/7
IL	10/10	10/10	10/10	10/10	10/10
IL/1 <sup>st</sup> child	4/4	4/4	4/4	4/4	4/4
PreOc	2/2	1/1	1/1	1/1	2/2
Pst0c	2/2	2/2	2/2	2/2	2/2
SuprOc	1/1	1/1	1/1	1/1	1/1
ATem	2/2	1/1	1/1	1/1	1/1
PTem	3/3	3/3	3/3	3/3	3/3
SVL (mm)	649.0	349.0	452.0	374.0	540.0
TaL (mm)	214.0	113.0 (–)	153.0	112.0	57.0 (–)
TL (mm)	863.0	462.0 (-)	605.0	486.0	597.0 (–)
HL (mm)	20.1	13.1	13.5	13.3	19.1
HW (mm)	10.1	8.1	8.3	8.0	13.0
Rel TL	0.24	0.24 (-)	0.25	0.23	0.9 (–)
Rel HW	0.50	0.61	0.61	0.60	0.68

1957; MZB 65 1), 252 km northwest from Buleleng Regency. In Indochina region, it was recorded in Phetchaburi Province of Thailand (Pauwels et al. 2003), more than 2700 km north of Bali and East Java, which means that this species has a highly disjunct distribution. This distribution pattern is similar to the Daboia siamensis (Smith, 1917), which is primarily distributed in Indochina region but has been recorded in East Java and on Lesser Sunda and inhabiting seasonally dry climates (de Lang 2017; Kurniawan et al. 2021). One may argue that the occurrence of P. indochinensis in Indonesia was accidentally introduced by trade (Liong, 1958) or transported with wood, as reported by Mertens (1957) for P. sibilans (Linnaeus, 1758). Previous phylogenetic studies (Kelly et al. 2008) and our phylogenetic trees (Fig. 5) showed that not only P. indochinensis, but also P. condanarus and P. lineolatus (Eurasian taxa) are the earliest-diverging lineages of Psammophis. This raises interesting questions on its biogeographic history and leads us to hypothesize the possibility of the intercontinental dispersal scenario of this African-origin snake.

Disjunct distribution among closely related taxa across continent is not uncommon and has been reported in lizards (Brandley et al. 2011), rat snakes (Burbrink and Lawson 2007), iguanas (Keogh et al. 2008), crocodiles (Oaks 2011), vipers (Wüster et al. 2008), and hingedteeth snakes (Chen et al. 2013). The family Psammophiidae first emerged around 30 Mya (Zaher et al. 2019) in Africa. Our phylogenetic analysis indicates that the most recent common ancestor of the earliest diverging and extant taxa (i.e., *P. indochinensis*, *P. condanarus*, and *P. lineolatus*) may be distributed via the Gomphotherium landbridge which connected the Eurasian and Afro-Arabian plates around 15 Mya (Rögl 1999) after the Arabian plate was separated from the African plate by the formation of the Red Sea around 25 Mya (Bosworth et al. 2005). At that time, the most recent common ancestor of the earliest diverging and extant taxa may have successfully colonized Eurasia in suitable habitats of dry deciduous forests, scrub, and savannah along the coast of central-southeast Asia (Morley 2018).

The next major process was the extirpation of these most recent common ancestors in the Arabian and African plates. If these recent common ancestors successfully colonized Eurasia continent, this indicates that they were once present in Arabian and African plates. Due to the progressive aridification, geological instability, and climatic changes in Arabia and Africa plates during the Miocene and Plio-Pleistocene (Tejero-Cicuéndez et al. 2020), Psammophis may have radiated into many clades in the southern, western, and northern coasts of Africa as well as the central part of the continent, similar to Acanthodactylus (Tamar et al. 2016). Presumably the earliest diverging Psammophis species in Africa and Arabia went extinct due to the aridification, although this is weakly supported due to the lack of extant species. The low species diversity in the Eurasian lineage (only three extant species), may be due to increased extinction rates



**Figure 5.** Maximum likelihood (ML; left) and Bayesian inference (BI; right) trees of cytB alignment showing the basal clade position of *Psammophis indochinensis* (NK 2007) from Gerokgak District, Buleleng Regency, Bali Province, Indonesia. The bootstrap supports for ML and Bayesian posterior probability for BI are indicated by numbers and colors (ML: 0–100, BI: 0–1; green: strongly supported, red: weakly supported). Sequence generated in this study is shown in red type.

as a result of environmental pressures and/or reduced speciation rate (Burbrink and Pyron 2010).

Similar to the above scenario, the occurrence of P. indochinensis in Indonesia could be explained by inquiring to last glacial period (LGP), when the "Savanah corridor" existed through the Sundaland continent (Bird et al. 2005). This corridor-formerly proposed by Heaney (1991)-encompassed some parts of the present-day Gulf of Thailand, eastern coast of peninsular Malaysia, Java Sea, eastern Java, and Bali. This corridor may have provided a distribution route for P. indochinensis to Bali, leading to the possibility of intercontinental dispersal. This corridor submerged due to a sea-level rising after the LGP, leaving only several savannahs in eastern Java and Bali. By inference, this hypothesis supports the previously reported occurrences of P. indochinensis in East Java and Bali (Mertens 1957; Ineich and Dueve 1990), which would mean that this species is native there. However, this explanation can only be justified by further analysis of a divergence time estimation.

Overall, our new record provides additional data on the distribution and morphological variation required for resolving the biogeographical history of *P. indochinensis*. Continuous and intensive survey efforts will surely yield more knowledge on the extent of distribution of herpetofauna species in these regions.

# Acknowledgements

We express our gratitude to Sunchai Makchai for allowing us to examine specimens in the THNHM (Pathum Thani, Thailand). We thank Gunther Köhler, Senckenberg Forschungsinstitut und Naturmuseum (Frankfurt, Germany) for providing the information on specimens and literature. We are especially grateful to Justin M. Bernstein (Rutgers University-Newark, USA), Matthew D. Buehler (Villanova University, USA), Evan S. H. Quah (Universiti Sains Malaysia, Malaysia), and Jesse Grismer (La Sierra University, USA) for their reviews and helpful comments on the manuscript. This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

# Authors' Contributions

Conceptualization: NK. Data curation: MF. Investigation: NK, GSW, PT. Resources: GSW, PT. Visualization: MF. Writing – original draft: NK. Writing – review and editing: LS.

#### References

- Amarasinghe AT, Putra CA, Henkanaththegedara SM, Dwiyahreni AA, Winarni NL, Margules C, Supriatna J (2021) Herpetofaunal diversity of West Bali National Park, Indonesia with identification of indicator species for long-term monitoring. Global Ecology and Conservation 2021: e01638. https://doi.org/10.1016/j.gecco.2021. e01638
- Bird MI, Taylor D, Hunt C (2005) Palaeoenvironments of insular Southeast Asia during the Last Glacial Period: a savanna corridor in Sundaland?. Quaternary Science Reviews 24 (20–21): 2228– 2242. https://doi.org/10.1016/j.quascirev.2005.04.004
- Bosworth W, Huchon P, McClay K (2005) The Red Sea and Gulf of Aden basins. Journal of African Earth Sciences 43 (1–3): 334–378. https://doi.org/10.1016/j.jafrearsci.2005.07.020
- Boulenger GA (1890) The fauna of British India, including Ceylon and Burma. Reptilia and Batrachia. Taylor & Francis, London, UK, 548 pp.
- Boulenger GA (1896) Catalogue of the snakes in the British Museum

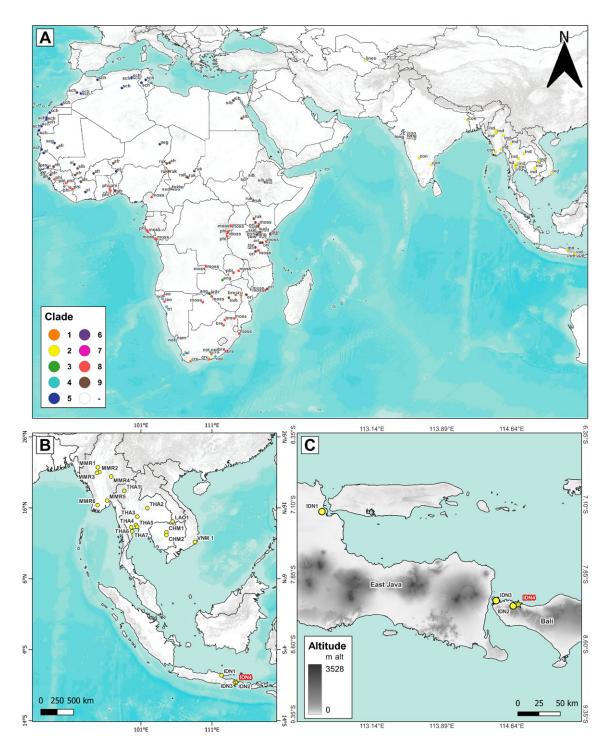


Figure 6. Overall geographic distribution of several Psammophis members in Africa and Eurasia. A. Distribution of Psammophis in Africa and Eurasia. Abbreviations for Fig. 6A are as follows, Clade 1: (cru) P. crucifer; Clade 2: (con) P. condanarus; (ind) P. indochinensis; (lineo) P. lineolatus; Clade 3: (ang) P. angolensis; Clade 4: (jall) P. jallae; (lei) P. leightoni; (nam) P. namibensis; (not) P. notostictus; (tri) P. trigrammus; Clade 5: (aeg) P. aegyptius; (el) P. elegans; (pra) P. praeornatus; (pun) P. punctulatus trivirgatus; (sch) P. schokari; Clade 6: (bis) P. biseriatus; (tan) P. tanganicus; Clade 7: (linea) P. lineatus; Clade 8: (bre) P. brevirostris; (leo) P. leopardinus; (moss) P. mossambicus; (phi) P. phillipsii; Clade 9: (afr) P. afroccidentalis; (ori) P. orientalis; (ruk) P. rukwae; (sib) P. sibilans; (sub) P. subtaeniatus; (sud) P. sudanensis; Clade - (not included in trees): (long) P. longifrons; (sp.); Psammophis sp. (see Supplemental material, Table S1 for details); where the color of dots corresponds to their respective clade (Fig. 5). B. Distribution of P. indochinensis in Indochina region and Indonesia. C. Distribution of P. indochinensis in Bali and Java. Abbreviations for Figs. 6B-C are as follows, MYANMAR: (MMR1) Sagaing Division (Wogan et al. 2008); (MMR2) Mandalay Division (Wogan et al. 2008); (MMR3) Magwe Division (Wogan et al. 2008); (MMR4) Shan State (Boulenger 1896); (MMR5) Bago Division (Boulenger 1890); THAILAND: (THA1) Chiang Mai Province (Taylor 1965); (THA2) Chaiyaphum Province (Das 2010); (THA3) Lopburi Province (Smith 1943); (THA4) Pathum Thani (this study; THNHM 1777–79, 1781); (THA5) Bangkok (Smith 1943; ZFMK 16 658); (THA6) Ratchaburi Province (Chan-Ard et al. 1999); (THA7) Phetchaburi Province (Pauwels et al. 2003); LAO PDR: (LAO1) Champasak Province (Stuart 1998; Teynié et al. 2004); CAMBODIA: (CHM1) Siem Reap Province (Hartmann et al. 2011); (CHM2) Kampong Thom Province (Hartmann et al. 2011); VIETNAM: (VNM1) Ninh Thuan Province (Smith 1943, Nguyen et al. 2009; ZFMK 88 831); INDONESIA: (IDN1) Gresik, East Java Province (Mertens 1957; MZB 65 1); (IDN2) Mount Merbuk, Bali Province (Ineich and Dueve 1990; MNHN 1989-49); (IDN3) Bali Barat National Park, Bali Province (Amarasinghe et al. 2021); (IDN4) Buleleng Regency, Bali Province (this study; NK 2007).

(Natural History). Volume III, containing the Colubridae (Opisthoglyphae and Proteroglyphae), Amblycephalidae, and Viperidae. British Museum, London, UK, 727 pp.

- Brandley MC, Wang Y, Guo X, Montes de Oca AN, Fería-Ortíz M, Hikida T, Ota H (2011) Accommodating heterogenous rates of evolution in molecular divergence dating methods: an example using intercontinental dispersal of *Plestiodon (Eumeces)* lizards. Systematic Biology 60 (1): 3–15. https://doi.org/10.1093/sysbio/ syq045
- Broadley DG (1987) Caudal autotomy in African Snakes of the Genera *Natriciteres* Loveridge and *Psammophis* Boie. The Journal of the Herpetological Association of Africa 33 (1): 18–19. https://doi. org/10.1080/04416651.1987.9650171
- Burbrink FT, Lawson R (2007) How and when did Old World ratsnakes disperse into the New World? Molecular phylogenetics and evolution 43 (1): 173–189. https://doi.org/10.1016/j.ympev.2006.09.009
- Burbrink FT, Lawson R, Slowinski, JB (2000) Mitochondrial DNA phylogeography of the polytypic North American rat snake (*Ela-phe obsoleta*): a critique of the subspecies concept. Evolution 54 (6): 2107–2118. https://doi.org/10.1554/0014-3820(2000)054[2107:md potp]2.0.co;2
- Burbrink FT, Pyron RA (2010) How does ecological opportunity influence rates of speciation, extinction, and morphological diversification in New World ratsnakes (tribe Lampropeltini)? Evolution: International Journal of Organic Evolution 64 (4): 934–943. https://doi.org/10.1111/j.1558-5646.2009.00888.x
- Chan-Ard T, Großmann W, Gumprecht A, Schulz KD (1999) Amphibians and reptiles of Peninsular Malaysia and Thailand: an illustrated checklist. Bushmaster Publications, Würselen, Germany, 240 pp.
- Chen X, Huang S, Guo P, Colli GR, de Oca ANM, Vitt LJ, Pyron RA, Burbrink FT (2013) Understanding the formation of ancient intertropical disjunct distributions using Asian and Neotropical hinged-teeth snakes (*Sibynophis* and *Scaphiodontophis*: Serpentes: Colubridae). Molecular Phylogenetics and Evolution 66 (1): 254–261. https://doi.org/10.1016/j.ympev.2012.09.032
- Cottone AM, Aaron MB (2009) Notes on sexual size dimorphism and reproduction in the Asian Sand Snake, *Psammophis condanarus* (Psammophiidae). Hamadryad 34 (1): 182–185.
- Cox MJ, Hoover MF, Chanhome L, Thirakhupt K (2012) The snakes of Thailand. Chulalongkorn University Museum of Natural History, Bangkok, Thailand, 845 pp.
- Darriba D, Taboada GL, Doallo R, Posada D (2012) jModelTest 2: more models, new heuristics and parallel computing. Nature Methods 9 (8): 772–772. https://doi.org/10.1038/nmeth.2109
- Das I (2010) A field guide to the reptiles of South-East Asia. New Holland Publishers, London, UK, 376 pp.
- Das I (2012) Naturalist's guide to the snakes of South-East Asia: Malaysia, Singapore, Thailand, Myanmar, Borneo, Sumatra, Java and Bali. John Beaufoy Publishing, Oxford, UK, 160 pp.
- de Haan C (1999) Malpolon monspessulanus. In: Böhme W (Ed.) Handbuch der Reptilien und Amphibien Europas. Aula Verlag, Wiesbaden, 661–756.
- de Lang R (2017) The snakes of Java, Bali and surrounding islands. Edition Chimaira, Frankfurt am Main, Germany, 299–300.
- Dowling HG (1951) A proposed standard system of counting ventrals in snakes. British Journal of Herpetology 1 (5): 97–99.
- Gonçalves DV, Martínez-Freiría F, Crochet PA, Geniez P, Carranza S, Brito JC (2018) The role of climatic cycles and trans-Saharan migration corridors in species diversification: biogeography of *Psammophis schokari* group in North Africa. Molecular Phylogenetics and Evolution 118: 64–74. https://doi.org/10.1016/j. ympev.2017.09.009
- Grismer LL, Quah ES, Anuar S, Muin MA, Wood Jr PL, Nor SAM (2014) A diminutive new species of cave-dwelling wolf snake (Colubridae: *Lycodon* Boie, 1826) from Peninsular Malaysia. Zootaxa 3815 (1): 51–67. https://doi.org/10.11646/zootaxa.3815.1.3

Haidar IK, Romon BB, Chowdhury MA (2020) First confirmed re-

cord of the Condanarous Sand Snake (*Psammophis condanarus*) (Reptilia: Squamata: Lamprophildae) from Bangladesh. Journal of Asia-Pacific Biodiversity 13: 716–719. https://doi.org/10.1016/j. japb.2020.08.013

- Hartmann T, Handschuh M, Böhme W (2011) First record of *Psammophis indochinensis* Smith, 1943 from Cambodia, within the context of a distributional species account. Cambodian Journal of Natural History 7: 7–10.
- Heaney LR (1991) A synopsis of climatic and vegetational shange in Southeast Asia. In: Myers N (Ed.) Tropical forests and climate. Springer, Dordrecht, Germany, 53–61. https://doi. org/10.1007/978-94-017-3608-4 6
- Hoang DT, Chernomor O, von Haeseler A, Minh BQ, Vinh, LS (2018) UFBoot2: improving the ultrafast bootstrap approximation. Molecular Biology and Evolution 35 (2): 518–522. https://doi. org/10.1186/s12862-018-1131-3
- Hughes B (1999) Critical review of a revision of *Psammophis* (Linnaeus 1758) (Serpentes, Reptilia) by Frank Brandstätter. African Journal of Herpetology 48 (1–2): 63–70.
- Ineich I, Dueve J (1990) *Psammophis condanarus*. Herpetological Review 21: 23.
- Kelly CM, Barker NP, Villet MH, Broadley DG, Branch WR (2008) The snake family Psammophiidae (Reptilia: Serpentes): phylogenetics and species delimitation in the African sand snakes (*Psammophis* Boie, 1825) and allied genera. Molecular Phylogenetics and Evolution 47 (3): 1045–1060. https://doi.org/10.1016/j. ympev.2008.03.025
- Keogh JS, Edwards DL, Fisher RN, Harlow PS (2008) Molecular and morphological analysis of the critically endangered Fijian iguanas reveals cryptic diversity and a complex biogeographic history. Philosophical Transactions of the Royal Society B: Biological Sciences 363 (1508): 3413–3426. https://doi.org/10.1098/ rstb.2008.0120
- Kumar S, Stecher G, Tamura K (2016) MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. Molecular biology and evolution 33 (7): 1870–1874. https://doi.org/10.1093/ molbev/msw054
- Kurniawan N, Fathoni M, Fatchiyah F, Septiadi L, Smith EN (2021) Composition, distribution, and habitat type of snakes in Java, with discussion on the impact of human-snake interactions during 2013–2019. Herpetology Notes 14: 691–711.
- Kusrini MD, Hamidy A, Prasetyo LB, Nugraha R, Andriani D, Fadhila N, Hartanto E, Afrianto A (2021) Creation of an amphibian and reptile atlas for the Indonesian islands of Java and Bali reveals gaps in sampling effort. Herpetology Notes 14: 1009–1025.
- Lawson R, Slowinski JB, Crother BI, Burbrink FT (2005) Phylogeny of the Colubroidea (Serpentes): new evidence from mitochondrial and nuclear genes. Molecular phylogenetics and evolution 37 (2): 581–601. https://doi.org/10.1016/j.ympev.2005.07.016
- Liong ALG (1958) Sedikit tentang daerah disekitar Gresik. Penggemar Alam 103–104: 67–77.
- McKay JL (2006) A field guide to the amphibians and reptiles of Bali. Krieger Publishing Company, Malabar, USA, 129–131.
- Mertens R (1957) Zur Herpetofauna von Ostjava und Bali. Senckenbergiana Biologica 38: 23–32.
- Miller MA, Pfeiffer W, Schwartz T (2012) The CIPRES science gateway: enabling high-impact science for phylogenetics researchers with limited resources. In: Proceedings of the 1st Conference of the Extreme Science and Engineering Discovery Environment: bridging from the extreme to the campus and beyond 39: 1–8. https://doi.org/10.1145/2335755.2335836
- Ministry of Environment and Forestry (2018) Peraturan Menteri Lingkungan Hidup dan Kehutanan Tentang Perubahan Atas Peraturan Menteri Lingkungan Hidup dan Kehutanan Nomor P.20/ MENLHK/SETJEN/KUM.1/6/2018 Tentang Jenis Tumbuhan dan Satwa yang Dilindungi. Ministry of Environment and Forestry, Jakarta, Indonesia, 30 pp.

- Morley RJ (2018) Assembly and division of the South and South-East Asian flora in relation to tectonics and climate change. Journal of Tropical Ecology 34 (4): 209–234. https://doi.org/10.1017/ S0266467418000202
- Nguyen LT, Schmidt HA, von Haeseler A, Minh BQ (2015) IQ-TREE: a fast and effective stochastic algorithm for estimating maximumlikelihood phylogenies. Molecular Biology and Evolution 32 (1): 268–274. https://doi.org/10.1093/molbev/msu300
- Nguyen VS, Ho TC, Nguyen QT (2009) Herpetofauna of Vietnam. Edition Chimaira, Frankfurt am Main, Germany, 768 pp.
- Oaks JR (2011) A time-calibrated species tree of Crocodylia reveals a recent radiation of the true crocodiles. Evolution: International Journal of Organic Evolution 65 (11): 3285–3297. https://doi. org/10.1111/j.1558-5646.2011.01373.x
- Pauwels OS, David P, Chimsunchart C, Thirakhupt K (2003) Reptiles of Phetchaburi Province, Western Thailand: a list of species, with natural history notes, and a discussion on the biogeography at the Isthmus of Kra. Tropical Natural History 3 (1): 23–53.
- Peters JA (1964) Dictionary of Herpetology. Hafner Publishing Company, New York, USA, 392 pp.
- Rögl F (1999) Mediterranean and Paratethys, facts and hypotheses of an Oligocene to Miocene paleogeography (short overview). Geologica carpathica 50 (4): 339–349.
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic biology 61 (3): 539–542. https://doi.org/10.1093/sysbio/sys029
- Rusli N (2020) Snake of Java: a photographic guide. Indonesia Herpetofauna Foundation, Bogor, Indonesia, 143 pp.
- Smith MA (1943) The fauna of British India, Ceylon and Burma, including the whole of the Indochinese subregion. Reptilia and Amphibia. Vol. III, Serpentes. Taylor & Francis, London, UK, 584 pp.
- Somaweera R (2017) A naturalist's guide to the reptiles and amphibians of Bali. John Beaufoy Publishing, Oxford, UK, 176 pp.
- Stuart B (1998) A survey of amphibians and reptiles in Dong Khantung Proposed National Biodiversity Conservation Area, Champasak Province, Lao PDR. CPAWN/ Wildlife Conservation Society, Vientiane, Lao PDR, 35 pp.
- Tamar K, Carranza S, Sindaco R, Moravec J, Trape JF, Meiri S (2016) Out of Africa: phylogeny and biogeography of the widespread genus *Acanthodactylus* (Reptilia: Lacertidae). Molecular Phylogenetics and Evolution 103: 6–18. https://doi.org/10.1016/j. ympev.2016.07.003
- Taylor EH (1965) The serpents of Thailand and adjacent waters. University of Kansas Science Bulletin 45: 609–1096.
- Tejero-Cicuéndez H, Patton AH, Caetano DS, Šmíd J, Harmon LJ, Carranza S (2021) Reconstructing squamate biogeography in Afro-Arabia reveals the influence of a complex and dynamic geologic past. Systematic Biology 2021: syab025 https://doi.

#### org/10.1093/sysbio/syab025

- Teynié A, David P, Ohler A, Luanglath K (2004) Notes on a collection of amphibians and reptiles from southern Laos, with a discussion of the occurrence of Indo-Malayan species. Hamadryad 29: 33–62.
- Trape JF, Crochet PA, Broadley, DG, Sourouille P, Mané Y, Burger M, Böhme W, Saleh M, Karan A, Lanza B, Mediannikov O (2019) On the *Psammophis sibilans* group (Serpentes, Lamprophiidae, Psammophiinae) north of 12 S, with the description of a new species from West Africa. Bonn Zoological Bulletin 68 (1): 61–91.
- Vidal N, Branch WR, Pauwels OS, Hedges SB, Broadley DG, Wink M, Cruaud C, Joger U, Nagy ZT (2008) Dissecting the major African snake radiation: a molecular phylogeny of the Lamprophiidae Fitzinger (Serpentes, Caenophidia). Zootaxa 1945 (1): 51–66. https://doi.org/10.11646/zootaxa.1945.1.3
- Vyas R, Patel H (2013) Notes on distribution and natural history of *Psammophis longifrons* Boulenger 1896) (Serpentes: Psammophiidae: Psammophiinae) in Gujarat, India. Russian Journal of Herpetology 20 (3): 217–222. https://doi.org/10.30906/1026-2296-2013-20-3-217-222
- Whitaker R, Captain A (2004) Snakes of India: the field guide. Draco Books, Chengelpat, India, 479 pp.
- Wogan GOU, Vindum JV, Wilkinson JA, Koo MS, Slowinski JB, Win H, Thin T, Kyi SW, Oo SL, Lwin KS, Shein AK (2008) New country records and range extensions for Myanmar amphibians and reptiles. Hamadryad 33: 83–96.
- Wüster W, Peppin L, Pook CE, Walker DE (2008) A nesting of vipers: phylogeny and historical biogeography of the Viperidae (Squamata: Serpentes). Molecular Phylogenetics and Evolution 49 (2) 445–459. https://doi.org/10.1016/j.ympev.2008.08.019
- Zaher H, Murphy RW, Arredondo JC, Graboski R, Machado-Filho PR, Mahlow K, Montingelli GG, Quadros AB, Orlov NL, Wilkinson M, Zhang YP (2019). Large-scale molecular phylogeny, morphology, divergence-time estimation, and the fossil record of advanced caenophidian snakes (Squamata: Serpentes). PloS ONE 14 (5): e0216148. https://doi.org/10.1371/journal.pone.0216148

## Supplemental Materials

**Table S1.** Species abbreviations, locality, latitude and longitude (in decimal minutes; datum WGS 84), and specimen ID of taxa used to map the overall geographic distribution of Psammophis species in Africa and Eurasia (Fig. 6) corresponds to their respective clade (Fig. 5).

**Table S2.** Uncorrected *p*-distance between cytB sequences of *Psammophis* species included in phylogenetic analysis.