



Mucor variicolumellatus L. Wagner & G. Walther (Mucorales, Mucoromycota): a first record for the Neotropics

Carlos Alberto Fragoso de Souza¹, Diogo Xavier Lima², Diogo Paes da Costa¹, Catarina Letícia Ferreira de Lima², Erika Valente de Medeiros¹, André Luiz Cabral Monteiro de Azevedo Santiago²

1 Programa de Pós-Graduação em Produção Agrícola, Universidade Federal Rural de Pernambuco, Av. Bom Pastor s/n, Boa Vista, Garanhuns, PE, CEP 55292-270, Brazil. **2** Programa de Pós-Graduação em Biologia de Fungos, Departamento de Micologia, Universidade Federal de Pernambuco, Av. Professor Nelson Chaves, s/n, CEP 50670-420, Recife, PE, Brazil.

Corresponding author: Carlos Alberto Fragoso de Souza, carlos_fragoso1@hotmail.com

Abstract

Two specimens of *Mucor variicolumellatus* L. Wagner & G. Walther were isolated from soil samples collected in an upland rainforest area located in Pernambuco state, Brazil. Their identity were confirmed by morphophysiology and ITS rDNA sequence analysis. Both specimens are distinguished from other species within the *Mucor circinelloides* complex by producing obovoid, ovate and strawberry-shaped columellae. A detailed description and illustration of the specimens are presented. This is the first record of *M. variicolumellatus* in the Neotropics.

Keywords

ITS rDNA; Mucoromyceta; taxonomy.

Academic editor: Roger Fagner Ribeiro Melo | Received 4 March 2020 | Accepted 28 May 2020 | Published 16 June 2020

Citation: De Souza CAF, Lima DX, da Costa DP, Lima CLF, de Medeiros EV, Santiago ALCMA (2020) *Mucor variicolumellatus* L. Wagner & G. Walther (Mucorales, Mucoromycota): a first record for the Neotropics. Check List 16 (3): 743–747. <https://doi.org/10.15560/16.3.743>

Introduction

Upland rainforest are disjunctions of Montane Seasonal Semideciduous Forest, which is one of the vegetation types belonging to the Brazilian Atlantic Forest domain (Freire et al. 2018). In northeastern Brazil, upland rainforest areas are commonly inserted in the semiarid region, surrounded by Caatinga vegetation. When compared to the ecosystems of Caatinga, the upland rainforest areas exhibit higher humidity and lower average temperatures (Medeiros and Cestaro 2019), thus favoring the high diversity of plants, animals and microorganisms, including fungi. However, the disordered using of the Brazilian upland rainforest areas has caused irreversible degradation processes, with the fragmentation and

loss of habitats and ecosystems, impacting on biological diversity and putting at risk this still poorly studied forests (Tabarelli and Santos 2004).

Despite of the important ecological role played by fungi in edaphic environments as saprobes, parasites and symbionts (Stamford et al. 2005), the species richness of fungi in the Brazilian upland rainforest areas is still underestimated (Flora do Brasil 2020). The distribution records of the order Mucorales Dumort. from these ecosystems are restricted to the contributions of Santiago et al. (2011, 2013), Crous et al. (2018), de Souza et al. (2018), and Lima et al. (2018), including three new recently described species of *Mucor* Fresen.

The genus *Mucor* is characterized by rapid mycelial growth and formation of non-apophysate sporangia

in simple or branched sporangiophores (Benny et al. 2014). Molecular studies have demonstrated that this genus is polyphyletic, and includes some species complexes, such as the *M. circinelloides* complex (MCC) (Pawłowska et al. 2013; Walther et al. 2013; Wagner et al. 2019). The MCC consists of 14 species, including the recently described *Mucor variicolumellatus* L. Wagner & G. Walther (Wagner et al. 2019).

Mucor variicolumellatus was previously reported by Zycha (1935) in a *Tremella* Dill. ex L. carpophore and it was mistakenly identified as *M. fragilis* Bainier. Wagner et al. (2019) described the new species based on morphophysiological and molecular data. Here we present a detailed description and illustration of *M. variicolumellatus* isolated from soil samples in an upland rainforest area of the semi-arid region of Brazil. This is the first record of *M. variicolumellatus* in the Neotropics.

Methods

Isolation, purification, and identification. Soil samples were collected in an upland rainforest area, in the city of Taquaritinga do Norte (Fig. 1), state of Pernambuco, Brazil. The samples were placed in clean plastic bags and stored in styrofoam boxes for transportation to the laboratory. For isolation, five milligrams of soil were added to Petri dishes containing wheat germ agar medium (Benny 2008) supplemented with chloramphenicol (100 mg/L) to prevent bacterial growth. The dishes were stored in the laboratory at room temperature (28 ± 2 °C) for seven days under alternate light and

dark periods. Fragments of mycelium were removed from the Petri dishes under a stereomicroscope (Leica EZ4) and transferred to Petri dishes with malt extract agar (MEA) (Benny 2008). The specimens were identified by comparing the macroscopic and microscopic characteristics as described by Schipper (1976), Álvarez et al. (2011), and Wagner et al. (2019). Lyophilized cultures of *M. variicolumellatus* are deposited in the culture collection Micoteca URM of the Federal University of Pernambuco.

Molecular analysis. Fungal biomass was obtained from MEA cultures in test tubes stored at (25 ± 2 °C) for up to six days. Genomic DNA was extracted as described by Oliveira et al. (2016). The primer pairs ITS1/ITS4 (White et al. 1990) were used to amplify the rDNA ITS1–5.8S–ITS2 region. The polymerase chain reaction was conducted as described by Oliveira et al. (2014). The final amplicons were purified with the Invitrogen Pure-Link PCR Purification Kit and sequenced. The newly obtained sequences were deposited in GenBank.

Phylogenetic reconstructions were obtained by analyzing the ITS1–5.8S–ITS2 sequence data. The sequences of *Mucor* spp. were obtained from GenBank and aligned with the sequences generated in the present study. The sequences were aligned with Clustal X (Larkin et al. 2007) and edited with BioEdit (Hall 1999). Prior to phylogenetic analysis, the optimal nucleotide substitution model was estimated using Topali 2.5 (Milne et al. 2004). Bayesian inference analysis (two runs over 1×10^6 generations with a burn-in of 2500) was performed using

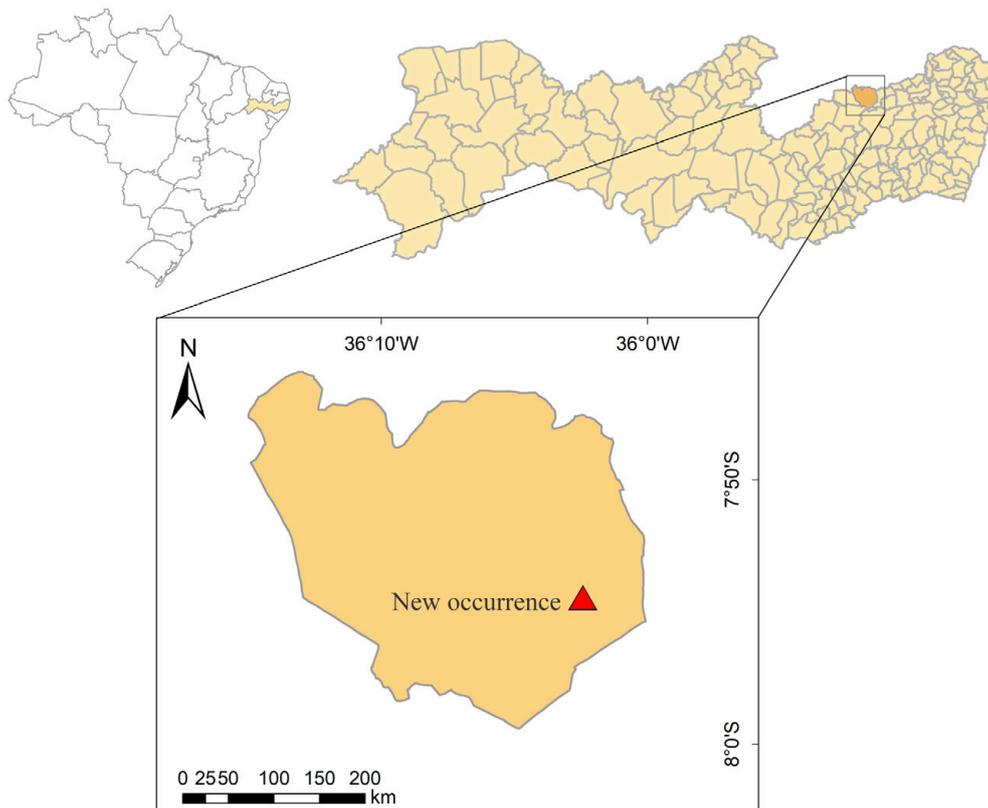


Figure 1. The location in Taquaritinga do Norte city, where *Mucor variicolumellatus* URM 7769 was found.

MrBayes 3.1.2, and maximum likelihood analysis (with support estimated by bootstrap analysis with 1000 replicates) was performed using PhyML (Guindon and Gascuel 2003), launched from Topali 2.5.

Results

Mucor variicolumellatus L. Wagner & G. Walther, *Persoonia* 44: 67–97. 2019.
Figure 2A–H

Material examined. BRAZIL• Pernambuco, Taquaritinga do Norte; 07°54'28"S, 036°02'25"W; 1.056 m a.s.l.; 26 Dec. 2017; Carlos Alberto Fragoso de Souza leg.; habitat: soil; URM 7769; GenBank: MT124621.

Additional material examined. BRAZIL• Pernambuco,

Taquaritinga do Norte; 07°54'28"S, 036°02'25"W; 1.056 m a.s.l.; 03 Apr. 2018, Carlos Alberto Fragoso de Souza leg.; habitat: soil; URM 7867; GenBank: MT124622.

Distribution. Brazil, Germany, Mawali, and USA.

Identification. Colonies firstly white then turning into pale gray, cottony, colonizing the entire Petri dish (9 cm diam) and touching the plate lid in five days, at 25°C, in MEA, reverse uncolored. Rhizoids present, slightly branched, up to 32 µm length, hyaline. Sporangiohores, hyaline to slightly pallid gray, 10–20 µm in diameter, repeatedly sympodially branched, with long or short branches, erect, some slightly curved, smooth-walled. Sporangia initially hyaline to pale orange then becoming brown gray to dark gray, globose, 20–90 µm in diameter, wall deliquescent. Columellae hyaline, often

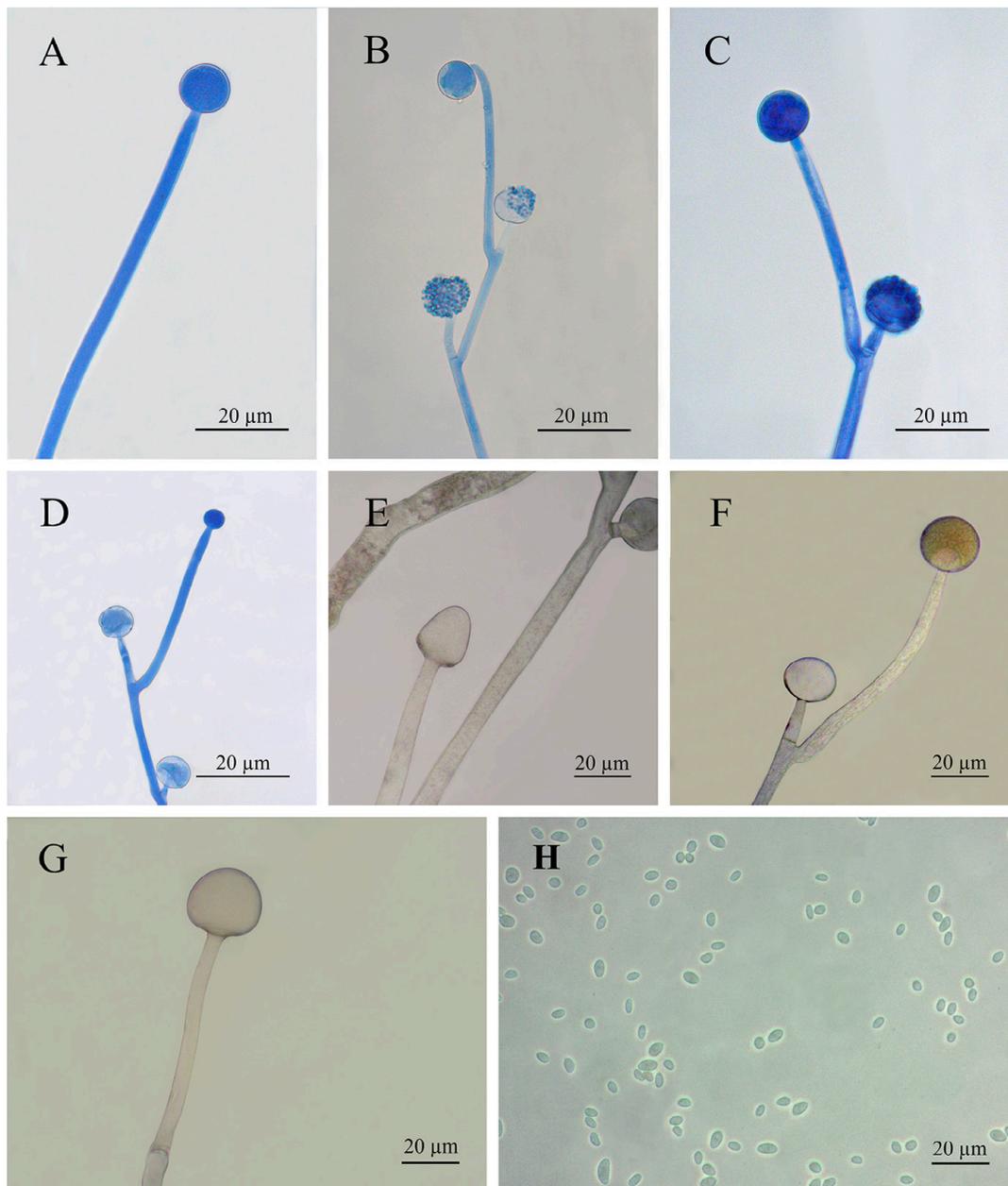


Figure 2. *Mucor variicolumellatus*. **A.** Unbranched sporangiohore with immature sporangium. **B, D.** Sympodially branched sporangiohore with globose sporangia and columella. **C.** Once branched sporangiohore with sporangia. **E.** Sporangiohore with conical columella. **F.** Once-branched sporangiohore with sporangium and columella. **G.** Unbranched sporangiohore with columella. **H.** Sporangiospores.

globose, 13–50 (–55) μm in diameter, obovoid, ovate and strawberry-shaped, 13.5–52 \times 13–45 (–60) μm , rarely ellipsoidal to cylindrical 22–33.5 \times 20–30, and conical 20–30 \times 13–25 μm , smooth-walled, with collar commonly observed. Sporangiospores hyaline, variable in size, ellipsoidal, (3–) 4–8 \times 2.5–4.5 μm , smooth-walled. Chlamydo-spores ellipsoidal to elongated ellipsoidal. Zygosporangia not observed.

Discussion

The *Mucor circinelloides* complex (MCC), as defined by Walther et al. (2013), currently comprises 14 recognized species, including *M. variicolumellatus*. According to Wagner et al. (2019), despite the morphological similarity between members of the MCC, all species can be distinguished by morphophysiological characteristics and phylogenetically, through molecular markers like ITS rDNA region. Our phylogenetic and morphological analyses support *M. variicolumellatus* as distinct from the other described MCC species.

Species of the MCC constitute a heterogeneous group of fungi, with oligotrophic or mesotrophic representatives adapted to a wide variety of substrates (Wagner et al. 2019). Although species of this complex are admittedly cosmopolitan, data on its geographic distribution are still quite fragmented, especially in Neotropical regions. *Mucor variicolumellatus* has been reported only in Germany, Malawi and the USA, isolated from *Tremella*, maize meal and humans (Álvarez et al. 2011; Wagner et al. 2019). Here we report the first record of *M. variicolumellatus* for the Neotropics, that was isolated from soil in an upland rainforest area of Pernambuco, Brazil.

Morphologically, *M. variicolumellatus* is distinguished from other MCC species by the production of obovoid, ovate and strawberry-shaped columellae. The morphological characteristics described here show close similarity to the original description given by Wagner et al. (2019) and Álvarez et al. (2011) (as *M. fragilis*), despite some features of our isolate, such as strawberry-shaped to conical and cylindrical columellae, were not reported by them. However, we do not consider these differences enough to characterize a new variety.

In the ITS rDNA phylogenetic tree (Fig. 3), *M. variicolumellatus* sequences grouped closely with those of *M. pseudolusitanicus* L. Wagner & G. Walther, that share morphological similarities. However, the sporangiospores of *M. variicolumellatus* do not show swellings at the base as frequently observed in *M. pseudolusitanicus*. Differences in sporangia diameters have also been observed, with *M. pseudolusitanicus* producing smaller sporangia (up to 75 μm). Additionally, *M. pseudolusitanicus* produces globose, semiglobose or occasionally slightly elongated but never obovoid columellae, different from those of *M. variicolumellatus*, which are obovoid, ovate and strawberry-shaped (Wagner et al. 2019).

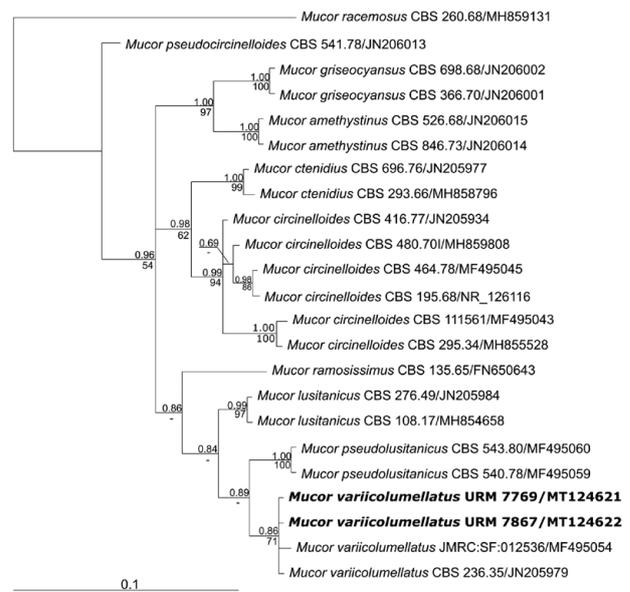


Figure 3. Phylogenetic tree of *Mucor variicolumellatus* and related species constructed using sequences of the ITS region. *Mucor racemosus* was used as outgroup. Sequences are labeled with their database accession numbers. Support values are from Bayesian inference and maximum likelihood analysis. The new Brazilian sequences are in boldface.

Our specimens, isolated as saprobe from soil, represent the first record in the Neotropics thereby expanding the knowledge of the geographical distribution of Mucoralean fungi.

Acknowledgements

This manuscript was financed by the project Diversity of Mucoromycotina in the different ecosystems of the Atlantic Rainforest of Pernambuco (FACEPE—First Projects Program PPP/FACEPE/CNPq-APQ-0842-2.12/14). The authors also thank the Coordination for the Improvement of Higher Education Personnel for the scholarships provided to CAFS and the National Council for Scientific and Technological Development for the research grant awarded to ALCMAS and EVM.

Authors' Contributions

DXL and CLFL collected the material; DPC performed the specified methodology; CAFS, ALCMAS and EVM wrote the text; ALCMAS and CAFS identified the species and built the plate.

References

- Álvarez E, Cano J, Stchigel AM, Sutton DA, Fothergill A, Salas A, Rinaldi MG, Guarro J (2011) Two new species of *Mucor* from clinical samples. *Medical Mycology* 49: 62–72. <https://doi.org/10.3109/13693786.2010.499521>
- Benny GL (2008) The methods used by Dr. R.K. Benjamin, and other Mycologists to isolate Zygomycetes. *Aliso* 26 (1): 37–61. <https://doi.org/10.5642/aliso.20082601.08>

- Benny GL, Humber RA, Voigt K (2014) Zygomycetous Fungi: Phylum Entomophthoromycota and Subphyla Kickxellomycotina, Mortierellomycotina, Mucoromycotina, and Zoopagomycotina. In: McLaughlin DJ, Spatafora JW (Eds) *The Mycota VII Part A, Systematics and evolution*. Springer-Verlag, Berlin, 209–250.
- Crous PW, Wingfield MJ, Burgess TI, Hardy GESTJ, Gené J, Guarro J, Baseia IG, García D, Gusmão LFP, Souza-Motta CM, Thangavel, Adamčík S, Barili A, Barnes CW, Bezerra JDP, Bordallo JJ, Cano-Lira JF, de Oliveira RJV, Ercole E, Hubka V, Iturrigata-González I, Kubátová A, Martín MP, Moreau P-A, Morte A, Ordoñez ME, Rodríguez A, Stchige AM, Vizzini A, Abdollahzadeh J, Abreu VP, Adamčíková K, Albuquerque GMR, Alexandrova AV, Duarte EÁ, Armstrong-Cho C, Banniza S, Barbosa RN, Bellanger J-M, Bezerra JL, Cabral TS, Caboň M, Caicedo E, Cantillo T, Carnegie AJ, Carmo LT, Castañeda-Ruiz RF, Clement CR, Čmoková A, Conceição LB, Cruz RHF, Damm U, da Silva BDB, da Silva GA, da Silva RMF, Santiago ALCMA, Oliveira LF, Souza CAF, Déniel F, Dima B, Dong G, Edwards J, Félix CR, Fournier J, Gibertoni TB, Hosaka K, Iturriaga T, Jadan M, Jany J-L, Jurjević Ž, Kolařík M, Kušan I, Landell MF, Cordeiro TRL, Lima DX, Loizides M, Luo S, Machado AR, Madrid H, Magalhães OMC, Marinho P, Matočec N, Mešić A, Miller AN, Morozova OV, Neves RP, Nonaka K, Nováková A, Oberlies NH, Oliveira-Filho JRC, Oliveira TGL, Papp V, Pereira OL, Perrone G, Peterson SW, Pham THG, Raja HA, Raudabaugh DB, Řehulka J, Rodríguez-Andrade E, Saba M, Schauflerová A, Shivas RG, Simonini G, Siqueira JPZ, Sousa JO, Stajsis V, Svetasheva T, Tan YP, Tkalčec Z, Ullah S, Valente P, Valenzuela-Lopez N, Abrinbana M, Viana Marques DA, Wong PTW, Xavier de Lima V, Groenewald JZ (2018) Fungal Planet description sheets: 716–784. *Persoonia* 40: 240–393. <https://doi.org/10.3767/persoonia.2018.40.10>
- de Souza CA, Voigt K, Gurgel LS, Cordeiro TR, Oliveira RJ, Lima DX, Santiago ALCMA (2018) A new species of *Mucor* (Mucoromycotina, Mucorales) isolated from an enclave of Upland Atlantic Forest in the semi-arid region of Brazil. *Phytotaxa* 351 (1): 53–62. <http://doi.org/10.11646/phytotaxa.351.1.4>
- Flora do Brasil (2020) <http://floradobrasil.jbrj.gov.br/>. Accessed on: 2020-2-26.
- Freire NCF, Moura DC, Silva JB, Moura ALSS, Melo JIM, Pacheco AP (2018) Atlas das caatingas: o único bioma exclusivamente brasileiro. Fundação Joaquim Nabuco, Massangana, Recife, 160–200 pp.
- Guindon S, Gascuel O (2003) A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. *Systematic Biology* 52: 696–704. <https://doi.org/10.1080/10635150390235520>
- Hall TA (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* 41: 95–98.
- Larkin M, Blackshields G, Brown NP, Chenna R, McGettigan PA, McWilliam H, Valentin F, Wallace IM, Wilm A, Lopez R, Thompson JD, Gibson TJ, Higgins DG (2007) Clustal W and Clustal X version 2.0. *Bioinformatics* 23: 2947–2948. <https://doi.org/10.1093/bioinformatics/btm404>
- Lima CLF, Lima DX, Souza CAF, Oliveira RJV, Cavalcanti IB, Gurgel LMS, Santiago ALCMA (2018) Description of *Mucor pernambucoensis* (Mucorales, Mucoromycota), a new species isolated from the Brazilian Upland Rainforest. *Phytotaxa* 350 (3): 274–282. <https://doi.org/10.11646/phytotaxa.350.3.6>
- Medeiros JF, Cestaro LA (2019) As diferentes abordagens utilizadas para definir brejos de altitude, áreas de exceção do nordeste brasileiro. *Sociedade e Território* 31 (2): 97–119. <https://doi.org/10.21680/2177-8396.2019v31n2ID16096>
- Milne I, Wright F, Rowe G, Marshal DF, Husmeier D, McGuire G (2004) TOPALi: software for automatic identification of recombinant sequences within DNA multiple alignments. *Bioinformatics* 20: 1806–1807. <https://doi.org/10.1093/bioinformatics/bth155>
- Oliveira RJV, Bezerra JL, Lima TEF, Silva GA, Cavalcanti MADQ (2016) *Phaeosphaeria nodulispora*, a new endophytic coelomycete isolated from tropical palm (*Cocos nucifera*) in Brazil. *Nova Hedwigia* 103 (1–2): 185–192. https://doi.org/10.1127/nova_hedwigia/2016/0343
- Oliveira RJV, Lima TEF, Cunha IB, Coimbra VRM, Silva GA, Bezerra JL, Cavalcanti MAQ (2014) *Corniculariella brasiliensis*, a new species of coelomycetes in the rhizosphere of *Caesalpinia echinata* (Fabaceae, Caesalpinioideae) in Brazil. *Phytotaxa* 178 (3): 197–204. <http://doi.org/10.11646/phytotaxa.178.3.5>
- Pawłowska J, Walther G, Wilk M, de Hoog S, Wrzosek M (2013) The use of compensatory base change analysis of ITS2 as a tool in the phylogeny of Mucorales, illustrated by the *Mucor circinelloides* complex. *Organisms Diversity & Evolution* 13: 497–502. <https://doi.org/10.1007/s13127-013-0139-1>
- Santiago ALCMA, Benny GL, Maia LC (2011) *Syncephalis aggregata*, a new species from the semiarid region of Brasil. *Mycologia* 103: 135–138. <https://doi.org/10.3852/10-049>
- Santiago ALCMA, Santos PJP, Maia LC (2013) Mucorales from the semiarid of Pernambuco. *Brazilian Journal of Microbiology* 44 (1): 299–305. <https://doi.org/10.1590/S1517-83822013005000027>
- Schipper MAA (1976) On *Mucor circinelloides*, *Mucor racemosus* and related species. *Studies in Mycology* 12: 1–40.
- Stamford NP, Rodrigues JJV, Heck RJ, Andrade DEGT (2005) Microbiota dos solos tropicais. In: Michereff SJ, Andrade DEGT, Menezes M (Eds) *Ecologia e Manejo de Patógenos Radiculares em Solos Tropicais*. Imprensa Universitária, Recife, 61–93.
- Tabarelli M, Santos AMM (2004) Uma breve descrição sobre a história natural dos brejos nordestinos. In: Porto KC, Cabral JJP, Tabarelli M (Eds) *Brejos de altitude em Pernambuco e Paraíba: história natural, ecologia e conservação*. Ministério do Meio Ambiente, Brasília, 299–302.
- Wagner L, Stielow JB, de Hoog GS, Bensch K, Schwartze VU, Voigt K, Alastruey-Izquierdo A, Kurzai O, Walther G (2019) A new species concept for the clinically relevant *Mucor circinelloides* complex. *Persoonia* 44: 67–97. <https://doi.org/10.3767/persoonia.2020.44.03>
- Walther G, Pawłowska J, Alastruey-Izquierdo A, Wrzosek M, Rodriguez-Tudela JL, Dolatabadi S, Chakrabarti A, de Hoog GS (2013) DNA barcoding in Mucorales: an inventory of biodiversity. *Persoonia* 30: 11–47. <https://doi.org/10.3767/003158513X665070>
- White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (Eds) *PCR protocols: a guide to methods and applications*. Academic Press, New York, 315–322.
- Zycha H (1935) Pilze II. Mucorineae. *Kryptogamenflora der Mark Brandenburg*. Gebrüder Bornträger, Leipzig, 264 pp.