



New taxonomic and faunistic records of fungus gnats (Insecta, Diptera) from Montenegro, Romania, and Serbia

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Abstract

A total of 95 fungus gnats species were identified from sweep net samples collected from Montenegro, Romania and Serbia during 2010–2017. A mycetophilid species, *Trichonta comis* Gagné, 1981, is redescribed and a key is provided for the separation of European *Boletina trivittata* (Meigen, 1818)-group species. Ten species from Montenegro, 27 from Romania and 1 species from Serbia are reported for the first time. The presence of *Stigmatomeria crassicornis* (Stannius, 1831) in Romania is verified.

Key words

Mycetophilidae; Keroplatidae; Dityomyiidae; Bolitophilidae; checklist; Europe.

Academic editor: Eliana Buenaventura | Received 4 October 2016 | Accepted 17 August 2017 | Published 29 September 2017

Citation: Kolcsár L-P (2017) New taxonomic and faunistic records of fungus gnats (Insecta, Diptera) from Montenegro, Romania, and Serbia. Check List 13 (5): 533–559. <https://doi.org/10.15560/13.5.533>

Introduction

The Sciaroidea is a species-rich superfamily, having 6 families occurring in the West Palearctic region: Dityomyiidae, Diadocidiidae, Keroplatidae, Bolitophilidae, Mycetophilidae (often collectively treated as fungus gnats), Sciariidae (black-winged fungus gnats), and Cecidomyiidae (gall midges) (Ševčík et al. 2016). In this paper Sciariidae and Cecidomyiidae are not discussed. More than 1365 species have been described from the Palearctic region (Bechev 2000) and new species are regularly found even from rather well-surveyed western Europe (Laštovka and Ševčík 2006, Kurina 2008). Larvae of European Sciaroidea are most often associated with microhabitats of humid forests, such as soil, dead wood, fruiting bodies of fungi and rarely plant tissues, eating mostly decomposing organic matter, while some

genera are predaceous. However, some species are only met in wetlands or in subterranean habitats (Søli et al. 2000, Jakovlev et al. 2014).

The first Romanian Sciaroidea records were published in the mid-19th century as a part of a larger monograph by Winnertz (1863). Later material collected from Romania was treated in the works of Strobl (1896), Thalhammer (1899), Lundström (1911a, 1911b, 1912, 1913, 1916), and Landrock (1914, 1925). Additions to the Romanian list of species were also provided by Tollet (1955), Decu-Burghel (1963), Burghel-Balacesco (1965, 1967, 1968), and Matile and Burghel-Balacesco (1969), based mostly on data about species found in caves. An additional publication added new information on the occurrence of Mycetophilidae in Romania (Sóos and Papp 1988). The expansion of knowledge on the Romanian fungus gnats fauna continued in the early 2000s during surveys

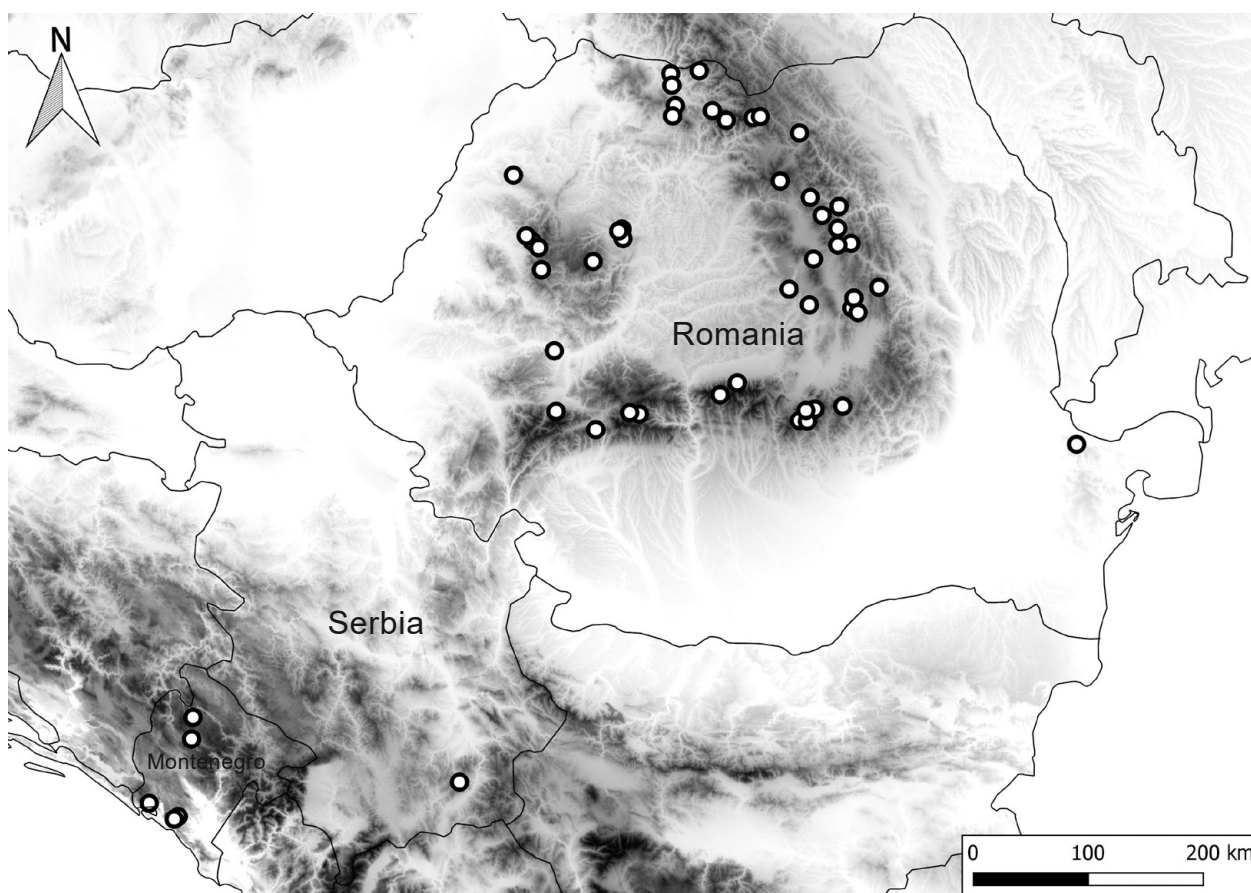


Figure 1. Location of collection sites in Montenegro, Romania, and Serbia of fungus gnats.

of Carpathians and corridor of Danube (Pârnu 2005). This Carpathian survey was limited to the territory of the Maramureş Basin (Pârnu 2002, 2003) and Piatra Craiului National Park (Pârnu 2004a, 2004b). In the *Checklist of Romanian Fauna* (Moldovan et al. 2007), 205 species of fungus gnats (Ditomyiidae, Diadocidiidae, Keroplatidae, Bolitophilidae, Mycetophilidae) are listed though without any references and some previously reported species seem to have been missed out. The online resource *Fauna Europaea* (Chandler 2013) states that 206 species of Sciaroidea (except Sciaridae and Cecidomyiidae) are known from Romania. Both above mentioned checklists are deficient, not including all species reported previously from Romania. In the present paper we suggest that a total of 270 fungus gnats species are known from the country, previous to this paper and even this number should be regarded as an underestimate due to poor faunistic knowledge.

The Sciaroidea of Montenegro and Serbia has been much less investigated and no national checklist has hitherto been published. The last published checklist under the former Yugoslavia (focusing upon Serbia and Montenegro) by Bechev (1997) listed 56 fungus gnats species. *Fauna Europaea* (Chandler 2013) lists 166 sciaroid species (excluding Sciaridae and Cecidomyiidae), but this list combines the fauna of Montenegro and Serbia jointly. We suggest that a total of 168 fungus gnats species are known from these countries, previous to this paper.

The first author (L-PK) and his colleagues had collected insects from Montenegro, Romania, and Serbia during the years 2010–2017. Fungus gnats were sorted from the collected material and were sent to the second author (JS) for identification. Thus, the present paper aims to improve the faunistic knowledge of fungus gnats in the countries in question through providing an updated checklist of fungus gnats (Ditomyiidae, Diadocidiidae, Keroplatidae, Bolitophilidae, Mycetophilidae) recorded from Montenegro, Romania, and Serbia jointly.

Methods

The material was collected by sweep netting in Montenegro, Romania and Serbia (Fig. 1, Table 1), between 2010 and 2017, and identified by Jukka Salmela. All the material listed here, are stored in 96% ethanol and deposited in the natural history collection of the Regional Museum of Lapland, Rovaniemi, Finland (LMM). Individual catalogue numbers of the vouchers are given (e.g., DIPT-JS-2016-0301). The holotype of *Trichonta comis* Gagné, 1981 was loaned from Finnish Museum of Natural History (Zoological Museum) (MZHF), University of Helsinki, Helsinki, Finland. The arrangement of the treated families and genera follows Bechev (2000) and *Fungus Gnats Online* (<http://sciaroidea.info/>) that may better reflect the phylogeny of the families and sub-families than *Fauna Europaea* (Chandler 2013), which

Table 1. Collection data in Montenegro (ME), Romania (RO) and Serbia (SRB), between 2010 and 2017.

Species	Specimen	Date	Location	Lat. (° N)	Long. (° E)	Collector(s)
<i>Allodia (Brachycampta) barbata</i> (Lundstrom, 1909)	1♂	28-V-2014	RO, Sacele, Baiul Mts, Bratocea Pass, 1177 m	45.4790	025.8936	Kolcsár L.-P.
<i>Allodia (Brachycampta) grata</i> (Meigen, 1830)	1♂	23-VIII-2014	RO, Balan, Hasmas Mts, Galkut valley, small brook, 1050 m	46.6493	025.8415	Kolcsár L.-P.
<i>Allodia (Brachycampta) grata</i> (Meigen, 1830)	1♂	25-VIII-2014	RO, Casinu Nou, Bodoc Mt., Balaj pass, brook, 880 m	46.1949	025.9887	Kolcsár L.-P.
<i>Allodia (Allodia) lugens</i> (Wiedemann, 1817)	1♂	22-V-2014	RO, Remeti, Bihor Mts, Iad River, 870 m	46.7157	022.5788	Kolcsár L.-P.
<i>Allodia (Allodia) lugens</i> (Wiedemann, 1817)	1♂	28-V-2014	RO, Sacele, Baiul Mts, Bratocea Pass, 1177 m	45.4790	025.8936	Kolcsár L.-P.
<i>Allodia (Allodia) lundstroemi</i> Edwards, 1921	1♂	25-IX-2014	RO, Vanvucesti, Vartop Mt., Varciorog waterfall, 1000 m	46.4714	022.7376	Kolcsár L.-P.
<i>Allodia (Allodia) ornaticollis</i> (Meigen, 1818)	1♂	25-VIII-2014	RO, Casinu Nou, Bodoc Mt., Balaj pass, brook, 880 m	46.1949	025.9887	Kolcsár L.-P.
<i>Allodiopsis rustica</i> (Edwards, 1941)	1♂	25-VIII-2014	RO, Sanmartin, Ciuc Mts, Rugat Valley, 950 m	46.2669	026.0103	Kolcsár L.-P.
<i>Anatella ciliata</i> Winnertz, 1864	1♂	30-IX-2014	RO, Cluj-Napoca, Miko Garden, 373 m	46.7635	023.5804	Kolcsár L.-P.
<i>Antlemon (Antlemonopsis) brevisanum</i> (Loew, 1871)	1♂	26-VIII-2014	RO, Azuga, Baiul Mts, Limbasel Valley, 1250 m	45.4578	025.6015	Kolcsár L.-P.
<i>Boletina basalis</i> (Meigen, 1818)	1♂	16-V-2014	RO, Cluj-Napoca, Cartierul Manastur, 434 m	46.7497	023.5467	Kolcsár L.-P.
<i>Boletina basalis</i> (Meigen, 1818)	1♂	18-V-2014	RO, Botiza, Tibles Mts, Sasul River, springs, 905 m	47.6411	024.1400	Kolcsár L.-P.
<i>Boletina cincticornis</i> (Walker, 1848)	1♂	27-V-2014	RO, Capatanenii Ungureni, Fagaras Mts, Transfagaras, 1280 m	45.5626	024.6093	Kolcsár L.-P.
<i>Boletina cincticornis</i> (Walker, 1848)	1♂	28-V-2014	RO, Azuga, Baiul Mts, Limbasel Valley, 1250 m	45.4578	025.6015	Kolcsár L.-P.
<i>Boletina cincticornis</i> (Walker, 1848)	2♂	6-VIII-2014	RO, Muntele Baisorii, Bihor Mts, Buscat Mt., 1634 m	46.5310	023.2788	Kolcsár L.-P.
<i>Boletina gripha</i> Dziedzicki, 1885	2♂	17-V-2013	RO, Rona de Sus, Maramures Hills, 425 m	47.8664	024.0912	Kolcsár L.-P.
<i>Boletina joosti</i> Plassmann, 1987	1♂	21-VIII-2014	RO, Gura Haitii, Caliman Mts, near to mine, 1640 m	47.1071	025.2385	Kolcsár L.-P.
<i>Boletina sciarina</i> Staeger, 1840	1♂	22-VIII-2014	RO, Pintec, Giurgiu Mts, Pintec valley, 820 m	46.9255	025.8533	Kolcsár L.-P.
<i>Boletina trivittata</i> (Meigen, 1818)	1♂	11-V-2010	ME, Zabljak, Durmitor Mt., Crna lake, 1448 m	43.1486	019.0882	Kolcsár L.-P.
<i>Boletina trivittata</i> (Meigen, 1818)	1♂	17-V-2014	RO, Moisei, Rodnei Mts, Izei River, springs, 1014 m	47.6065	024.5274	Kolcsár L.-P.
<i>Boletina trivittata</i> (Meigen, 1818)	1♂	25-V-2014	RO, Carnic, Retezat Mts, Pietrele Valley, 978 m	45.4405	022.8913	Kolcsár L.-P.
<i>Boletina trivittata</i> (Meigen, 1818)	2♂	26-V-2014	RO, Oborsia Lotrului, Parang Mts, 1450 m	45.4319	023.6618	Kolcsár L.-P.
<i>Boletina trivittata</i> (Meigen, 1818)	1♂	28-V-2014	RO, Azuga, Baiul Mts, Limbasel Valley, 1205 m	45.4578	025.6015	Kolcsár L.-P.
<i>Boletina trivittata</i> (Meigen, 1818)	1♂	19-VIII-2014	RO, Anies, Rodnei Mts, Cepelor spring, 1227 m	47.5380	024.6728	Kolcsár L.-P.
<i>Bolitophila (Bolitophila) austriaca</i> (Mayer, 1950)	1♂	11-V-2010	ME, Zabljak, Durmitor Mt., Crna lake, 1448 m	43.1486	019.0882	Kolcsár L.-P.
<i>Bolitophila (Bolitophila) austriaca</i> (Mayer, 1950)	1♂	26-V-2014	RO, Voineasa, Parangului Mts, Vidra lake, valley Mogosu, 1320 m	45.4212	023.7621	Kolcsár L.-P.
<i>Bolitophila (Bolitophila) cinerea</i> Meigen, 1818	1♂	5-V-2014	RO, Varghis, Varghis Canyon, 630 m	46.2174	025.5435	Török E.
<i>Bolitophila (Bolitophila) cinerea</i> Meigen, 1818	1♂	18-V-2014	RO, Botiza, Tibles Mts, Sasul River, springs, 905 m	47.6411	024.1400	Kolcsár L.-P.
<i>Bolitophila (Cliopisa) melanoleuci</i> Polevoi, 1996	1♂	13-V-2010	ME, Herceg Novi, Kotori bay, Morinj, 285 m	42.4894	018.6290	Kolcsár L.-P.
<i>Bolitophila (Cliopisa) modesta</i> Lackschewitz, 1937	1♂	1-V-2010	RO, Bradesti, Tarnava Mare River, 490 m	46.3314	025.3311	Kolcsár L.-P.
<i>Bolitophila (Cliopisa) occlusa</i> Edwards, 1913	1♂	22-V-2014	RO, Stana de Vale, Bihor Mts, Ciripa Valley, 1375 m	46.6756	022.6412	Kolcsár L.-P.
<i>Bolitophila (Bolitophila) saundersi</i> (Curtis, 1836)	1♂	3-V-2014	ME, Cetinje, Lovcen NP, Bjelosi, 950 m	42.3670	018.8911	Kolcsár L.-P.
<i>Bolitophila (Bolitophila) saundersi</i> (Curtis, 1836)	1♂	17-VII-2013	RO, Trei Fantani, Hasmas Mts, Bicaz-Mic brook, 900 m	46.7690	025.8412	Kolcsár L.-P.
<i>Bolitophila (Bolitophila) tenella</i> Winnertz, 1864	1♂	1-V-2010	RO, Bradesti, Tarnava Mare River, 490 m	46.3314	025.3311	Kolcsár L.-P.
<i>Bolitophila (Bolitophila) tenella</i> Winnertz, 1864	1♂	25-V-2014	RO, Carnic, Retezat Mts, Pietrele Valley, 978 m	45.4405	022.8913	Kolcsár L.-P.
<i>Bolitophila (Bolitophila) tenella</i> Winnertz, 1864	1♂	26-V-2014	RO, Oborsia Lotrului, Parang Mts, 1450 m	45.4319	023.6618	Kolcsár L.-P.
<i>Bolitophila (Bolitophila) tenella</i> Winnertz, 1864	1♂	26-V-2014	RO, Voineasa, Parangului Mts, Vidra lake, valley Mogosu, 1320 m	45.4212	023.7621	Kolcsár L.-P.
<i>Bolitophila (Bolitophila) tenella</i> Winnertz, 1864	1♂	25-VIII-2014	RO, Casinu Nou, Bodoc Mt., Balaj pass, brook, 880 m	46.1949	025.9887	Kolcsár L.-P.

Continued

Table 1. Continued.

Species	Specimen	Date	Location	Lat. (° N)	Long. (° E)	Collector(s)
<i>Brevicornu sericoma</i> (Meigen, 1830)	1♂	11-V-2010	ME, Zabljak, Durmitor Mt., Crna lake, 1448 m	43.1486	019.0882	Kolcsár L.-P.
<i>Coelophthiria thoracica</i> (Winnertz, 1864)	2♂	26-VIII-2014	RO, Azuga, Baiul Mts, Limbasel Valley, 1250 m	45.4578	025.6015	Kolcsár L.-P.
<i>Coelosia fusca</i> Bezzi, 1892	1♂	11-V-2010	ME, Zabljak, Durmitor Mt., Crna lake, 1448 m	43.1486	019.0882	Kolcsár L.-P.
<i>Cerotelion striatum</i> (Gmelin, 1790)	1♂	30-VI-2017	SRB, Sikirje, Kukavica Mts, 648 m	42.6525	021.8801	Kolcsár L.-P., Török E.
<i>Diadocidia (Diadocidia) spinosula</i> Tollet, 1948	1♂	26-V-2014	RO, Oborsia Lotrului, Parang Mts, 1450 m	45.4319	023.6618	Kolcsár L.-P.
<i>Ditomyia fasciata</i> (Meigen, 1818)	1♀	16-V-2014	RO, Cluj-Napoca, Cartierul Manastur, 434 m	46.7497	023.5467	Kolcsár L.-P.
<i>Docosia expectata</i> Lastovka & Sevcik, 2006	1♂	18-V-2014	RO, Grosii Tiblesului, Tibles Mts, Minghet Valley, 777 m	47.5685	024.1117	Kolcsár L.-P.
<i>Docosia flavicoxa</i> Strobl, 1900	1♂	25-V-2014	RO, Salatruc, Ciuc Mts, Uz Valley, 645 m	46.3452	026.2715	Kolcsár L.-P.
<i>Docosia gilvipes</i> (Walker, 1856)	1♂	11-V-2010	ME, Zabljak, Durmitor Mt., Crna lake, 1448 m	43.1486	019.0882	Kolcsár L.-P.
<i>Exechia fusca</i> (Meigen, 1804)	1♂	25-V-2014	RO, Carnic, Retezat Mts, Pietrele Valley, 978 m	45.4405	022.8913	Kolcsár L.-P.
<i>Exechia fusca</i> (Meigen, 1804)	1♂	22-VIII-2014	RO, Pintec, Giurgiu Mts, Pintec valley, 820 m	46.9255	025.8533	Kolcsár L.-P.
<i>Exechia pseudocincta</i> Strobl, 1910	1♂	23-VIII-2014	RO, Baratos, Hasmas Mts, Muhos valley, 1111 m	46.6608	025.9786	Kolcsár L.-P.
<i>Exechia seriata</i> (Meigen, 1830)	1♂	25-IX-2014	RO, Vanvucesti, Vartop Mt., Varciorog waterfall, 1000 m	46.4714	022.7376	Kolcsár L.-P.
<i>Exechiopsis (Exechiopsis) magnicauda</i> (Lundstrom, 1911)	2♂	23-VIII-2014	RO, Balan, Hasmas Mts, Galkut valley, small brook, 1050 m	46.6493	025.8415	Kolcsár L.-P.
<i>Exechiopsis (Exechiopsis) subulata</i> (Winnertz, 1864)	1♂	23-VIII-2014	RO, Balan, Hasmas Mts, Galkut valley, small brook, 1050 m	46.6493	025.8415	Kolcsár L.-P.
<i>Exechiopsis (Exechiopsis) subulata</i> (Winnertz, 1864)	1♂	25-IX-2014	RO, Vanvucesti, Vartop Mt., Varciorog waterfall, 1000 m	46.4714	022.7376	Kolcsár L.-P.
<i>Exechiopsis (Exechiopsis) unguiculata</i> (Lundstrom, 1911)	1♂	11-V-2010	ME, Zabljak, Durmitor Mt., Crna lake, 1448 m	43.1486	019.0882	Kolcsár L.-P.
<i>Exechiopsis (Exechiopsis) unguiculata</i> (Lundstrom, 1911)	1♂	11-V-2010	ME, Zabljak, Durmitor Mt., Crna lake, 1448 m	43.1486	019.0882	Kolcsár L.-P.
<i>Exechiopsis (Exechiopsis) unguiculata</i> (Lundstrom, 1911)	1♂	29-V-2014	RO, Sambata de Sus, Fagaras Mts, Sambata Valley, 1020 m	45.6508	024.7889	Kolcsár L.-P.
<i>Grzegorzekia collaris</i> (Meigen, 1818)	1♂	16-V-2014	RO, Cluj-Napoca, Cartierul Manastur, 434 m	46.7497	023.5467	Kolcsár L.-P.
<i>Keroplatus dispar</i> (Dufour, 1839)	1♂	30-IX-2014	RO, Cluj-Napoca, Miko Garden, 373 m	46.7635	023.5804	Kolcsár L.-P.
<i>Keroplatus reaumurii reaumurii</i> Dufour, 1839	1♂	25-IX-2014	RO, Vanvucesti, Vartop Mt., Varciorog waterfall, 1000 m	46.4714	022.7376	Kolcsár L.-P.
<i>Macrocera centralis</i> Meigen, 1818	2♂	30-III-2014	ME, Cetinje, 670 m	42.3873	018.9330	Kolcsár L.-P.
<i>Macrocera longibrachia</i> Landrock, 1917	1♂	22-V-2014	RO, Stana de Vale, Bihor Mts, Ciripa Valley, 1375 m	46.6756	022.6412	Keresztes L.
<i>Macrocera pilosa</i> Landrock, 1917	1♂	5-V-2014	RO, Varghis, Varghis Canyon, 630 m	46.2174	025.5435	Török E.
<i>Macrocera vittata</i> Meigen, 1830	1♂	30-IV-2014	ME, Savnik, Petnjica, 1057 m	42.9836	019.0733	Kolcsár L.-P.
<i>Macrorrhyncha rostrata</i> (Zetterstedt, 1851)	2♂	26-VII-2014	RO, Dobresti, Bucegi Mts, Saua Dichiului, 1575 m	45.3736	025.4391	Kolcsár L.-P.
<i>Macrorrhyncha rostrata</i> (Zetterstedt, 1851)	1♂	30-IX-2014	RO, Cluj-Napoca, Miko Garden, 373 m	46.7635	023.5804	Kolcsár L.-P.
<i>Monocentrotia matilei</i> Bechev, 1989	1♂	5-VIII-2012	RO, Padurea Neagra, Plopi Mt., Bistra brook, 441 m	47.1492	022.4441	Török E.
<i>Monoclona rufilatera</i> (Walker, 1836)	1♂	28-V-2014	RO, Sacele, Baiul Mts, Bratocea Pass, 1177 m	45.4790	025.8936	Kolcsár L.-P.
<i>Mycetophila abbreviata</i> Landrock, 1914	1♂	22-V-2014	RO, Remeti, Bihor Mts, Iad River, 870 m	46.7157	022.5788	Kolcsár L.-P.
<i>Mycetophila abbreviata</i> Landrock, 1914	1♂	25-IX-2014	RO, Vanvucesti, Vartop Mt., Varciorog waterfall, 1000 m	46.4714	022.7376	Kolcsár L.-P.
<i>Mycetophila alea</i> Laffoon, 1965	1♂	11-V-2010	ME, Zabljak, Durmitor Mt., Crna lake, 1448 m	43.1486	019.0882	Kolcsár L.-P.
<i>Mycetophila curviseta</i> Lundstrom, 1911	1♂	23-VIII-2014	RO, Balan, Hasmas Mts, Galkut valley, small brook, 1050 m	46.6493	025.8415	Kolcsár L.-P.
<i>Mycetophila curviseta</i> Lundstrom, 1911	1♂	25-VIII-2014	RO, Casinu Nou, Bodoc Mt., Balaj pass, brook, 880 m	46.1949	025.9887	Kolcsár L.-P.
<i>Mycetophila deflexa</i> Chandler, 2001	2♂	25-VIII-2014	RO, Sanmartin, Ciuc Mts, Rugat Valley, 950 m	46.2669	026.0103	Kolcsár L.-P.
<i>Mycetophila dentata</i> Lundstrom, 1913	1♂	22-VIII-2014	RO, Pintec, Giurgiu Mts, Pintec valley, 820 m	46.9255	025.8533	Kolcsár L.-P.
<i>Mycetophila edwardsi</i> Lundstrom, 1913	1♂	25-V-2014	RO, Salatruc, Ciuc Mts, Uz Valley, 645 m	46.3452	026.2715	Kolcsár L.-P.

Continued

Table 1. Continued.

Species	Spec-imen	Date	Location	Lat. (° N)	Long. (° E)	Collector(s)
<i>Mycetophila fraterna</i> Winnertz, 1864	1♂	6-VIII-2014	RO, Padis, Bihor Mts, Cetatile Radesei Cave (around), 1294 m	46.6301	022.7081	Kolcsár L.-P.
<i>Mycetophila fraterna</i> Winnertz, 1864	1♂	20-VIII-2014	RO, Gura Lalei, Rodnei Mts, Lalei Valley, 1182 m	47.5560	024.9549	Kolcsár L.-P.
<i>Mycetophila fraterna</i> Winnertz, 1864	1♂	22-VIII-2014	RO, Pintec, Giurgiu Mts, Pintec valley, 820 m	46.9255	025.8533	Kolcsár L.-P.
<i>Mycetophila lamellata</i> Lundstrom, 1911	1♂	22-V-2014	RO, Remeti, Bihor Mts, Iad River, 870 m	46.7157	022.5788	Kolcsár L.-P.
<i>Mycetophila lastovkai</i> Caspers, 1984	1♂	22-V-2014	RO, Remeti, Bihor Mts, Iad River, 870 m	46.7157	022.5788	Kolcsár L.-P.
<i>Mycetophila lastovkai</i> Caspers, 1984	1♂	25-V-2014	RO, Carnic, Retezat Mts, Pietrele Valley, 978 m	45.4405	022.8913	Kolcsár L.-P.
<i>Mycetophila lastovkai</i> Caspers, 1984	1♂	26-V-2014	RO, Vulcan, Vulcan Mt., Vulcan Pass, 1420 m	45.3067	023.3064	Kolcsár L.-P.
<i>Mycetophila lastovkai</i> Caspers, 1984	1♂	25-IX-2014	RO, Vanvucesti, Vartop Mt., Varciorog waterfall, 1000 m	46.4714	022.7376	Kolcsár L.-P.
<i>Mycetophila marginata</i> Winnertz, 1864	1♂	11-V-2010	ME, Zabljak, Durmitor Mt., Crna lake, 1448 m	43.1486	019.0882	Kolcsár L.-P.
<i>Mycetophila ocellus</i> Walker, 1848	1♂	6-VIII-2014	RO, Padis, Bihor Mts, Cetatile Radesei Cave (around), 1294 m	46.6301	022.7081	Kolcsár L.-P.
<i>Mycetophila signatoides</i> Dziedzicki, 1884	1♂	22-VIII-2014	RO, Pintec, Giurgiu Mts, Pintec valley, 820 m	46.9255	025.8533	Kolcsár L.-P.
<i>Mycetophila stylata</i> (Dziedzicki, 1884)	1♂	22-VIII-2014	RO, Pintec, Giurgiu Mts, Pintec valley, 820 m	46.9255	025.8533	Kolcsár L.-P.
<i>Mycetophila trinitata</i> Staeger, 1840	1♂	25-V-2014	RO, Salatruc, Ciuc Mts, Uz Valley, 645 m	46.3452	026.2715	Kolcsár L.-P.
<i>Mycomya (Mycomya) alpina</i> Matile, 1972	1♂	21-VIII-2014	RO, Borsec, Giurgeu Mts, pine forest, 1125 m	46.9901	025.5534	Kolcsár L.-P.
<i>Mycomya (Mycomya) alpina</i> Matile, 1972	1♂	26-VIII-2014	RO, Azuga, Baiul Mts, Limbasel Valley, 1250 m	45.4578	025.6015	Kolcsár L.-P.
<i>Mycomya (Mycomya) alpina</i> Matile, 1972	1♂	26-VIII-2014	RO, Azuga, Baiul Mts, Limbasel Valley, 1250 m	45.4578	025.6015	Kolcsár L.-P.
<i>Mycomya (Mycomya) cinerascens</i> (Macquart, 1826)	1♂	22-VIII-2014	RO, Pintec, Giurgiu Mts, Pintec valley, 820 m	46.9255	025.8533	Kolcsár L.-P.
<i>Mycomya (Mycomya) egregia</i> (Dziedzicki, 1885)	1♂	19-VIII-2014	RO, Anies, Rodnei Mts, Cepelor spring, 1227 m	47.538	024.6728	Kolcsár L.-P.
<i>Mycomya (Mycomya) marginata</i> (Meigen, 1818)	1♂	18-V-2014	RO, Slatioara, Izei Valley, Slatioara brook, 345 m	47.7846	024.1035	Kolcsár L.-P.
<i>Mycomya (Mycomya) marginata</i> (Meigen, 1818)	1♂	26-VIII-2014	RO, Azuga, Baiul Mts, Limbasel Valley, 1250 m	45.4578	025.6015	Kolcsár L.-P.
<i>Mycomya (Mycomya) neohyalinata</i> Vaisanen, 1984	1♂	28-V-2014	RO, Azuga, Baiul Mts, Limbasel Valley, 1250 m	45.4578	025.6015	Kolcsár L.-P.
<i>Mycomya (Mycomya) neohyalinata</i> Vaisanen, 1984	1♂	20-VIII-2014	RO, Gura Lalai, Maramures Mts, Bistrita River small spring, 1025 m	47.5644	025.0301	Kolcsár L.-P.
<i>Mycomya (Mycomya) prominens</i> (Lundstrom, 1913)	1♂	22-VIII-2014	RO, Pintec, Giurgiu Mts, Pintec valley, 820 m	46.9255	025.8533	Kolcsár L.-P.
<i>Mycomya (Mycomya) tenuis</i> (Walker, 1856)	1♂	19-VII-2013	RO, Valea Putnei, Giumalau Mt., Padurea Seculara Giumalau, 1111 m	47.4483	025.4408	Kolcsár L.-P.
<i>Mycomya (Mycomyopsis) trilineata</i> (Zetterstedt, 1838)	1♂	30-IV-2014	ME, Savnik, Petnjica, 1057 m	42.9836	019.0733	Kolcsár L.-P.
<i>Mycomya (Mycomya) tumida</i> (Winnertz, 1864)	1♂	30-IX-2014	RO, Cluj-Napoca, Miko Garden, 373 m	46.7635	023.5804	Kolcsár L.-P.
<i>Mycomya (Mycomya) vittiventris</i> (Zetterstedt, 1852)	1♂	21-VIII-2014	RO, Gura Haitii, Caliman Mts, near to mine, 1640 m	47.1071	025.2385	Kolcsár L.-P.
<i>Mycomya (Mycomya) winnertzi</i> (Dziedzicki, 1885)	1♂	3-V-2014	ME, Cetinje, Lovcen NP, Bjelosi, 950 m	42.3670	018.8911	Kolcsár L.-P.
<i>Mycomya (Mycomya) winnertzi</i> (Dziedzicki, 1885)	2♂	23-VIII-2014	RO, Balan, Hasmas Mts, Galkut valley, small brook, 1050 m	46.6493	025.8415	Kolcsár L.-P.
<i>Neoempheria pictipennis</i> (Haliday, 1833)	1♂	24-V-2014	RO, Salatruc, Nemira Mts, Uz Valley, 480 m	46.3452	026.2715	Kolcsár L.-P.
<i>Neuratelia nemoralis</i> (Meigen, 1818)	1♂	18-V-2014	RO, Botiza, Tibles Mts, Sasul River, springs, 905 m	47.6411	024.1400	Kolcsár L.-P.
<i>Neuratelia nemoralis</i> (Meigen, 1818)	1♂	25-V-2014	RO, Carnic, Retezat Mts, Pietrele Valley, 978 m	45.4405	022.8913	Kolcsár L.-P.
<i>Neuratelia nemoralis</i> (Meigen, 1818)	1♂	6-VIII-2014	RO, Padis, Bihor Mts, Cetatile Radesei Cave (around), 1294 m	46.6301	022.7081	Kolcsár L.-P.

Continued

Table 1. Continued.

Species	Specimen	Date	Location	Lat. (° N)	Long. (° E)	Collector(s)
<i>Notolopha cristata</i> (Staeger, 1840)	1♂	25-VIII-2014	RO, Catrusa, Bodoc Mt., Fagus forest, 740 m	46.1620	026.0534	Kolcsár L.-P.
<i>Orfelia fasciata</i> (Meigen, 1804)	1♂	16-V-2014	RO, Cluj-Napoca, Cartierul Manastur, 434 m	46.7497	023.5467	Kolcsár L.-P.
<i>Orfelia fasciata</i> (Meigen, 1804)	1♂	24-V-2014	RO, Deva, Cetatea Valley, 487 m	45.8842	022.8725	Kolcsár L.-P.
<i>Orfelia fasciata</i> (Meigen, 1804)	1♂	27-XII-2014	RO, Hagota, Giurgeu Mts, Tisasul brook, 860 m	46.8618	025.6772	Kolcsár L.-P.
<i>Phronia forcipula</i> Winnertz, 1864	1♂	3-V-2014	ME, Cetinje, Lovcen NP, Bjelosi, 950 m	42.3670	018.8911	Kolcsár L.-P.
<i>Phronia forcipula</i> Winnertz, 1864	1♂	11-V-2010	ME, Zabljak, Durmitor Mt., Crna lake, 1448 m	43.1486	019.0882	Kolcsár L.-P.
<i>Phronia forcipula</i> Winnertz, 1864	1♂	26-V-2014	RO, Vulcan, Vulcan Mt., Vulcan Pass, 1420 m	45.3067	023.3064	Kolcsár L.-P.
<i>Phronia forcipula</i> Winnertz, 1864	1♂	27-VII-2013	RO, Liban, Harghita Mts, Lacul Dracului bog, 1175 m	46.5481	025.5870	Kolcsár L.-P.
<i>Phronia forcipula</i> Winnertz, 1864	1♂	21-VIII-2014	RO, Borsec, Giurgeu Mts, pine forest, 1125 m	46.9901	025.5534	Kolcsár L.-P.
<i>Phronia forcipula</i> Winnertz, 1864	1♂	23-VIII-2014	RO, Balan, Hasmas Mts, Galkut valley, small brook, 1050 m	46.6493	025.8415	Kolcsár L.-P.
<i>Phronia humeralis</i> Winnertz, 1864	1♂	22-VIII-2014	RO, Pintec, Giurgiu Mts, Pintec valley, 820 m	46.9255	025.8533	Kolcsár L.-P.
<i>Phronia nigricornis</i> (Zetterstedt, 1852)	2♂	25-V-2014	RO, Salatruc, Ciuc Mts, Uz Valley, 645 m	46.3452	026.2715	Kolcsár L.-P.
<i>Phthiria humilis</i> Winnertz, 1864	1♂	27-V-2014	RO, Capatanenii Ungureni, Fagaras Mts, Transfagaras, 1280 m	45.5626	024.6093	Kolcsár L.-P.
<i>Platyura marginata</i> Meigen, 1804	1♂	24-V-2014	RO, Deva, Cetatea Valley, 487 m	45.8842	022.8725	Kolcsár L.-P.
<i>Platyura marginata</i> Meigen, 1804	1♂	25-VII-2014	RO, Sinaia, Bucegi Mts, Stana Regala, 1260 m	45.3684	025.5222	Kolcsár L.-P.
<i>Rymosia lundstroemi</i> Dziedzicki, 1910	1♂	11-V-2010	ME, Zabljak, Durmitor Mt., Crna lake, 1448 m	43.1486	019.0882	Kolcsár L.-P.
<i>Rymosia lundstroemi</i> Dziedzicki, 1910	1♂	25-IX-2014	RO, Vanvucesti, Vartop Mt., Varciorog waterfall, 1000 m	46.4714	022.7376	Kolcsár L.-P.
<i>Stigmatomeria crassicornis</i> (Stannius, 1831)	1♂	16-V-2014	RO, Cluj-Napoca, Cartierul Manastur, 434 m	46.7497	023.5467	Kolcsár L.-P.
<i>Symmerus (Symmerus) annulatus</i> (Meigen, 1830)	1♂	18-VII-2014	RO, Busteni, Bucegi Mts, Cerbului Valley, 1082 m	45.4474	025.5074	Kolcsár L.-P.
<i>Symmerus (Symmerus) annulatus</i> (Meigen, 1830)	1♂	12-VII-2014	RO, Feleacu, Morii valley, Fagus-Carpinus forest, 620 m	46.6958	023.5912	Kolcsár L.-P.
<i>Synapha vitripennis</i> (Meigen, 1818)	1♂	18-V-2014	RO, Grosii Tiblesului, Tibles Mts, Minghet Valley, 777 m	47.5685	024.1117	Kolcsár L.-P.
<i>Synplasta gracilis</i> (Winnertz, 1864)	1♂	5-V-2014	RO, Varghis, Varghis Canyon, 630 m	46.2174	025.5435	Török E.
<i>Synplasta rufilatera</i> (Edwards, 1941)	1♂	16-V-2014	RO, Cluj-Napoca, Cartierul Manastur, 434 m	46.7497	023.5467	Kolcsár L.-P.
<i>Tarnania fenestralis</i> (Meigen, 1818)	1♂	5-V-2014	RO, Varghis, Varghis Canyon, 630 m	46.2174	025.5435	Török E.
<i>Trichonta comis</i> Gagne, 1981	1♂	17-V-2013	RO, Repedea, Maramures Mt., Repedea Valley, 788 m	47.8867	024.3904	Kolcsár L.-P.
<i>Trichonta girschneri</i> Landrock, 1912	1♂	17-V-2014	RO, Luncavita, Macin, Fagilor Valley, 151 m	45.1984	028.3422	Kolcsár L.-P.
<i>Trichonta vulcani</i> (Dziedzicki, 1889)	1♂	23-VIII-2014	RO, Balan, Hasmas Mts, Galkut valley, small brook, 1050 m	46.6493	025.8415	Kolcsár L.-P.
<i>Urytalpa rhapsodica</i> Chandler, 1995	1♂	21-VIII-2014	RO, Gura Haitii, Caliman Mts, near to mine, 1640 m	47.1071	025.2385	Kolcsár L.-P.
<i>Zygomyia humeralis</i> (Wiedemann, 1817)	3♂	28-V-2014	RO, Sacele, Baiul Mts, Bratocea Pass, 1177 m	45.4790	025.8936	Kolcsár L.-P., Török E.

classifies taxa according to alphabetical order.

In order to assess sample completeness of material collected from Romania, an individual rarefaction analysis was performed. Based on the analysis, the total number of studied specimens (131, x-axis) was plotted against the total number of observed species (86, y-axis). Data from Montenegro was omitted due to the rather small number of specimens collected and the remoteness of the sampling sites in the Carpathian Mountains. The analysis was calculated by using PAST 3.11 program (Hammer et al. 2001).

Morphological structures were examined using an Olympus CH2 microscope, Optika B-150 equipped with a Canon 650D camera and a LM Digital SLR Adapter. Layer photos were combined using the software Zerene Stacker.

The morphological terminology used here follows (Søli et al. 2000) and wing venation (Amorim and Rindal 2007).

Results

During this short and non-targeted survey 95 species collected from Montenegro, Romania, and Serbia (see Table 1) were identified. Our rarefaction curve based on material collected from Romania is still rising, meaning that the survey was incomplete (Fig. 2); more thorough faunistic study would yield more species from the area. Some rare and poorly known species were identified, such as *Ditomyia fasciata* (Meigen, 1818), *Antlemon brevimanum* (Loew, 1871), and *Mycetophila curviseta* Lundström, 1911 (Table 1).

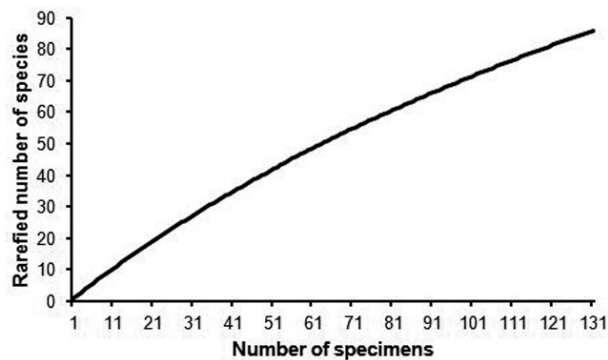


Figure 2. Individual rarefaction curve of specimens identified from Romania.

List of fungus gnats (Keroplatidae, Bolitophilidae and Mycetophilidae) species new to Montenegro, Romania, and Serbia

Family Keroplatidae Rondani, 1856

Subfamily Keroplatinae Rondani, 1856

Cerotelion striatum (Gmelin, 1790)

Material examined. Figure 1. Serbia: Sikirje, Kuka-vica Mts, small brook in deciduous forest, 648 m above sea level (a.s.l.), 42.6526° N, 021.8802° E, 30 June 2017, Kolcsár L.-P., Török E. leg., 1 male.

A Western Palaearctic species, widely distributed in Europe (Chandler 2013), here reported for the first time from Serbia. A relatively rare species, being associated with old trees (Ševčík and Vonička 2008) and Red-Listed in Czech Republic (Máca 2008). Larvae develop in webs and have been reared from *Auricularia auricula-judae* (Jakovlev 2011).

Keroplatus dispar Dufour, 1839

Material examined. Figure 1. Romania: Cluj-Napoca, Mikó Garden, unused old botanical garden with old trees, 373 m a.s.l., 46.7635° N, 023.5804° E, 15 September 2014–20 October 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0277.

A very rare European species, reported from France, Kaliningrad, Sweden (Matile 1986), Russian Karelia (Polevoi 2000), Norway (Økland and Sølvi 1992), Czech Republic and Slovakia (Ševčík and Košel 2009a), and Belgium (Chandler 2013). *Keroplatus dispar* was redescribed by Matile (1986), but Kjærandsen et al. (2007) claimed that both *K. dispar* and *K. tuvensis* Zaitzev, 1991 may actually fall within the intraspecific variation of the widespread *K. testaceus* (Dalman, 1818). The male specimen studied by JS fits well to the illustrations provided by Matile (1986, 1990) (inner view of gonostylus, 9th tergite and proctiger), and thus we report the species here for the first time from Romania. *Keroplatus* larvae spin mucous webs on the surfaces of polyporous fungi and feed mostly on fungal spores (Matile 1990).

Keroplatus reaumurii ssp. *reaumurii* Dufour, 1839

Material examined. Figure 1. Romania: Vanvucești, Vârtop Mt., Vârciorog waterfall, mixed forest, 1000 m

a.s.l., 46.4714° N, 022.7376° E, 25 September 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0268.

Matile (1986) revised this large and conspicuous species. The nominate subspecies was listed by Matile from France and Iran, and has subsequently been reported from Central and southern Europe, including Hungary, Greece, and Azerbaijan (Zaitzev 1994, Chandler 2013). Matile (1986) ranked *K. pentophthalmus* Giglio-Tos, 1890 as a subspecies of *K. reaumurii* and listed this taxon from Romania. The specimen studied by J. Salmela has a very narrow ventromedial appendage on gonocoxite, thus fitting the description of the nominate subspecies (Matile 1986, fig. 11).

Macrorrhyncha rostrata (Zetterstedt, 1851)

Material examined. Figure 1. Romania: Dobrești, Bucegi Mt., Săua Dichiului, spruce forest, 1575 m a.s.l., 45.3736° N, 025.4391° E, 26 July 2014, Keresztes L. leg., 1 male, DIPT-JS-2014-0451; Cluj-Napoca, Mikó Garden, unused old botanical garden with old trees, 373 m a.s.l., 46.7635° N, 023.5804° E, 15 September 2014–20 October 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0228.

A rather widespread European species, known from Central and Northern Europe and Russia (Chandler 2013), here reported for the first time from Romania. This species is, however, quite rare, being associated with pristine forests in Fennoscandia (Kjærandsen and Chandler 2011, Westling 2015); it is Red-Listed in Finland (Penttinen et al. 2010) and Sweden (Westling 2015). Immature stages are unknown, but most likely larvae of the genus dwell in decaying wood (Hutson et al. 1980).

Monocentrotro matilei Bechev, 1989

Material examined. Figure 1. Romania: Pădurea Neagră, Plopiș Mt., Bistra brook, hornbeam-beech forest, 441 m a.s.l., 47.1492° N, 022.4441° E, 8 May 2012, Török E. leg., 1 male, DIPT-JS-2016-0009.

Monocentrotro matilei was originally described from Bulgaria and Algeria (Bechev 1989). Papp (2003) reported this species from Hungary, and Chandler (2009) redescribed it based on specimens collected from Sardinia. The species is here reported for the first time from Romania. The species apparently has a fragmented range confined to mountainous areas of southeastern Europe and the Mediterranean area. Immature stages of *M. matilei* are unknown, but the European species *M. lundstromi* Edwards, 1925 is probably associated with decaying wood (Jakovlev et al. 2014).

Urytalpa rhapsodica Chandler, 1995

Material examined. Figure 1. Romania: Gura Haitii, Călimani Mt., spruce forest near to mine, 1640 m a.s.l., 47.107088° N, 025.2385° E, 21 August 2014, Kolcsár L.-P. leg., 3 males, DIPT-JS-2016-0239.

The species was described from Slovakia (Chandler 1995) and has been later found from the Czech Republic (Ševčík 2004), Switzerland, Bulgaria (Bechev and Koç 2008), and Italy (Kjærandsen et al. 2009) and is here reported for the first time from Romania. Immature stages

are unknown, but larvae of *Urytalpa* probably dwell among soil, moss carpets, or decaying wood (Jakovlev et al. 2014). Based on those few published localities, the species is most likely associated with European montane forests.

Subfamily Macrocerinae Rondani, 1856

Macrocera longibrachiata Landrock, 1917

Material examined. Figure 1. Romania: Stâna de Vale, Bihor Mt., Valea Ciripa, spruce forest around cold springs, 1375 m a.s.l., 46.6756° N, 022.6412° E, 22 May 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0188.

Landrock (1917) described this species from Germany. Also known from Great Britain (Hutson et al. 1980), Slovakia (Ševčík 2007), Estonia (Kurina 1998), Belgium, France, Italy (Chandler 2013), and new for the Romanian fauna. A very rare and poorly known species, immature stages are unknown. Larvae of *Macrocera* are predaceous, living on various substrates, such as soil and dead wood (Hutson et al. 1980, Ševčík and Roháček 2008).

Macrocera pilosa Landrock, 1917

Material examined. Figure 1. Romania: Vârghiș, Vârghiș Canyon, hornbeam-beech forest, 630 m a.s.l., 46.2174° N, 025.5435° E, 5 May 2014, Török E. leg., 2 males, DIPT-JS-2016-0252.

Rather widespread Palearctic species, known from Central and northern Europe, Bulgaria, and Siberia (Zaitzev 1994, Chandler 2013), and new for Romanian fauna. Immature stages are unknown.

Family Bolitophilidae Winnertz, 1863

Bolitophila (Bolitophila) austriaca (Mayer, 1950)

Material examined. Figure 1. Montenegro: Žabljak, Durmitor Mt., Crna lake, spruce forest along a small stream, 1448 m a.s.l., 43.1486° N, 019.0882° E, 11 May 2010, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0246; Romania: Voineasa, Parâng Mt., Vidra lake, valley Mogoșu, spruce forest along a small stream near to a deforestation area, 988 m a.s.l., 45.4212° N, 023.7621° E, 26 May 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2015-0301.

A rather widespread Palearctic species. In Europe *B. austriaca* is known from central and northern parts and Russia but is mostly absent or non-detected from a wide belt ranging from Greece to Lithuania (Chandler 2013). It is new for Romania and Montenegro. The species has been reared from a fruiting body of *Tricholoma focale* in Russian Karelia (Jakovlev 1994).

Bolitophila (Bolitophila) saundersi (Curtis, 1836)

Material examined. Figure 1. Montenegro: Cetinje, Bjelosi, Lovćen NP, dry beech forest, 950 m a.s.l., 42.3670° N, 018.8911° E, 3 May 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0265. Romania: Trei Fântâni, Hășmaș Mt., Bicaz-Mic brook, spruce (*Picea abies*) and hazelnut (*Corylus avellana*) trees around a peat bog, 900 m a.s.l., 46.769° N, 025.8412° E, 17 July 2013, Kolcsár

L.-P. leg., 1 male, DIPT-JS-2016-0181.

A widespread palaeartic species (Chandler 2013), here reported for the first time from Montenegro. It was reared from a number of agaric fruiting bodies (Jakovlev 1994), especially from *Hypholoma fasciculare* (Ševčík 2010).

Bolitophila (Bolitophila) tenella Winnertz, 1864

Material examined. Figure 1. Romania: Voineasa, Parâng Mt., Vidra lake, valley Mogoșu, spruce forest along a small stream near to a deforestation area, 988 m a.s.l., 45.4212° N, 023.7621° E, 26 May 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2015-0302; Cașinu Nou, Bodoc Mt., Balaj pass, spruce forest along a brook with fallen trees, 880 m a.s.l., 46.1949° N, 025.9887° E, 25 August 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0218; Carnic, Retezat Mt., Pietrele Valley, mixed forest, dominated by spruce, 978 m a.s.l., 45.4405° N, 022.8913° E, 25 May 2014, Kolcsár L.-P. leg., 1 male; Obârșia Lotrului, Parâng Mt., peat bog with fallen spruce trees, 1450 m a.s.l., 45.4319° N, 023.6618° E, 26 May 2014, Kolcsár L.-P. leg., 1 male; Brădești, Târnava Mare River, willow forest among the river, 490 m a.s.l., 46.3314° N, 025.3311° E, 1 May 2010, Kolcsár L.-P. leg., 1 male.

It is a widespread Palearctic species, known from several European countries (Chandler 2013), and new for the Romanian fauna. Larvae are polymycophagous, feeding on several agaric mushrooms (Jakovlev 1994, 2011). According to Ševčík (2010), *B. tenella* is an uncommon species in the Czech Republic, perhaps preferring *Pholiota* species as host fungi.

Bolitophila (Cliopisa) melanoleuci Polevoi, 1996

Material examined. Figure 1. Montenegro: Herceg Novi, Morinj, mediterranean forest dominated by *Quercus* near a small town, 285 m a.s.l., 42.4894° N, 018.6290° E, 13 May 2010, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0250.

It is a very rare and poorly known European species. The species was described from Russian Karelia (Polevoi 1996) and there are unpublished records from Norway and Sweden (Kjærandsen 2012). Thus, *B. melanoleuci* is here reported for the first time outside Fennoscandia. Adults have been reared from the fruiting bodies of *Melanoleuca melaleuca* and *M. brevipes* (Polevoi 1996), and these fungi grow in grassy habitats, including man-made parks (Salo et al. 2016); the type material of the species was actually reared from mushrooms growing on city parks (Jakovlev 1993), as *B. latipes* Tollet, 1943).

Bolitophila (Cliopisa) occlusa Edwards, 1913

Material examined. Figure 1. Romania: Stâna de Vale, Bihor Mt., Valea Ciripa, spruce forest around cold springs, 1375 m a.s.l., 46.6756° N, 022.6412° E, 22 May 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0204.

It is a widespread Palearctic species, known from several European countries (Chandler 2013) and new to the Romanian fauna. It was reared from soft-bodied polypore fungi (*Postia* spp., Ševčík 2010). Records

from agarics *Hypholoma fasciculare* and *Pleurotus* are reported (Jakovlev 2011) but may represent another *Bolitophila* species.

Family Mycetophilidae Newman, 1834

Subfamily Mycomyinae Edwards, 1925

Mycomya (Mycomya) alpina Matile, 1972

Material examined. Figure 1. Romania: Azuga, Baiul Mts, Limbășel Valley, spruce forest, 1250 m a.s.l., 45.4989° N, 025.6079° E, 26 August 2014, Kolcsár L.-P. leg., 2 males, DIPT-JS-2016-0208 and DIPT-JS-2016-0211; Borsec, Giurgeu Mts, harvested spruce forest, 1125 m a.s.l., 46.9901° N, 025.5534° E, 21 August 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0221.

Matile (1972) described this species from the French and Italian Alps. Later Väisänen (1984) redescribed this species and also reported it from Austria and Russian Far East (Kuril Islands). Thus, this species has a highly disjunct Palaearctic range. *Mycomya alpina* is here reported for the first time in Europe outside the Central European Alps. Immature stages are unknown, but *Mycomya* larvae are mostly associated with decaying wood and wood-growing fungi (Väisänen 1984), and some species dwell in soil, litter, *Carex*-tussocks (Ševčík and Roháček 2008, Jakovlev 2011) and even on (semi) aquatic lakeshores (Przhiboro 2012).

Mycomya (Mycomya) egregia (Dziedzicki, 1885)

Material examined. Figure 1. Romania: Anieș, Rodnei Mt., Cepelor spring, spruce forest with fallen trees, 1165 m a.s.l., 47.5380° N, 024.6728° E, 19 August 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0180.

This species has been reported from Central and northern Europe and Russia (Väisänen 1984, Chandler 2013) and is here reported as a new for the Romanian fauna. Immature stages are unknown.

Mycomya (Mycomya) neohyalinata Väisänen, 1984

Material examined. Figure 1. Romania: Gura Lalai, Maramureș Mts, Bistrița River, young spruce forest around small springs, 1230 m a.s.l., 47.5548° N, 024.9451° E, 20 August 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2015-0303.

This is a common and widespread Holarctic species but is new for the Romanian fauna. Larvae of the species are associated with fungal fruiting bodies (Väisänen 1984).

Mycomya (Mycomyopsis) trilineata (Zetterstedt, 1838)

Material examined. Figure 1. Montenegro: Šavnik, Petnjica, dry beech forest, 1057 m a.s.l., 42.9836° N, 019.0733° E, 30 April 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0202.

A widespread Palaearctic species (Väisänen 1984), here reported for the first time from Montenegro. Larvae are mycophagous; adults have been reared from *Leccinum scabrum* (Väisänen 1984), *Laxitextum bicolor*, and *Phlebia tremellosa* (Jakovlev 2011).

Subfamily Sciophilinae Dziedzicki, 1885

Phthinia humilis Winnertz, 1864

Material examined. Figure 1. Romania: Căpățânenii Ungureni, Făgăraș Mt., Transfăgăraș, spruce forest, 1280 m a.s.l., 45.5626° N, 024.6093° E, 27 May 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0251.

A rather widespread European species (Chandler 2013), here reported for the first time from Romania. The species is associated with forests; adults have been reared from decaying wood, moss carpet, and soil (Jakovlev 2011).

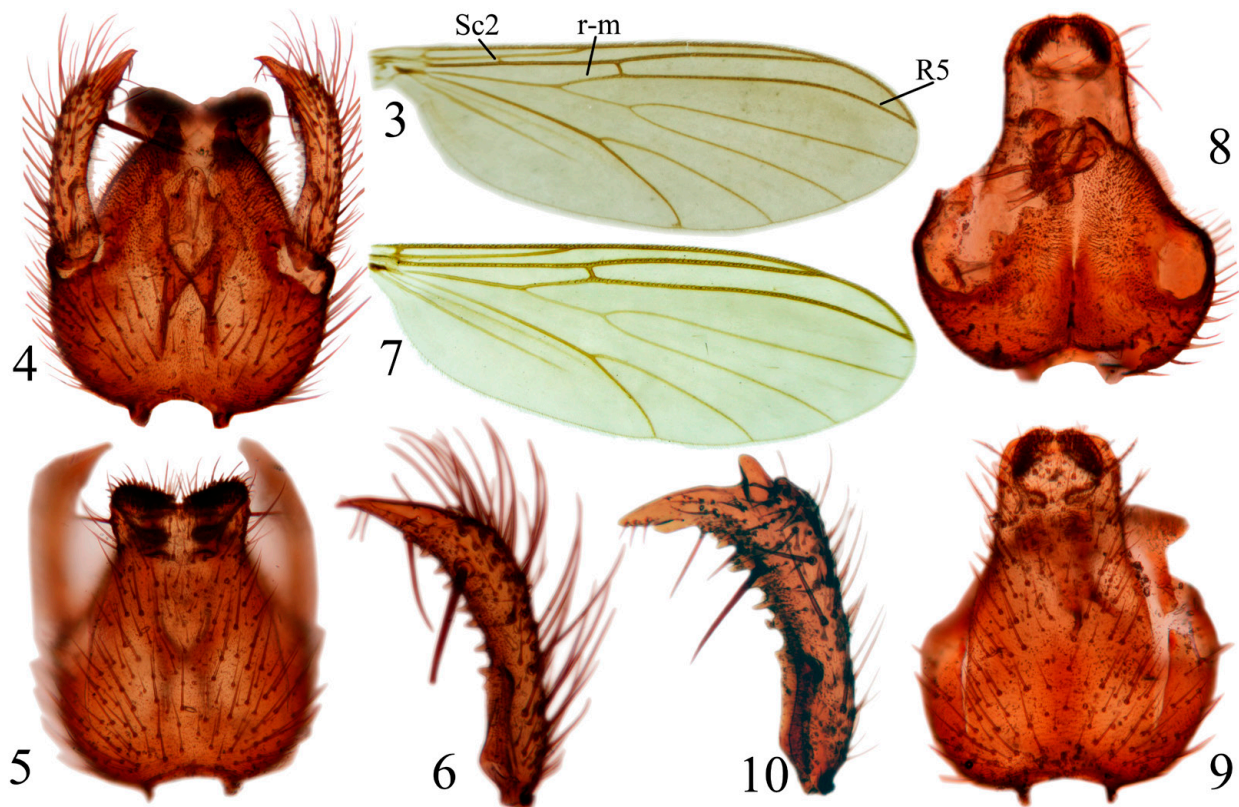
Subfamily Gnoristinae Edwards, 1925

Boletina joosti Plassmann, 1987

Material examined. Figure 1. Romania: Gura Haitii, Călimani Mt., spruce forest near to mine, 1640 m a.s.l., 47.1071° N, 025.2385° E, 21 August 2014, 1 male, DIPT-JS-2016-0240.

This is a rare European species, hitherto known only from Germany. Plassmann (1986) described this species based on a holotype and a paratype males collected from Thuringia and Bavaria, respectively. The male specimens studied by Plassmann were collected in late April and late June. Instead, the specimen from Romania was collected in late August, and thus, *B. joosti* is on the wing during early and late season and adults probably overwinter. The closely related *B. trivittata* (Meigen, 1818) has a similar phenology pattern in Fennoscandia (J. Salmela, pers. obs.). Immature stages of *B. joosti* are unknown, but *B. trivittata* has been reared from rotting wood and soil litter; *Boletina* larvae are seldom found from fungal fruiting bodies (see Salmela et al. 2016 for a review). *Boletina joosti* is probably a very rare species, perhaps having a restricted and fragmented Central European range in mountainous areas.

The genus *Boletina* is still relatively poorly known. Phylogeny and natural classification of the genus are still unresolved (Martinsson et al. 2011), and new species are regularly found, especially from the boreal zone (Salmela et al. 2016). However, in a phylogenetic assessment based on molecular characters, the *Boletina trivittata*-group (consisting of *B. trivittata* and *B. subtrivittata* Zaitzev, 1994) formed a monophyletic clade (Martinsson et al. 2011). This *trivittata*-group is perhaps a sister group to all other recent *Boletina*, and *Katatopygia* Martinsson and Kjærandsen, 2012 is a basal sister group to the *trivittata*-group + remaining *Boletina* (Martinsson et al. 2011). The *trivittata*-group consists of 3 Nearctic species (*B. gracilis* Johannsen, 1912, *B. nacta* Johannsen, 1912, *B. sedula* Johannsen, 1912) and 4 Palaearctic species (*B. augusta* Chandler & Blasco-Zumeta, 2001, *B. joosti*, *B. trivittata*, *B. subtrivittata*). Of the latter, *B. subtrivittata* is known from the Russian Far East and Hokkaido, Japan (Sasakawa and Kimura 1974), as *B. trivittata*, (Zaitzev 1994), while 3 species occur in Europe (Chandler and Blasco-Zumeta 2001).



Figures 3–10. *Boletina trivittata* (Meigen, 1818) (3–6) and *B. joosti* Plassmann, 1987 (7–10), male specimens. 3, 7: wing, 4, 8: hypopygium, ventral view, 5, 9: hypopygium, dorsal view, 6, 10: gonostylus, ventral view.

Key to European *trivittata*-group species

- 1 Costa ending at the apex of R5, cross vein r-m horizontal with R5, i.e. angle ca 180°, (Figs 3, 7). Male gonostylus unbranched (= with a single branch), elongated, very strong, about as long as gonocoxite (Figs 4–6, 8–10) *trivittata*-group ... 2
- Costa extending beyond apex of R5, cross vein r-m forming a shallow angle (ca 160°) with R5. Male gonostylus 1- or 2-branched; if unbranched, then mostly less strong in structure, variable in shape and mostly shorter than gonocoxite
..... *Boletina* other than *trivittata*-gr.
- 2 Wing narrow; veins thick, with brown seams along veins *B. augusta* Chandler & Blasco-Zumeta (known from Spain)
- Wing normal; veins not thickened, with no brown seams along veins (Figs 3, 7) 3
- 3 Sc2 absent (Fig. 7). Male gonostylus with a subapical, tooth-like outgrowth (Fig. 10) *B. joosti* Plassmann (known from Germany and Romania)
- Sc2 present (Fig. 3). Male gonostylus with no subapical, tooth-like outgrowth (Fig. 6)
..... *B. trivittata* Staeger (widespread in Europe)

Coelophthinia thoracica (Winnertz, 1864)

Material examined. Figure 1. Romania: Azuga, Baiul Mts, Limbășel Valley, spruce forest, 1250 m a.s.l., 45.4989° N, 025.6079° E, 26 August 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0209

A rather widespread European species occurring in

northern and central parts, the British Isles, and Russia (Chandler 2013), here reported for the first time from Romania. The species is associated with fungal fruiting bodies (Jakovlev 1994).

Coelosia fusca Bezzi, 1892

Material examined. Figure 1. Montenegro: Žabljak, Durmitor Mt., Crna lake, spruce forest along a small stream, 1448 m a.s.l., 43.1486° N, 019.0882° E, 11 May 2010, Kolcsár L.-P. leg., 1 male, DIPT-JS-2015-0304.

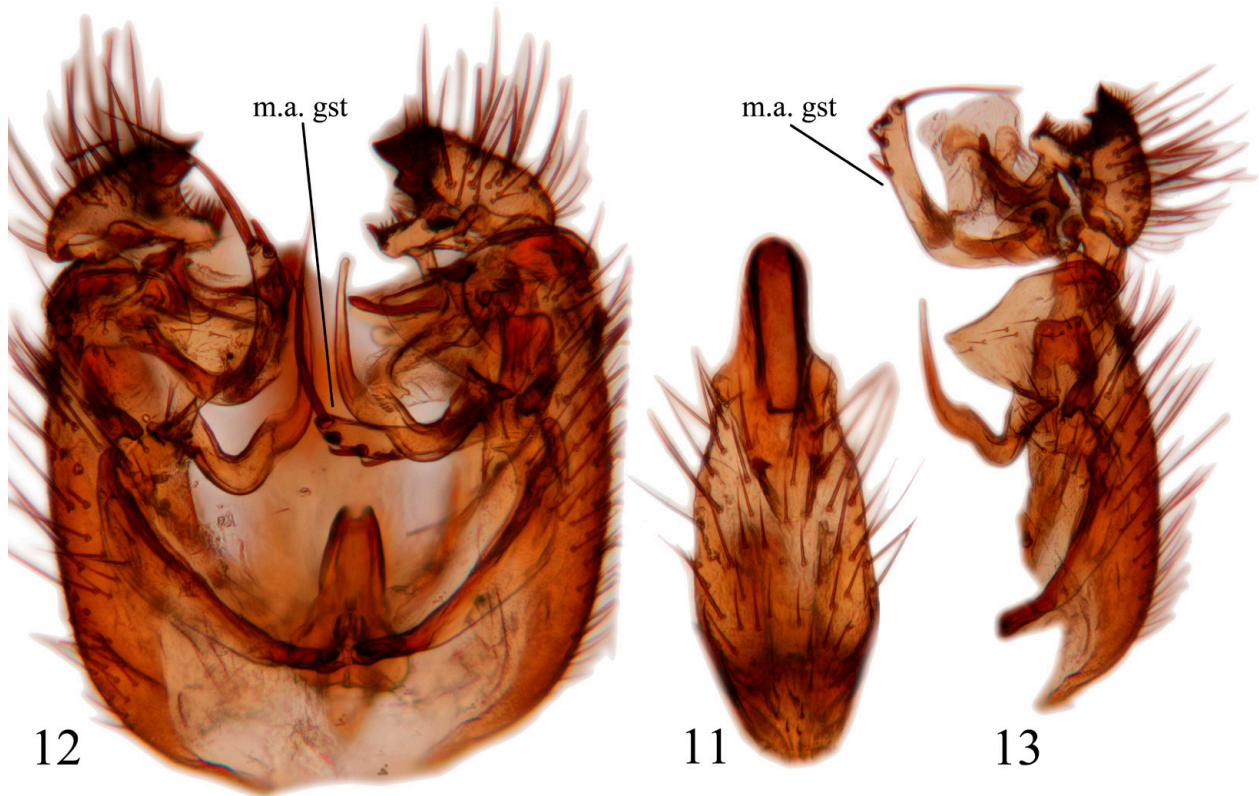
A common and widespread western Palearctic species (Søli 1997, Chandler 2013), here reported as new for the fauna of Montenegro. Members of the genus *Coelosia* occur in humid habitats or even wetlands (Søli 1997, Jakovlev et al. 2014). *Coelosia tenella* Zetterstedt is a fungivorous species (Jakovlev 2011).

Subfamily Leiinae Edwards, 1925

Docosia expectata Laštovka & Ševčík, 2006

Material examined. Figure 1. Romania: Groșii Țibleșului, Tibleș Mt., Minghet Valley, mixed forest, dominated by *Fagus sylvatica*, 777 m a.s.l., 47.5685° N, 024.1117° E, 18 May 2014, Kolcsár L.-P. leg., 1 male DIPT-JS-2016-0226.

The species was described from the Czech Republic and Slovakia (Laštovka and Ševčík 2006), and is also known from Great Britain, Germany, Sweden and Finland (see Jakovlev et al. 2014). The species is new for Romanian fauna. Biology of *D. expectata* is still poorly known, but the species is perhaps associated with broad-leaved forests (Kurina et al. 2004, Jakovlev et al. 2014).



Figures 11–13. *Exechiopsis (Exechiopsis) unguiculata* (Lundström, 1911), male hypopygium. **11:** median appendage of gonocoxite, **12:** hypopygium, dorsal view (cerci omitted), **13:** gonostylus, ventrolateral view; m.a. gst: median appendage of gonostylus.

Docosia larvae are associated with various decaying organic matter, including fungi and rotting wood (Jakovlev et al. 2014). *Docosia* has a close relationship with *Boletina* and should be classified with the Gnoristinae (Rindal et al. 2009, Martinsson et al. 2011).

***Docosia gilvipes* (Walker, 1856)**

Material examined. Figure 1. Montenegro: Žabljak, Durmitor Mt., Crna lake, spruce forest along a small stream, 1448 m a.s.l., 43.1486° N, 019.0882° E, 11 May 2010, Kolcsár L.-P. leg., 1 male, DIPT-JS-2015-0306.

The species is the most common and widespread *Docosia* in the Palearctic region (Laštovka and Ševčík 2006), but not previously reported from Montenegro. Male hypopygium of *D. gilvipes* has been recently illustrated by Laštovka and Ševčík (2006) and Kurina (2008). Larvae are polymycophagous, known to occur in more than 40 species of fungi (Ševčík 2010).

Subfamily Mycetophilinae Newman, 1834

***Allodia (Allodia) lundstroemi* Edwards, 1921**

Material examined. Figure 1. Romania: Vanucești, Vârtoș Mt., Vârciorog waterfall, mixed forest, 1000 m a.s.l., 46.4714° N, 022.7376° E, 25 September 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0270.

A widespread Palearctic species (Zaitzev 2003), here reported for the first time from Romania. Larvae are fungivorous, reared from fruiting bodies of *Lentinus lepideus* and *Laccaria laccata* (Jakovlev 1994).

***Anatella ciliata* Winnertz, 1864**

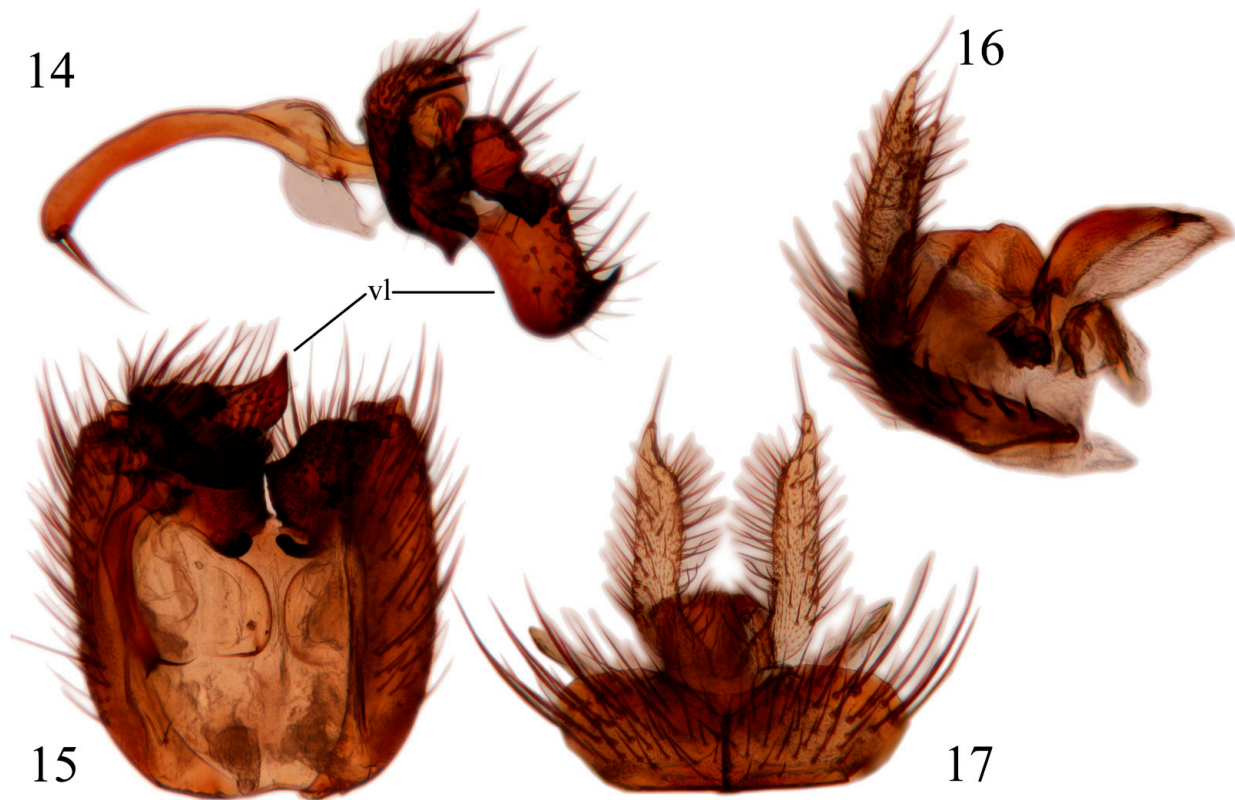
Material examined. Figure 1. Romania: Cluj-Napoca, Mikó Garden, unused old botanical garden with old trees, 373 m a.s.l., 46.7635° N, 023.5804° E, 15 September 2014–20 October 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0230.

A widespread Holarctic species (Chandler 2013), here reported for the first time from Romania. Larvae of *Anatella* are most likely fungivorous (Ševčík 2010), and *A. ciliata* has been reared once from dead wood (Jakovlev 2011).

***Exechiopsis (Exechiopsis) unguiculata* (Lundström, 1911)**

Material examined. Figure 1. Montenegro: Žabljak, Durmitor Mt., Crna lake, spruce forest along a small stream, 1448 m a.s.l., 43.1486° N, 019.0882° E, 11 May 2010, Kolcsár L.-P. leg., 2 males, DIPT-JS-2015-0309 and DIPT-JS-2016-0244. Romania: Sâmbăta de Sus, Făgăraș Mt., Sâmbăta Valley, 1020 m a.s.l., 45.6508° N, 024.7889° E, 29 May 2014, Kolcsár L.-P. leg., 1020 m a.s.l., 1 male.

The description of the species (as *Exechia unguiculata*) was based on material collected from modern Croatia and Austria (Lundström 1911a). The species has a European range, known from southern and Central Europe (Chandler 2013), including Great Britain (Edwards 1925), Spain (Mederos and Chandler 2014), Ukraine (Zaitzev 2003), and is here reported from Montenegro. The species was recently removed from the list of Finnish fungus gnats (Jakovlev 2014). Note that *E. unguiculata* has not been recorded from Japan, as was mentioned by Jakovlev



Figures 14–17. *Rymosia lundstroemi* Dziedzicki, 1910, male hypopygium. **14:** gonostylus, lateral view, **15:** hypopygium, dorsal view (cerci omitted), **16:** 9th tergite, cerci and parameres, lateral view, **17:** 9th tergite and cerci, dorsal view.

(2011), citing Sasakawa and Ishizaki (1999). Immature stages of the species are unknown, but *Exechiopsis* larvae have been reared from both soil-dwelling agarics and saproxylic fungi (Jakovlev 2011). *Exechiopsis unguiculata* has been illustrated only by Lundström (1911a) and Zaitzev (2003), and thus photos of the male hypopygium are provided here. The male hypopygium is characterised by a ventroapical lobe of the gonocoxite that is narrowing apically, its apex rounded, margins straight and bordered by a black frame (Figure 11). The medial appendage of the gonostylus is curved, long and narrow, bearing three subapical black spines arranged in a row and a long apical spine (Figs 12, 13).

***Rymosia lundstroemi* Dziedzicki, 1910**

Material examined. Figure 1. Montenegro: Žabljak, Durmitor Mt., Crna lake, spruce forest along a small stream, 1448 m a.s.l., 43.1486° N, 019.0882° E, 11 May 2010, Kolcsár L.-P. leg., 1 male, DIPT-JS-2015-0308. Romania: Vanucești, Vârtope Mt., Vârciorog waterfall, mixed forest, 1000 m a.s.l., 46.4714° N, 022.7376° E, 25 September 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0259.

The species has a rather restricted Central European range (Chandler 2013), here reported for the first time from Montenegro. An adult male of the species was collected from a cave in Romania (Burghel-Balacesco 1968, as *R. matilei*), but immature stages are unknown. *Rymosia* in general dwell in forests or wetlands (Chandler 1994), and larvae of some species are associated with fungal fruiting bodies (Jakovlev 1994). This species has

been illustrated only twice (Dziedzicki 1910, Burghel-Balacesco 1968, as *R. matilei*). Both illustrations are rather general, and photos of the male hypopygium of *R. lundstroemi* are provided here. Ventral lobe of gonostylus is short and black, truncated (Figure 14); gonocoxites internally with prominent, black mesial hooks (Fig. 15); parameres about as long as cerci, having black, pointed apical spines (Figs 16, 17).

***Stigmatomeria crassicornis* (Stannius, 1931)**

Material examined. Figure 1. Romania: Cluj-Napoca, Cartierul Mănăstur, hornbeam-beech forest near the city, 434 m a.s.l. 46.7497° N, 023.5467° E, 16 May 2014, Kolcsár L.-P. leg., 2 males, DIPT-JS-2016-0273.

The species is probably very widespread in Europe (Chandler 2013) and has already been reported by Pârnu (2003, 2004a) from Romania. *Stigmatomeria crassicornis* is, however, close to *S. obscura* (Winnertz, 1864), a species that had been for decades considered as a junior synonym of *S. crassicornis* (see Kjærandsen et al. 2007, Salmela and Kaunisto 2015). Pârnu (2003) reported 2 females from Maramureș Depression, and the sex of the specimen examined from Piatra Craiului National Park was not specified (Pârnu 2004a). Because Pârnu's records may in principle also belong to *S. obscura*, the presence of *S. crassicornis* in Romania is verified herein. Larvae of the species may be associated with truffles (Edwards 1925, Chandler 2010), but Jakovlev (2011) reared the species from *Ascocoryne sarcoides* on birch log.

Synplasta gracilis Winnertz, 1864

Material examined. Figure 1. Romania: Vârghiș, Vârghiș Canyon, hornbeam-beech forest, 630 m a.s.l., 46.2174° N, 025.5435° E, 5 May 2014, Török E. leg., 2 males, DIPT-JS-2016-0254.

This is a rather widespread European species, here reported for the first time from Romania. Biology of the genus is poorly known, but *S. gracilis* has been reared from *Mycoacia uda* and *Pleurotus dryinus* (Chandler 1993), and Zaitzev (2003) referred to a record from *Coprinus* fruiting bodies. *Synplasta* species are generally rare in the boreal zone, mainly associated with deciduous or old growth forests (J. Jakovlev, pers. comm.), and thus, *S. gracilis* is Red-Listed (NT) in Finland (Penttinen et al. 2010).

Synplasta rufilatera (Edwards, 1941)

Material examined. Figure 1. Romania: Cluj-Napoca, Cartierul Mănăstur, hornbeam-beech forest near the city, 434 m a.s.l. 46.7497° N, 023.5467° E, 16 May 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0273.

This rare and poorly known European species was described from Great Britain (Edwards 1941, as *Rhymosia*) and was later recorded only from Poland (Zatwarnicki 2001, as *Allodiopsis*), Germany (Ševčík 2001), Finland (Jakovlev 2014), Sweden (Kjærandsen et al. 2007), Russian Karelia, Ukraine and Siberia (Zaitzev 2003); it is here reported for the first time from Romania. Immature stages are unknown, but most likely it is a forest associated species. In Fennoscandia, this species is observed from both nemoral and north boreal vegetation zones (Kjærandsen et al. 2007, J. Jakovlev pers. comm.). However, the species may consist of 2 taxa, proper *S. rufilatera* sensu Edwards and an undescribed taxon sensu Zaitzev (2003) (P.J. Chandler, pers. comm.).

Mycetophila deflexa Chandler, 2001

Material examined. Figure 1. Romania: Sânmartin, Ciuc Mts, Rugat Valley, young spruce forest along a small stream with old fallen trees, 950 m a.s.l., 46.2669° N, 026.0103° E, 25 August 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0008.

The species has a wide range, extending from Great Britain (Chandler 2000) and Portugal (Ribeiro 2003) to the Russian Far East (Zaitzev 2003). In Europe the species is also known from France (Chandler 2013), Finland (Jakovlev 2014), and Russian Karelia (Zaitzev 2003). This species is very close to *M. gratiosa* Winnertz, and some of the records published as *M. gratiosa* may actually belong to *M. deflexa* (e.g., Polevoi 2001) from eastern Finland). The species is Red-Listed in Finland (Penttinen et al. 2010), and it has been found from forests characterised by a high amount of dead woody material (J. Jakovlev, pers. comm.). Immature stages are unknown, but *Mycetophila* larvae live inside fungal fruiting bodies (Ševčík 2010, Jakovlev 2011).

Mycetophila lastovkai Caspers, 1984

Material examined. Figure 1. Romania: Vulcan Mt.,

Vulcan Pass, spruce forest along a stream with old fallen trees, 1420 m a.s.l., 45.3067° N, 023.3064° E, 26 May 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2014-0452; Remeți, Bihor Mt., Iad River, 870 m a.s.l., mixed forest, 46.7195° N, 022.5640° E, 22 May 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0201. Carnic, Retezat Mt., Pietrele Valley, mixed forest, dominated by spruce, 978 m a.s.l., 45.4405° N, 022.8913° E, 25 May 2014, Kolcsár L.-P. leg., 1 male; Vanvucești, Vârtop Mt., Vârciorog waterfall, mixed forest, 1000 m a.s.l., 46.4714° N, 022.7376° E, 25 September 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0260.

This species was described from Germany (Caspers 1984) and was later found in the British Isles (Chandler 1988), Bulgaria, Central Europe (Chandler 2013), Sweden, Norway, and Denmark (Kjærandsen 2012). However, a recently described species, *M. gemerensis* Ševčík & Kurina 2011, is close to *M. lastovkai*, and thus older records of the latter species should be verified due to possible confusion of these 2 taxa (Salmela and Kaunisto 2015). Immature stages of *M. lastovkai* are unknown.

Mycetophila marginata Winnertz, 1864

Material examined. Figure 1. Montenegro: Žabljak, Durmitor Mt., Crna lake, spruce forest along a small stream, 1448 m a.s.l., 43.1486° N, 019.0882° E, 11 May 2010, Kolcsár L.-P. leg., 1 male, DIPT-JS-2015-0307.

A widespread European species (Chandler 2013), here reported for the first time from Montenegro. Larvae are fungivorous (Zaitzev 2003; Ševčík 2010), most often reared from saproxylic fungi.

Mycetophila stylata (Dziedzicki, 1884)

Material examined. Figure 1. Romania: Pintec, Giurgiu Mt., Pintec valley, deforestation area, 820 m a.s.l., 46.9255° N, 025.8533° E, 22 August 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2014-0465.

A widespread European species (Chandler 2013), here reported for the first time from Romania. *Mycetophila stylata* has been reared once from a *Lactarius* fruiting body (Jakovlev 1994).

Phronia forcipula Winnertz, 1864

Material examined. Figure 1. Montenegro: Žabljak, Durmitor Mt., Crna lake, spruce forest along a small stream, 1448 m a.s.l., 43.1486° N, 019.0882° E, 11 May 2010, Kolcsár L.-P. leg., 1 male, DIPT-JS-2015-0305. Cetinje, Bjelosi, Lovćen NP, dry beech forest, 950 m a.s.l., 42.3670° N, 018.8911° E, 3 May 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0264. Romania: Vulcan Mt., Vulcan Pass, spruce forest along a stream with old fallen trees, 1420 m a.s.l., 45.3067° N, 023.3064° E, 26 May 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2014-0453. Bălan, Hășmaș Mt., Gălkút valley, spruce forest along a small brook, 1050 m a.s.l., 46.6493° N, 025.8415° E, 23 August 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0195.

A widespread Holarctic species (Gagné 1975, Zaitzev 2003), here reported for the first time from Montenegro.



Figures 18–24. *Trichonta comis* Gagné, 1981, male hypopygium. **18:** 9th tergite and cerci, dorsal view, **19:** hypopygium, dorsal view (cerci omitted), **20:** gonostylus, inner lateral view, **21:** dorsal lobe of gonostylus, **22:** apical half of the ventral lobe of gonostylus, **23:** median projection of ventral lobe of gonostylus, dorsolateral view, **24:** aedeagus.

Larvae have been once observed from the surface of *Cor-ticium* fruiting body (Edwards 1925).

***Phronia nigricornis* (Zetterstedt, 1852)**

Material examined. Figure 1. Romania: Sălătruc, Ciuc Mt., Uz Valley, beech forest, 645 m a.s.l., 46.3452° N, 026.2715° E, 25 August 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0256.

A widespread Holarctic species (Gagné 1975, Zaitzev 2003), here reported for the first time from Romania. Larvae of the genus *Phronia* dwell on decaying logs, often associated with slime moulds (Myxomycetes) (Zaitzev 2003). Immature stages of *P. nigricornis* are unknown.

***Trichonta comis* Gagné, 1981**

Gagné 1981: 17 (figs 126–128)

Material examined. Holotype male. Finland, Kuusamo, Juuma, 21 August 1964 R. Tuomikoski leg., pinned specimen, abdomen in a microvial in glycerol (MZHF). Romania: Repedea, Maramureş Mt., wet naturally opened area in a spruce forest with fallen trees, 788 m a.s.l., 47.8867° N, 024.3904° E, 17 May 2014, Kolcsár L.-P. leg., DIPT-JS-2016-0220, 1 male. Figure 1.

Redescription of male. Head. Dark brown, vertex with numerous short pale setae. Lateral ocelli touching eye margins, median ocellus small, arranged in a shallow triangle. Ommatidia pubescent. Clypeus rounded, length/width ratio 0.30. Face and clypeus light brown, bearing light setae. Mouthparts light brown. Palpus 5-segmented, segments yellowish brown, segments 3–5 apically

slightly infuscated. Palpal segments 2–5 bearing strong dark dorso-apical setae, their length about as long as or shorter than respective palpus width. Ratios of 3 apical palpal segments (3:4, 4:5): 0.79, 0.65. Sensilla cochleariformis present on base of third palpomere, arranged in a regular circular shape. Scape and pedicel yellowish brown, first flagellomere basally yellowish brown, apically darker; flagellomeres 2–14 dark brown. Scape elongated, conical, pedicel globular, their length ratio (length of scape/pedicel) 1.69. Flagellomeres cylindrical, except last one slightly conical, length/width ratios of first, second, fifth and last flagellomeres 2.72, 1.82, 2.0 and 4.1, respectively.

Thorax. Mainly brown, anterior corners of scutellum, antepronotum and proepisternum yellowish. Anepisternum with 5 or 6 strong setae and with several inconspicuous setae. Laterotergite setose, other pleural plates glabrous. Halteres yellow, stem sparsely setose.

Legs. Coxae yellowish, apically infuscated, anterior edge of fore coxa bearing several light setae. Hind coxa with a posterobasal seta and hind tibia without posteroventral and anteroventral setae. Trochanters yellowish, ventrally black. Femora yellowish, with a narrow ventral band fading toward femoral apices. Tibiae and tarsi yellowish. Anteroapical depressed area of fore tibia ovate, bearing a comb of circa 26 hyaline setae. Length ratios of femur to tibia for fore-, mid- and hind legs: 1.0, 0.95, 0.88. Length ratios of tibia to basitarsus for fore-, mid- and hind legs: 1.24, 1.5, 1.62.

Wing. Hyaline, length 3.2 mm. C and R veins dark brown, M1 and M2 mostly brown, CuA and M4 brown; veins r-m, first sector of M4, M1+2 and bases of M1 and M2 pale. These pale veins and Rs are also glabrous, bearing no stout, dark setae.

Abdomen. Brown, bearing light setae. Ninth tergite and cerci as in Figure 18. Gonocoxite dark brown, bearing numerous light setae (Fig. 19); ventrocaudal margin with a very modest median elevation. Gonostylus composed of two main lobes, dorsal and ventral (Fig. 20). Dorsal lobe (Fig. 21) is simple, lingulate, widest medially and narrowing apically; 14 stout setae on inner surface and four setae in a row on dorsal margin. Ventral lobe is intricate, consisting of the lobe itself and median projections. Apical half of the ventral lobe of gonostylus is rectangular; caudal outer corner is angular, bearing a dense dark setosity; inner corner with a finger-like projection and with a long subapical seta (Fig. 22); caudal margin even/truncated, with rather long pale setae. One of the projections of the ventral lobe is conspicuous, erect, dark brown; its stalk is widest basally, glabrous, narrowing toward apex; the apex is a beak-like structure, having a larger, pointed tooth and a comb consisting of ca 7–10 blunt teeth (Fig. 23). Aedeagus is rather wide, with a deep U-shaped median incision; apex truncated, rounded; caudal lateral corners with several sharp teeth (Fig. 24).

Female. Unknown.

Biology. Immature stages of the species are unknown, but adults have often been collected from forests (see Taxon discussion below).

Taxonomic discussion. Holarctic species of the genus *Trichonta* were revised by Gagné (1981), and since then, 20 northern hemisphere species have been described (Chandler 1992, 2009, Bragina 1994, Chandler and Ribeiro 1995, Wu et al. 1995; Zaitzev 2003, Hong et al. 2007). *Trichonta comis* is characterized by the presence of single posterobasal seta on hind coxa, absence of ventral setae on hind tibia, setosity of Cu petiole and hyaline wing lamina. In addition, the apical half of ventral lobe of gonostylus is bears a long seta. The aedeagus, not illustrated or verbally described by Gagné (1981), is also quite peculiar due to its spinose caudal corners. The type locality in Finland (Kuusamo, Juuma) is not known in detail, but is most likely within Oulanka National Park; the holotype male is the only specimen recorded from Finland. In Norway the species has been collected from both birch (*Betula*) (Søli 1994) and spruce (*Picea*) (Økland and Zaitzev 1997) dominated forests. The species is also known from the Czech Republic (Ševčík 1999, collected from a virgin beech forest) and Slovakia (Ševčík and Košel 2009b). It is new to the Romanian fauna. *Trichonta comis* is most likely a rare, boreo-montane forest-dwelling species, showing affinities to pristine forests. Larvae of *Trichonta* are predominantly associated with wood-decaying fungi (e.g., Jakovlev 2011)

Trichonta vulcani (Dziedzicki, 1889)

Material examined. Figure 1. Romania: Bălan, Hășmaș Mt., Gálkút valley, spruce forest along a small brook, 1050 m a.s.l., 46.6493° N, 025.8415° E, 23 August 2014, Kolcsár L.-P. leg., 1 male, DIPT-JS-2016-0192.

A widespread Holarctic species (Gagné 1981, Zaitzev 2003), here reported as a new to the Romanian fauna. Immature stages are unknown.

Discussion

Based on individual rarefaction analysis, the obtained curve is still steadily rising, showing no signs of leveling off (Fig. 2). Thus, additional sampling and further examination of specimens is expected to increase the number of observed species. Hence, these results suggest a potentially species-rich, yet poorly researched, Sciaroidea fauna. In general, the lower Diptera fauna of Romania is still little investigated, evidenced by the discovery of several new species and new regional records (Ujvárosi and Krzeminska 2002, Ujvárosi et al. 2011a, 2011b, Kolcsár et al. 2012, 2016, Török et al. 2013, Tkoč and Roháček 2014). Regarding the Carpathians, the systematic surveys performed so far suggest a diverse and poorly known Tipuloidea and Ptychopteridae fauna of this mountain range (Keresztes et al. 2012, Dénes et al. 2015, 2016, Török et al. 2015), and it is also definitely true in the case of Sciaroidea. Only 2 articles have been specifically dealt with the Carpathian Sciaroidea (Pârnu 2002, 2004a), and they added no less than 67 species to the Romanian fauna. All faunistic data (Table 1) and the actual checklist of fungus gnats (Ditomyiidae, Diadocidiidae, Keroplatidae, Bolitophilidae, Mycetophilidae) of Romania (Table 2) and Montenegro and Serbia jointly (Table 3) are available on *TransDiptera Online Database* (Kolcsár et al. 2017).

The Balkan area is a biodiversity hotspot, however, the Diptera fauna of the region is poorly known and has not been sufficiently explored (Ivković and Pont 2015). Only a few articles added new faunistic information to the native Diptera fauna of Montenegro and Serbia in the last 10 years (van Steenis et al. 2015, Plóciennik et al. 2014, Ivković et al. 2013, Stanković et al. 2014). No articles have been specifically dealt with the fungus gnat fauna of Montenegro or Serbia.

Systematic, comprehensive and long-term surveys are necessary to expand our knowledge on the Montenegrin, Romanian, and Serbian dipteran fauna, including the highly diverse Sciaroidea. Long-term passive sampling devices like Malaise traps or bait traps should be used to explore phenology, population dynamics, and assemblage composition of different habitats. Increased faunistic knowledge will provide information about the formation and connection of the Montenegro, Romania, and Serbia fauna with surrounding areas.

Table 2. Checklist of fungus gnats (Ditomyiidae, Diadocidiidae, Keroplatidae, Bolitophilidae, Mycetophilidae) of Romania. New records are listed with asterisk (*).

Family	Subfamily	Species
Bolitophilidae		* <i>Bolitophila (Bolitophila) austriaca</i> (Mayer, 1950)
Bolitophilidae		<i>Bolitophila (Bolitophila) cinerea</i> Meigen, 1818
Bolitophilidae		<i>Bolitophila (Bolitophila) cooremani</i> (Tollet, 1955)
Bolitophilidae		<i>Bolitophila (Bolitophila) lengersdorfi</i> (Tollet, 1955)
Bolitophilidae		<i>Bolitophila (Bolitophila) leruthi</i> (Tollet, 1955)
Bolitophilidae		* <i>Bolitophila (Bolitophila) tenella</i> Winnertz, 1864
Bolitophilidae		<i>Bolitophila (Bolitophila) saundersi</i> (Curtis, 1836)
Bolitophilidae		<i>Bolitophila (Bolitophila) spinigera</i> Edwards, 1925
Bolitophilidae		<i>Bolitophila (Cliopisa) dubia</i> Siebke, 1863
Bolitophilidae		<i>Bolitophila (Cliopisa) edwardsiana</i> Stackelberg, 1969
Bolitophilidae		<i>Bolitophila (Cliopisa) hybrida</i> (Meigen, 1804)
Bolitophilidae		<i>Bolitophila (Cliopisa) latipes</i> Tollet, 1943
Bolitophilidae		<i>Bolitophila (Cliopisa) maculipennis</i> Walker, 1835
Bolitophilidae		<i>Bolitophila (Cliopisa) modesta</i> Lackschewitz, 1937
Bolitophilidae		* <i>Bolitophila (Cliopisa) occlusa</i> Edwards, 1913
Bolitophilidae		<i>Bolitophila (Cliopisa) spelaeicola</i> Tollet, 1955
Diadocidiidae		<i>Diadocidia (Diadocidia) ferruginosa</i> (Meigen, 1830)
Diadocidiidae		<i>Diadocidia (Diadocidia) spinosula</i> Tollet, 1948
Ditomyiidae		<i>Ditomyia fasciata</i> (Meigen, 1818)
Ditomyiidae		<i>Symmerus (Symmerus) annulatus</i> (Meigen, 1830)
Keroplatidae	Keroplatinae	<i>Antlemon (Antlemonopsis) brevimanum</i> (Loew, 1871)
Keroplatidae	Keroplatinae	<i>Cerotelion racovitzai</i> Matile & Burgehele-Balacesco, 1969
Keroplatidae	Keroplatinae	<i>Cerotelion striatum</i> (Gmelin, 1790)
Keroplatidae	Keroplatinae	* <i>Keroplatus dispar</i> (Dufour, 1839)
Keroplatidae	Keroplatinae	<i>Keroplatus reaumurii pentophthalmus</i> Giglio-Tos, 1890
Keroplatidae	Keroplatinae	* <i>Keroplatus reaumurii reaumurii</i> (Dufour, 1839)
Keroplatidae	Keroplatinae	<i>Keroplatus testaceus</i> (Dalman, 1818)
Keroplatidae	Keroplatinae	<i>Macrorrhyncha flava</i> Winnertz, 1846
Keroplatidae	Keroplatinae	* <i>Macrorrhyncha rostrata</i> (Zetterstedt, 1851)
Keroplatidae	Keroplatinae	* <i>Monocentrotroa matilei</i> Bechev, 1989
Keroplatidae	Keroplatinae	<i>Orfelia discoloria</i> (Meigen, 1818)
Keroplatidae	Keroplatinae	<i>Orfelia fasciata</i> (Meigen, 1804)
Keroplatidae	Keroplatinae	<i>Orfelia lugubris</i> (Zetterstedt, 1851)
Keroplatidae	Keroplatinae	<i>Orfelia nemoralis</i> (Meigen, 1818)
Keroplatidae	Keroplatinae	<i>Orfelia nigricornis</i> (Fabricius, 1805)
Keroplatidae	Keroplatinae	<i>Orfelia ochracea</i> (Meigen, 1918)
Keroplatidae	Keroplatinae	<i>Orfelia pallida</i> (Staeger, 1840)
Keroplatidae	Keroplatinae	<i>Platyura marginata</i> Meigen, 1804
Keroplatidae	Keroplatinae	<i>Pyratula zonata</i> (Zetterstedt, 1855)
Keroplatidae	Keroplatinae	<i>Rocetelion humerale</i> (Zetterstedt, 1850)
Keroplatidae	Keroplatinae	* <i>Urytalpa rhapsodica</i> Chandler, 1995
Keroplatidae	Macrocerinae	<i>Macrocera angulata</i> Meigen, 1818
Keroplatidae	Macrocerinae	<i>Macrocera centralis</i> Meigen, 1818
Keroplatidae	Macrocerinae	<i>Macrocera fasciata</i> Meigen, 1804
Keroplatidae	Macrocerinae	<i>Macrocera inversa</i> Loew, 1869
Keroplatidae	Macrocerinae	<i>Macrocera kerteszi</i> Lundstrom, 1911
Keroplatidae	Macrocerinae	* <i>Macrocera longibrachiata</i> Landrock, 1917
Keroplatidae	Macrocerinae	<i>Macrocera parva</i> Lundstrom, 1914
Keroplatidae	Macrocerinae	<i>Macrocera phalerata</i> Meigen, 1818
Keroplatidae	Macrocerinae	* <i>Macrocera pilosa</i> Landrock, 1917
Keroplatidae	Macrocerinae	<i>Macrocera stigma</i> Curtis, 1837
Keroplatidae	Macrocerinae	<i>Macrocera stigmoides</i> Edwards, 1925
Keroplatidae	Macrocerinae	<i>Macrocera vittata</i> Meigen, 1830
Keroplatidae	Macrocerinae	<i>Macrocera zetterstedti</i> Lundstrom, 1914
Mycetophilidae	Gnoristinae	<i>Apolephthisa subincana</i> (Curtis, 1837)
Mycetophilidae	Gnoristinae	<i>Boletina basalis</i> (Meigen, 1818)
Mycetophilidae	Gnoristinae	<i>Boletina cincticornis</i> (Walker, 1848)
Mycetophilidae	Gnoristinae	<i>Boletina dispecta</i> Dziedzicki, 1885
Mycetophilidae	Gnoristinae	<i>Boletina gripha</i> Dziedzicki, 1885
Mycetophilidae	Gnoristinae	* <i>Boletina joosti</i> Plassmann, 1987

Continued

Table 2. Continued.

Family	Subfamily	Species
Mycetophilidae	Gnoristinae	<i>Boletina lundbecki</i> Lundstrom, 1912
Mycetophilidae	Gnoristinae	<i>Boletina moravica</i> Landrock, 1912
Mycetophilidae	Gnoristinae	<i>Boletina nasuta</i> (Haliday, 1839)
Mycetophilidae	Gnoristinae	<i>Boletina nitida</i> Grzegorzek, 1885
Mycetophilidae	Gnoristinae	<i>Boletina plana</i> (Walker, 1856)
Mycetophilidae	Gnoristinae	<i>Boletina sciarina</i> Staeger, 1840
Mycetophilidae	Gnoristinae	<i>Boletina trispinosa</i> Edwards, 1913
Mycetophilidae	Gnoristinae	<i>Boletina trivittata</i> (Meigen, 1818)
Mycetophilidae	Gnoristinae	* <i>Coelophthinia thoracica</i> (Winnertz, 1864)
Mycetophilidae	Gnoristinae	<i>Gnoriste bilineata</i> Zetterstedt, 1852
Mycetophilidae	Gnoristinae	<i>Grzegorzekia collaris</i> (Meigen, 1818)
Mycetophilidae	Gnoristinae	<i>Speolepta leptogaster</i> (Winnertz, 1864)
Mycetophilidae	Gnoristinae	<i>Synapha vitripennis</i> (Meigen, 1818)
Mycetophilidae	Leiinae	<i>Clastobasis alternans</i> (Winnertz, 1864)
Mycetophilidae	Leiinae	* <i>Docosia expectata</i> Lastovka & Sevcik, 2006
Mycetophilidae	Leiinae	<i>Docosia flavicoxa</i> Strobl, 1900
Mycetophilidae	Leiinae	<i>Docosia fuscipes</i> (Roser, 1840)
Mycetophilidae	Leiinae	<i>Docosia lastovkai</i> Chandler, 1994
Mycetophilidae	Leiinae	<i>Docosia moravica</i> Landrock, 1916
Mycetophilidae	Leiinae	<i>Ectrepesthoneura referta</i> Plassmann, 1976
Mycetophilidae	Leiinae	<i>Leia bilineata</i> (Winnertz, 1864)
Mycetophilidae	Leiinae	<i>Leia bimaculata</i> (Meigen, 1804)
Mycetophilidae	Leiinae	<i>Leia crucigera</i> Zetterstedt, 1838
Mycetophilidae	Leiinae	<i>Leia cylindrica</i> (Winnertz, 1864)
Mycetophilidae	Leiinae	<i>Leia maculosa</i> (Strobl, 1900)
Mycetophilidae	Leiinae	<i>Leia subfasciata</i> (Meigen, 1818)
Mycetophilidae	Leiinae	<i>Leia winthemi</i> Lehmann, 1822
Mycetophilidae	Leiinae	<i>Rondaniella dimidiata</i> (Meigen, 1804)
Mycetophilidae	Manotinae	<i>Manota unifurcata</i> Lundstrom, 1913
Mycetophilidae	Mycetophilinae	<i>Allodia (Allodia) anglofennica</i> Edwards, 1921
Mycetophilidae	Mycetophilinae	<i>Allodia (Allodia) lugens</i> (Wiedemann, 1817)
Mycetophilidae	Mycetophilinae	* <i>Allodia (Allodia) lundstroemi</i> Edwards, 1921
Mycetophilidae	Mycetophilinae	<i>Allodia (Allodia) ornaticollis</i> (Meigen, 1818)
Mycetophilidae	Mycetophilinae	<i>Allodia (Allodia) truncata</i> Edwards, 1921
Mycetophilidae	Mycetophilinae	<i>Allodia (Brachycampta) alternans</i> (Zetterstedt, 1838)
Mycetophilidae	Mycetophilinae	<i>Allodia (Brachycampta) barbata</i> (Lundstrom, 1909)
Mycetophilidae	Mycetophilinae	<i>Allodia (Brachycampta) foliifera</i> (Strobl, 1910)
Mycetophilidae	Mycetophilinae	<i>Allodia (Brachycampta) grata</i> (Meigen, 1830)
Mycetophilidae	Mycetophilinae	<i>Allodia (Brachycampta) pistillata</i> (Lundstrom, 1911)
Mycetophilidae	Mycetophilinae	<i>Allodiopsis domestica</i> (Meigen, 1830)
Mycetophilidae	Mycetophilinae	<i>Allodiopsis korolevi</i> Zaitzev, 1982
Mycetophilidae	Mycetophilinae	<i>Allodiopsis rustica</i> (Edwards, 1941)
Mycetophilidae	Mycetophilinae	* <i>Anatella ciliata</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Anatella turi</i> Dziedzicki, 1923
Mycetophilidae	Mycetophilinae	<i>Brevicornu canescens</i> (Zetterstedt, 1852)
Mycetophilidae	Mycetophilinae	<i>Brevicornu fissicauda</i> (Lundstrom, 1911)
Mycetophilidae	Mycetophilinae	<i>Brevicornu griseicollis</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Brevicornu kingi</i> (Edwards, 1925)
Mycetophilidae	Mycetophilinae	<i>Brevicornu proximum</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Brevicornu sericoma</i> (Meigen, 1830)
Mycetophilidae	Mycetophilinae	<i>Brevicornu spathulatum</i> (Lundstrom, 1911)
Mycetophilidae	Mycetophilinae	<i>Brevicornu verralli</i> (Edwards, 1925)
Mycetophilidae	Mycetophilinae	<i>Cordyla brevicornis</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Cordyla crassicornis</i> Meigen, 1818
Mycetophilidae	Mycetophilinae	<i>Cordyla fasciata</i> Meigen, 1830
Mycetophilidae	Mycetophilinae	<i>Cordyla fissa</i> Edwards, 1925
Mycetophilidae	Mycetophilinae	<i>Cordyla fusca</i> Meigen, 1804
Mycetophilidae	Mycetophilinae	<i>Cordyla insons</i> Lastovka & Matile, 1974
Mycetophilidae	Mycetophilinae	<i>Cordyla murina</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Cordyla pusilla</i> Edwards, 1925
Mycetophilidae	Mycetophilinae	<i>Cordyla semiflava</i> (Staeger, 1840)

Continued

Table 2. Continued.

Family	Subfamily	Species
Mycetophilidae	Mycetophilinae	<i>Dynatosoma cochleare</i> Strobl, 1895
Mycetophilidae	Mycetophilinae	<i>Dynatosoma fuscicorne</i> (Meigen, 1818)
Mycetophilidae	Mycetophilinae	<i>Dynatosoma majus</i> Landrock, 1912
Mycetophilidae	Mycetophilinae	<i>Dynatosoma nigromaculatum</i> Lundstrom, 1913
Mycetophilidae	Mycetophilinae	<i>Dynatosoma nobile</i> Loew, 1873
Mycetophilidae	Mycetophilinae	<i>Dynatosoma reciprocum</i> (Walker, 1848)
Mycetophilidae	Mycetophilinae	<i>Epicypta aterrima</i> (Zetterstedt, 1852)
Mycetophilidae	Mycetophilinae	<i>Epicypta torquata</i> Matile, 1977
Mycetophilidae	Mycetophilinae	<i>Exechia bicincta</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Exechia confinis</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Exechia contaminata</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Exechia dizona</i> Edwards, 1924
Mycetophilidae	Mycetophilinae	<i>Exechia dorsalis</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Exechia fusca</i> (Meigen, 1804)
Mycetophilidae	Mycetophilinae	<i>Exechia pseudocincta</i> Strobl, 1910
Mycetophilidae	Mycetophilinae	<i>Exechia separata</i> Lundstrom, 1912
Mycetophilidae	Mycetophilinae	<i>Exechia seriata</i> (Meigen, 1830)
Mycetophilidae	Mycetophilinae	<i>Exechia spinuligera</i> Lundstrom, 1912
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) dumitrescae</i> Burghel-Balacesco, 1972
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) furcata</i> (Lundstrom, 1911)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) indecisa</i> (Walker, 1856)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) ingraca</i> (Stackelberg, 1948)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) intersecta</i> (Meigen, 1818)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) januarii</i> (Lundstrom, 1913)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) jenkinsoni</i> (Edwards, 1925)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) lackschewitziana</i> (Stackelberg, 1948)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) landrocki</i> (Lundstrom, 1912)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) magnicauda</i> (Lundstrom, 1911)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) oltenica</i> (Burghel-Balacesco, 1965)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) pulchella</i> (Winnertz, 1864)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) subulata</i> (Winnertz, 1864)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) trisetula</i> Tolle, 1955
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) unguiculata</i> (Lundstrom, 1911)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) vizzavonensis</i> (Edwards, 1929)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Xenexechia) crucigera</i> (Lundstrom, 1909)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Xenexechia) davatchii</i> (Matile, 1969)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Xenexechia) palettata</i> (Burghel-Balacesco, 1965)
Mycetophilidae	Mycetophilinae	<i>Macrobrachius kowarzii</i> Dziedzicki, 1889
Mycetophilidae	Mycetophilinae	<i>Mycetophila abbreviata</i> Landrock, 1914
Mycetophilidae	Mycetophilinae	<i>Mycetophila alea</i> Laffoon, 1965
Mycetophilidae	Mycetophilinae	<i>Mycetophila bialorusica</i> Dziedzicki, 1884
Mycetophilidae	Mycetophilinae	<i>Mycetophila blanda</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Mycetophila confluens</i> Dziedzicki, 1884
Mycetophilidae	Mycetophilinae	<i>Mycetophila curviseta</i> Lundstrom, 1911
Mycetophilidae	Mycetophilinae	<i>Mycetophila czizeki</i> Landrock, 1911
Mycetophilidae	Mycetophilinae	<i>*Mycetophila deflexa</i> Chandler, 2001
Mycetophilidae	Mycetophilinae	<i>Mycetophila dentata</i> Lundstrom, 1913
Mycetophilidae	Mycetophilinae	<i>Mycetophila edwardsi</i> Lundstrom, 1913
Mycetophilidae	Mycetophilinae	<i>Mycetophila fraterna</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Mycetophila fungorum</i> (De Geer, 1776)
Mycetophilidae	Mycetophilinae	<i>Mycetophila gibbula</i> Edwards, 1925
Mycetophilidae	Mycetophilinae	<i>Mycetophila hetschkoi</i> Landrock, 1918
Mycetophilidae	Mycetophilinae	<i>Mycetophila idonea</i> Lastovka, 1972
Mycetophilidae	Mycetophilinae	<i>Mycetophila laeta</i> Walker, 1848
Mycetophilidae	Mycetophilinae	<i>Mycetophila lamellata</i> Lundstrom, 1911
Mycetophilidae	Mycetophilinae	<i>*Mycetophila lastovkai</i> Caspers, 1984
Mycetophilidae	Mycetophilinae	<i>Mycetophila magnicauda</i> Strobl, 1895
Mycetophilidae	Mycetophilinae	<i>Mycetophila marginata</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Mycetophila mikii</i> Dziedzicki, 1884
Mycetophilidae	Mycetophilinae	<i>Mycetophila occultans</i> Lundstrom, 1913
Mycetophilidae	Mycetophilinae	<i>Mycetophila ocellus</i> Walker, 1848

Continued

Table 2. Continued.

Family	Subfamily	Species
Mycetophilidae	Mycetophilinae	<i>Mycetophila ornata</i> Stephens, 1829
Mycetophilidae	Mycetophilinae	<i>Mycetophila perpallida</i> Chandler, 1993
Mycetophilidae	Mycetophilinae	<i>Mycetophila pumila</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Mycetophila rudis</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Mycetophila ruficollis</i> Meigen, 1818
Mycetophilidae	Mycetophilinae	<i>Mycetophila scotica</i> Edwards, 1941
Mycetophilidae	Mycetophilinae	<i>Mycetophila sigillata</i> Dziedzicki, 1884
Mycetophilidae	Mycetophilinae	<i>Mycetophila signatoides</i> Dziedzicki, 1884
Mycetophilidae	Mycetophilinae	<i>Mycetophila spectabilis</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Mycetophila stolidus</i> Walker, 1856
Mycetophilidae	Mycetophilinae	<i>Mycetophila strigata</i> Staeger, 1840
Mycetophilidae	Mycetophilinae	<i>Mycetophila strigatoides</i> (Landrock, 1927)
Mycetophilidae	Mycetophilinae	<i>Mycetophila strobili</i> Lastovka, 1972
Mycetophilidae	Mycetophilinae	* <i>Mycetophila stylata</i> (Dziedzicki, 1884)
Mycetophilidae	Mycetophilinae	<i>Mycetophila trinotata</i> Staeger, 1840
Mycetophilidae	Mycetophilinae	<i>Mycetophila tuberosa</i> Lundstrom, 1911
Mycetophilidae	Mycetophilinae	<i>Mycetophila unguiculata</i> Lundstrom, 1913
Mycetophilidae	Mycetophilinae	<i>Mycetophila unicolor</i> Stannius, 1831
Mycetophilidae	Mycetophilinae	<i>Mycetophila v-nigrum</i> Lundstrom, 1913
Mycetophilidae	Mycetophilinae	<i>Mycetophila zetterstedtii</i> Lundstrom, 1906
Mycetophilidae	Mycetophilinae	<i>Myrosia maculosa</i> (Meigen, 1818)
Mycetophilidae	Mycetophilinae	<i>Notolopha cristata</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Phronia biarcuata</i> (Santos Abreu, 1920)
Mycetophilidae	Mycetophilinae	<i>Phronia cinerascens</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Phronia flavipes</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Phronia forcipata</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Phronia forcipula</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Phronia humeralis</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Phronia maculata</i> Dziedzicki, 1889
Mycetophilidae	Mycetophilinae	* <i>Phronia nigricornis</i> (Zetterstedt, 1852)
Mycetophilidae	Mycetophilinae	<i>Phronia nitidiventris</i> (Wulp, 1858)
Mycetophilidae	Mycetophilinae	<i>Phronia obtusa</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Phronia petulans</i> Dziedzicki, 1889
Mycetophilidae	Mycetophilinae	<i>Phronia tenuis</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Phronia triangularis</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Platurocypta punctum</i> (Stannius, 1831)
Mycetophilidae	Mycetophilinae	<i>Platurocypta testata</i> (Edwards, 1925)
Mycetophilidae	Mycetophilinae	<i>Pseudexechia parallela</i> (Edwards, 1925)
Mycetophilidae	Mycetophilinae	<i>Pseudexechia trisignata</i> (Edwards, 1913)
Mycetophilidae	Mycetophilinae	<i>Pseudexechia trivittata</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Pseudobrachypeza helvetica</i> (Walker, 1856)
Mycetophilidae	Mycetophilinae	<i>Rymosia affinis</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Rymosia fasciata</i> (Meigen, 1804)
Mycetophilidae	Mycetophilinae	<i>Rymosia lundstroemi</i> Dziedzicki, 1910
Mycetophilidae	Mycetophilinae	<i>Rymosia spinipes</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Rymosia tolleti</i> Burghel-Balacesco, 1965
Mycetophilidae	Mycetophilinae	<i>Sceptonia concolor</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Stigmatomeria crassicornis</i> (Stannius, 1831)
Mycetophilidae	Mycetophilinae	* <i>Synplasta gracilis</i> (Winnertz, 1864)
Mycetophilidae	Mycetophilinae	* <i>Synplasta rufilata</i> (Edwards, 1941)
Mycetophilidae	Mycetophilinae	<i>Tarnania dziedzickii</i> (Edwards, 1941)
Mycetophilidae	Mycetophilinae	<i>Tarnania fenestralis</i> (Meigen, 1818)
Mycetophilidae	Mycetophilinae	<i>Tarnania tarnanii</i> (Dziedzicki, 1910)
Mycetophilidae	Mycetophilinae	<i>Trichonta aberrans</i> Lundstrom, 1911
Mycetophilidae	Mycetophilinae	<i>Trichonta brevicauda</i> Lundstrom, 1906
Mycetophilidae	Mycetophilinae	<i>Trichonta clavigera</i> Lundstrom, 1913
Mycetophilidae	Mycetophilinae	* <i>Trichonta comis</i> Gagne, 1981
Mycetophilidae	Mycetophilinae	<i>Trichonta concinna</i> Gagne, 1981
Mycetophilidae	Mycetophilinae	<i>Trichonta conjungens</i> Lundstrom, 1909
Mycetophilidae	Mycetophilinae	<i>Trichonta excisa</i> Lundstrom, 1916
Mycetophilidae	Mycetophilinae	<i>Trichonta falcata</i> Lundstrom, 1911

Continued

Table 2. Continued.

Family	Subfamily	Species
Mycetophilidae	Mycetophilinae	<i>Trichonta flavicauda</i> Lundstrom, 1914
Mycetophilidae	Mycetophilinae	<i>Trichonta foeda</i> Loew, 1869
Mycetophilidae	Mycetophilinae	<i>Trichonta fusca</i> Landrock, 1918
Mycetophilidae	Mycetophilinae	<i>Trichonta girschneri</i> Landrock, 1912
Mycetophilidae	Mycetophilinae	<i>Trichonta hamata</i> Mik, 1880
Mycetophilidae	Mycetophilinae	<i>Trichonta hungarica</i> Landrock, 1925
Mycetophilidae	Mycetophilinae	<i>Trichonta submaculata</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Trichonta trivittata</i> Lundstrom, 1916
Mycetophilidae	Mycetophilinae	<i>Trichonta venosa</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Trichonta vitta</i> (Meigen, 1830)
Mycetophilidae	Mycetophilinae	* <i>Trichonta vulcani</i> (Dziedzicki, 1889)
Mycetophilidae	Mycetophilinae	<i>Trichonta vulgaris</i> Loew, 1869
Mycetophilidae	Mycetophilinae	<i>Zygomyia humeralis</i> (Wiedemann, 1817)
Mycetophilidae	Mycetophilinae	<i>Zygomyia notata</i> (Stannius, 1831)
Mycetophilidae	Mycetophilinae	<i>Zygomyia pictipennis</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Zygomyia pseudohumeralis</i> Caspers, 1980
Mycetophilidae	Mycomyinae	<i>Mycomya (Cesamya) pectinifera</i> Edwards, 1924
Mycetophilidae	Mycomyinae	<i>Mycomya (Cymomya) circumdata</i> (Staeger, 1840)
Mycetophilidae	Mycomyinae	* <i>Mycomya (Mycomya) alpina</i> Matile, 1972
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) bicolor</i> (Dziedzicki, 1885)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) brunnea</i> (Dziedzicki, 1885)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) cinerascens</i> (Macquart, 1826)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) danielae</i> Matile, 1972
Mycetophilidae	Mycomyinae	* <i>Mycomya (Mycomya) egregia</i> (Dziedzicki, 1885)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) griseovittata</i> (Zetterstedt, 1852)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) levis</i> (Dziedzicki, 1885)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) marginata</i> (Meigen, 1818)
Mycetophilidae	Mycomyinae	* <i>Mycomya (Mycomya) neohyalinata</i> Vaisanen, 1984
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) occultans</i> (Winnertz, 1864)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) parva</i> (Dziedzicki, 1885)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) prominens</i> (Lundstrom, 1913)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) ruficollis</i> (Zetterstedt, 1852)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) tenuis</i> (Walker, 1856)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) trivittata</i> Zetterstedt, 1838
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) tumida</i> (Winnertz, 1864)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) vittiventris</i> (Zetterstedt, 1852)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) wankowiczii</i> (Dziedzicki, 1885)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) winnertzi</i> (Dziedzicki, 1885)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomyopsis) maura</i> (Walker, 1856)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomyopsis) trilineata</i> (Zetterstedt, 1838)
Mycetophilidae	Mycomyinae	<i>Neoempheria pictipennis</i> (Haliday, 1833)
Mycetophilidae	Mycomyinae	<i>Neoempheria winnertzi</i> Edwards, 1913
Mycetophilidae	Polylepta	<i>Polylepta guttiventris</i> (Zetterstedt, 1852)
Mycetophilidae	Sciophilinae	<i>Acnemis nitidicollis</i> (Meigen, 1818)
Mycetophilidae	Sciophilinae	<i>Anaclileia dispar</i> (Winnertz, 1864)
Mycetophilidae	Sciophilinae	<i>Leptomorphus quadrimaculatus</i> (Matsumura, 1916)
Mycetophilidae	Sciophilinae	<i>Monoclona furcata</i> Johannsen, 1910
Mycetophilidae	Sciophilinae	<i>Monoclona rufilatera</i> (Walker, 1836)
Mycetophilidae	Sciophilinae	<i>Neuratelia minor</i> (Lundstrom, 1912)
Mycetophilidae	Sciophilinae	<i>Neuratelia nemoralis</i> (Meigen, 1818)
Mycetophilidae	Sciophilinae	* <i>Phthinia humilis</i> Winnertz, 1864
Mycetophilidae	Sciophilinae	<i>Sciophila tenuis</i> (Winnertz, 1864)
Mycetophilidae	Sciophilinae	<i>Sciophila varia</i> (Winnertz, 1864)

Table 3. Checklist of fungus gnats (Ditomyiidae, Diadocidiidae, Keroplatidae, Bolitophilidae, Mycetophilidae) of Montenegro and Serbia, based Bechev (1997) and Chandler (2013). New records for Montenegro are listed with one asterisk (*) and new for Serbia with two asterisks (**).

Family	Subfamily	Species
Bolitophilidae		* <i>Bolitophila (Bolitophila) austriaca</i> (Mayer, 1950)
Bolitophilidae		<i>Bolitophila (Bolitophila) cinerea</i> Meigen, 1818
Bolitophilidae		<i>Bolitophila (Bolitophila) cooremani</i> (Tollet, 1955)
Bolitophilidae		<i>Bolitophila (Bolitophila) lengersdorfi</i> (Tollet, 1955)
Bolitophilidae		<i>Bolitophila (Bolitophila) tenella</i> Winnertz, 1864
Bolitophilidae		* <i>Bolitophila (Bolitophila) saundersi</i> (Curtis, 1836)
Bolitophilidae		<i>Bolitophila (Cliopisa) bimaculata</i> Zetterstedt, 1838
Bolitophilidae		<i>Bolitophila (Cliopisa) hybrida</i> (Meigen, 1804)
Bolitophilidae		<i>Bolitophila (Cliopisa) maculipennis</i> Walker, 1835
Bolitophilidae		* <i>Bolitophila (Cliopisa) melanoleuci</i> Polevoi, 1996
Bolitophilidae		<i>Bolitophila (Cliopisa) modesta</i> Lackschewitz, 1937
Bolitophilidae		<i>Bolitophila (Cliopisa) nigrolineata</i> Landrock, 1912
Diadocidiidae		<i>Diadocidia (Diadocidia) ferruginosa</i> (Meigen, 1830)
Diadocidiidae		<i>Diadocidia (Diadocidia) spinosula</i> Tollet, 1948
Ditomyiidae		<i>Ditomyia fasciata</i> (Meigen, 1818)
Keroplatidae	Keroplatinae	<i>Antlemon (Antlemonopsis) brevimanum</i> (Loew, 1871)
Keroplatidae	Keroplatinae	** <i>Cerotelion striatum</i> (Gmelin, 1790)
Keroplatidae	Keroplatinae	<i>Isoneuromyia pseudochracea</i> (Landrock, 1925)
Keroplatidae	Keroplatinae	<i>Macrorrhyncha brevirostris</i> (Lundstrom, 1911)
Keroplatidae	Keroplatinae	<i>Orfelia ochracea</i> (Meigen, 1918)
Keroplatidae	Macrocerinae	<i>Macrocera centralis</i> Meigen, 1818
Keroplatidae	Macrocerinae	<i>Macrocera fasciata</i> Meigen, 1804
Keroplatidae	Macrocerinae	<i>Macrocera inversa</i> Loew, 1869
Keroplatidae	Macrocerinae	<i>Macrocera lutea</i> Meigen, 1804
Keroplatidae	Macrocerinae	<i>Macrocera obscura</i> Winnertz, 1864
Keroplatidae	Macrocerinae	<i>Macrocera phalerata</i> Meigen, 1818
Keroplatidae	Macrocerinae	<i>Macrocera vittata</i> Meigen, 1830
Mycetophilidae	Gnoristinae	<i>Apolephthisa subincana</i> (Curtis, 1837)
Mycetophilidae	Gnoristinae	<i>Boletina basalis</i> (Meigen, 1818)
Mycetophilidae	Gnoristinae	<i>Boletina cincticornis</i> (Walker, 1848)
Mycetophilidae	Gnoristinae	<i>Boletina gripha</i> Dziedzicki, 1885
Mycetophilidae	Gnoristinae	<i>Boletina griphoides</i> Edwards, 1925
Mycetophilidae	Gnoristinae	<i>Boletina lundstroemi</i> Landrock, 1912
Mycetophilidae	Gnoristinae	<i>Boletina sciarina</i> Staeger, 1840
Mycetophilidae	Gnoristinae	<i>Coelophthinia thoracica</i> (Winnertz, 1864)
Mycetophilidae	Gnoristinae	* <i>Coelosia fusca</i> Bezzi, 1892
Mycetophilidae	Gnoristinae	<i>Palaeodocosia vittata</i> (Coquillett, 1901)
Mycetophilidae	Gnoristinae	<i>Speolepta leptogaster</i> (Winnertz, 1864)
Mycetophilidae	Gnoristinae	<i>Synapha vitripennis</i> (Meigen, 1818)
Mycetophilidae	Leiinae	* <i>Docosia gilvipes</i> (Walker, 1856)
Mycetophilidae	Leiinae	<i>Docosia lastovkai</i> Chandler, 1994
Mycetophilidae	Leiinae	<i>Ectrepesthoneura colyeri</i> Chandler, 1980
Mycetophilidae	Leiinae	<i>Leia bimaculata</i> (Meigen, 1804)
Mycetophilidae	Leiinae	<i>Tetragoneura ambigua</i> (Grzegorzek, 1885)
Mycetophilidae	Leiinae	<i>Tetragoneura sylvatica</i> (Curtis, 1837)
Mycetophilidae	Mycetophilinae	<i>Allodia (Allodia) lugens</i> (Wiedemann, 1817)
Mycetophilidae	Mycetophilinae	<i>Allodia (Allodia) ornatocollis</i> (Meigen, 1818)
Mycetophilidae	Mycetophilinae	<i>Allodia (Brachycampta) alternans</i> (Zetterstedt, 1838)
Mycetophilidae	Mycetophilinae	<i>Allodia (Brachycampta) foliifera</i> (Strobl, 1910)
Mycetophilidae	Mycetophilinae	<i>Allodia (Brachycampta) grata</i> (Meigen, 1830)
Mycetophilidae	Mycetophilinae	<i>Allodia (Brachycampta) pistillata</i> (Lundstrom, 1911)
Mycetophilidae	Mycetophilinae	<i>Allodia (Brachycampta) retracta</i> Plassmann, 1977
Mycetophilidae	Mycetophilinae	<i>Allodia (Brachycampta) triangularis</i> (Strobl, 1895)
Mycetophilidae	Mycetophilinae	<i>Allodiopsis domestica</i> (Meigen, 1830)
Mycetophilidae	Mycetophilinae	<i>Allodiopsis rustica</i> (Edwards, 1941)
Mycetophilidae	Mycetophilinae	<i>Anatella concava</i> Plassmann, 1990
Mycetophilidae	Mycetophilinae	<i>Anatella lenis</i> Dziedzicki, 1923
Mycetophilidae	Mycetophilinae	<i>Anatella minuta</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Anatella novata</i> Dziedzicki, 1923

Continued

Table 3. Continued.

Family	Subfamily	Species
Mycetophilidae	Mycetophilinae	<i>Brachypeza (Brachypeza) armata</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Brevicornu auriculatum</i> (Edwards, 1925)
Mycetophilidae	Mycetophilinae	<i>Brevicornu fissicauda</i> (Lundstrom, 1911)
Mycetophilidae	Mycetophilinae	<i>Brevicornu fuscipenne</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Brevicornu griseicollis</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Brevicornu intermedium</i> Santos Abreu, 1920
Mycetophilidae	Mycetophilinae	<i>Brevicornu proximum</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Brevicornu sericoma</i> (Meigen, 1830)
Mycetophilidae	Mycetophilinae	<i>Brevicornu subfissicauda</i> Zaitzev, 1985
Mycetophilidae	Mycetophilinae	<i>Brevicornu verralli</i> (Edwards, 1925)
Mycetophilidae	Mycetophilinae	<i>Cordyla brevicornis</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Cordyla crassicornis</i> Meigen, 1818
Mycetophilidae	Mycetophilinae	<i>Cordyla fasciata</i> Meigen, 1830
Mycetophilidae	Mycetophilinae	<i>Cordyla fissa</i> Edwards, 1925
Mycetophilidae	Mycetophilinae	<i>Cordyla flaviceps</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Cordyla fusca</i> Meigen, 1804
Mycetophilidae	Mycetophilinae	<i>Cordyla murina</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Cordyla nitidula</i> Edwards, 1925
Mycetophilidae	Mycetophilinae	<i>Dynatosoma cochleare</i> Strobl, 1895
Mycetophilidae	Mycetophilinae	<i>Dynatosoma majus</i> Landrock, 1912
Mycetophilidae	Mycetophilinae	<i>Exechia bicincta</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Exechia dorsalis</i> (Staeger, 1840)
Mycetophilidae	Mycetophilinae	<i>Exechia festiva</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Exechia fusca</i> (Meigen, 1804)
Mycetophilidae	Mycetophilinae	<i>Exechia nitidicollis</i> Lundstrom, 1913
Mycetophilidae	Mycetophilinae	<i>Exechia seriata</i> (Meigen, 1830)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) clypeata</i> (Lundstrom, 1911)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) intersecta</i> (Meigen, 1818)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) magnicauda</i> (Lundstrom, 1911)
Mycetophilidae	Mycetophilinae	<i>Exechiopsis (Exechiopsis) subulata</i> (Winnertz, 1864)
Mycetophilidae	Mycetophilinae	<i>*Exechiopsis (Exechiopsis) unguiculata</i> (Lundstrom, 1911)
Mycetophilidae	Mycetophilinae	<i>Mycetophila abiecta</i> (Lastovka, 1963)
Mycetophilidae	Mycetophilinae	<i>Mycetophila adumbrata</i> Mik, 1884
Mycetophilidae	Mycetophilinae	<i>Mycetophila alea</i> Laffoon, 1965
Mycetophilidae	Mycetophilinae	<i>Mycetophila attonsa</i> (Laffoon, 1957)
Mycetophilidae	Mycetophilinae	<i>Mycetophila bialorussica</i> Dziedzicki, 1884
Mycetophilidae	Mycetophilinae	<i>Mycetophila bohémica</i> (Lastovka, 1963)
Mycetophilidae	Mycetophilinae	<i>Mycetophila curviseta</i> Lundstrom, 1911
Mycetophilidae	Mycetophilinae	<i>Mycetophila czizeki</i> Landrock, 1911
Mycetophilidae	Mycetophilinae	<i>Mycetophila edwardsi</i> Lundstrom, 1913
Mycetophilidae	Mycetophilinae	<i>Mycetophila fraterna</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Mycetophila fungorum</i> (De Geer, 1776)
Mycetophilidae	Mycetophilinae	<i>Mycetophila gibbula</i> Edwards, 1925
Mycetophilidae	Mycetophilinae	<i>Mycetophila hetschkoi</i> Landrock, 1918
Mycetophilidae	Mycetophilinae	<i>Mycetophila ichneumonea</i> Say, 1823
Mycetophilidae	Mycetophilinae	<i>Mycetophila lamellata</i> Lundstrom, 1911
Mycetophilidae	Mycetophilinae	<i>Mycetophila luctuosa</i> Meigen, 1830
Mycetophilidae	Mycetophilinae	<i>*Mycetophila marginata</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Mycetophila occultans</i> Lundstrom, 1913
Mycetophilidae	Mycetophilinae	<i>Mycetophila ocellus</i> Walker, 1848
Mycetophilidae	Mycetophilinae	<i>Mycetophila perpallida</i> Chandler, 1993
Mycetophilidae	Mycetophilinae	<i>Mycetophila pictula</i> Meigen, 1830
Mycetophilidae	Mycetophilinae	<i>Mycetophila pumila</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Mycetophila ruficollis</i> Meigen, 1818
Mycetophilidae	Mycetophilinae	<i>Mycetophila scotica</i> Edwards, 1941
Mycetophilidae	Mycetophilinae	<i>Mycetophila sigillata</i> Dziedzicki, 1884
Mycetophilidae	Mycetophilinae	<i>Mycetophila signata</i> Meigen, 1830
Mycetophilidae	Mycetophilinae	<i>Mycetophila signatoides</i> Dziedzicki, 1884
Mycetophilidae	Mycetophilinae	<i>Mycetophila spectabilis</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Mycetophila stolidus</i> Walker, 1856
Mycetophilidae	Mycetophilinae	<i>Mycetophila strigatoides</i> (Landrock, 1927)

Continued

Table 3. Continued.

Family	Subfamily	Species
Mycetophilidae	Mycetophilinae	<i>Mycetophila stylata</i> (Dziedzicki, 1884)
Mycetophilidae	Mycetophilinae	<i>Mycetophila stylatiformis</i> Landrock, 1925
Mycetophilidae	Mycetophilinae	<i>Mycetophila subsigillata</i> Zaitzev, 1999
Mycetophilidae	Mycetophilinae	<i>Mycetophila sumavica</i> (Lastovka, 1963)
Mycetophilidae	Mycetophilinae	<i>Mycetophila uninotata</i> Zetterstedt, 1852
Mycetophilidae	Mycetophilinae	<i>Phronia biarcuata</i> (Santos Abreu, 1920)
Mycetophilidae	Mycetophilinae	<i>Phronia conformis</i> (Walker, 1856)
Mycetophilidae	Mycetophilinae	<i>Phronia electa</i> Dziedzicki, 1889
Mycetophilidae	Mycetophilinae	* <i>Phronia forcipula</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Phronia nigricornis</i> (Zetterstedt, 1852)
Mycetophilidae	Mycetophilinae	<i>Phronia nitidiventris</i> (Wulp, 1858)
Mycetophilidae	Mycetophilinae	<i>Phronia notata</i> Dziedzicki, 1889
Mycetophilidae	Mycetophilinae	<i>Phronia signata</i> Winnertz, 1863
Mycetophilidae	Mycetophilinae	<i>Phronia strenua</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Phronia tenuis</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Phronia triangularis</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Pseudexechia trisignata</i> (Edwards, 1913)
Mycetophilidae	Mycetophilinae	<i>Pseudobrachypeza helvetica</i> (Walker, 1856)
Mycetophilidae	Mycetophilinae	<i>Rymosia acta</i> Dziedzicki, 1910
Mycetophilidae	Mycetophilinae	<i>Rymosia affinis</i> Winnertz, 1864
Mycetophilidae	Mycetophilinae	<i>Rymosia fasciata</i> (Meigen, 1804)
Mycetophilidae	Mycetophilinae	* <i>Rymosia lundstroemi</i> Dziedzicki, 1910
Mycetophilidae	Mycetophilinae	<i>Sceptonia hamata</i> Sevcik, 2004
Mycetophilidae	Mycetophilinae	<i>Sceptonia membranea</i> Edwards, 1925
Mycetophilidae	Mycetophilinae	<i>Sceptonia nigra</i> (Meigen, 1804)
Mycetophilidae	Mycetophilinae	<i>Stigmatomeria crassicornis</i> (Stannius, 1831)
Mycetophilidae	Mycetophilinae	<i>Synplasta gracilis</i> (Winnertz, 1864)
Mycetophilidae	Mycetophilinae	<i>Tarnania dziedzickii</i> (Edwards, 1941)
Mycetophilidae	Mycetophilinae	<i>Tarnania fenestralis</i> (Meigen, 1818)
Mycetophilidae	Mycetophilinae	<i>Trichonta comica</i> Gagne, 1981
Mycetophilidae	Mycetophilinae	<i>Trichonta falcata</i> Lundstrom, 1911
Mycetophilidae	Mycetophilinae	<i>Trichonta girschneri</i> Landrock, 1912
Mycetophilidae	Mycetophilinae	<i>Trichonta subfusca</i> Lundstrom, 1909
Mycetophilidae	Mycetophilinae	<i>Trichonta vitta</i> (Meigen, 1830)
Mycetophilidae	Mycetophilinae	<i>Trichonta vulgaris</i> Loew, 1869
Mycetophilidae	Mycetophilinae	<i>Zygomyia humeralis</i> (Wiedemann, 1817)
Mycetophilidae	Mycetophilinae	<i>Zygomyia notata</i> (Stannius, 1831)
Mycetophilidae	Mycetophilinae	<i>Zygomyia pseudohumeralis</i> Caspers, 1980
Mycetophilidae	Mycetophilinae	<i>Zygomyia valida</i> Winnertz, 1864
Mycetophilidae	Mycomyinae	<i>Mycomya (Cymomya) circumdata</i> (Staeger, 1840)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) cinerascens</i> (Macquart, 1826)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) danielae</i> Matile, 1972
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) flavicollis</i> (Zetterstedt, 1852)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) griseovittata</i> (Zetterstedt, 1852)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) marginata</i> (Meigen, 1818)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) prominens</i> (Lundstrom, 1913)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) sigma</i> Johannsen, 1910
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) tenuis</i> (Walker, 1856)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) wankowiczii</i> (Dziedzicki, 1885)
Mycetophilidae	Mycomyinae	<i>Mycomya (Mycomya) winnertzi</i> (Dziedzicki, 1885)
Mycetophilidae	Mycomyinae	* <i>Mycomya (Mycomyopsis) trilineata</i> (Zetterstedt, 1838)
Mycetophilidae	Mycomyinae	<i>Mycomya (Neomycomya) fimbriata</i> (Meigen, 1818)
Mycetophilidae	Mycomyinae	<i>Neoempheria lineola</i> (Meigen, 1818)
Mycetophilidae	Mycomyinae	<i>Neoempheria pictipennis</i> (Haliday, 1833)
Mycetophilidae	Mycomyinae	<i>Neoempheria proxima</i> (Winnertz, 1864)
Mycetophilidae	Sciophilinae	<i>Monoclona rufilatera</i> (Walker, 1836)
Mycetophilidae	Sciophilinae	<i>Sciophila baltica</i> Zaitzev, 1982
Mycetophilidae	Sciophilinae	<i>Sciophila hirta</i> Meigen, 1818
Mycetophilidae	Sciophilinae	<i>Sciophila thoracica</i> Staeger, 1840

Acknowledgements

We would like to express our sincere thanks to Edina Török and Lujza Keresztes for collecting material. This study was financially supported by a grant of the Romanian Ministry of National Education, CNCS-UEFISCDI-nr. PN-II-ID-PCE-2012-4-0595. The first author received financial support with scholarships from the Collegium Talentum, Hungary. Pekka Vilkkamaa (MZHF) kindly arranged the loan of the holotype of *T. comis*. English text was improved by Pete Boardman (Shrewsbury). Constructive comments by 3 anonymous referees and Eliana Buenaventura (Smithsonian Institution) improved the manuscript.

Authors' Contributions

L-PK collected the samples and took photos; JS identified the specimens; LPK and JS wrote the text.

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